

1 **Australian lizards are outstanding models for reproductive biology research**

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## 34 **Abstract**

35 Australian lizards are a diverse group of animals distributed across the continent and  
36 inhabiting a wide range of environments. Together, they exhibit a remarkable diversity of  
37 reproductive morphologies, physiologies, and behaviours that is broadly representative of  
38 vertebrates more generally. Importantly, many of these reproductive traits have evolved  
39 independently in multiple Australian lizard groups, and these groups are closely related to  
40 taxa that exhibit ancestral forms of these traits. Australian lizards are thus outstanding model  
41 organisms for testing hypotheses about how reproductive traits function and evolve and they  
42 provide an important basis of comparison with other animals that exhibit similar traits. Here,  
43 we highlight how research on Australian lizard reproduction has contributed to answering  
44 broader evolutionary and ecological questions that apply to all animals. We review  
45 reproductive traits exhibited by Australian lizards that are important areas of current research.  
46 We highlight important questions that emerge from an understanding of this body of research  
47 when considered as a whole. Finally, we identify additional research questions that Australian  
48 lizards are well-suited for reproductive biologists to address.

## 49 **Introduction**

50 Australia is home to about 13 % of the world's lizard biodiversity (~850 of ~7,000 species;  
51 Uetz 2010), despite containing only 5 % of the world's land area. Australian lizards are  
52 taxonomically distributed across seven families: Carphodactylidae, Diplodactylidae,  
53 Gekkonidae, Pygopodidae (all from the infraorder Gekkota), Scincidae, Agamidae, and  
54 Varanidae (Cogger 2018). Of these, the Scincidae are the most diverse Australian lineage,  
55 with ~460 species (Wilson and Swan 2021). The Carphodactylide are endemic to Australia,  
56 and Australia is home to large proportions of the world's Diplodactylidae (~65 %),  
57 Pygopodidae (~97 %), Scincidae (~30 %), and Varanidae (~50 %). These species numbers

58 and proportions are not exact, however, because the taxonomy of Australian lizards is still  
59 developing. Together, Australian lizards are distributed across the entire continent, and  
60 experience habitats ranging from cool temperate forests and alpine meadows to hot deserts  
61 and tropical rainforests (Figure 1; Cogger 2018). Most of the lizard species in Australia exist  
62 in only a few of these habitats, but some, like the pygopodid *Lialis burtonis* and the scincid  
63 *Tiliqua scincoides*, thrive across a range of habitat types. Australia's diverse environments,  
64 and the abilities of different lizard taxa to specialise or generalise to survive within them, are  
65 likely major reasons for Australia's high lizard biodiversity (Pianka 1969; Skeels *et al.* 2020).  
66  
67 Australian lizard biodiversity, and its distribution across such variable environments,  
68 provides important opportunities for testing hypotheses about how traits evolve and function.  
69 In particular, Australian lizards exhibit multiple evolutionary origins of key innovations  
70 related to reproduction, including social behaviours (Gardner *et al.* 2016), signalling and  
71 reproductive tactics (Dong CM 2021; Stuart-Fox *et al.* 2021), viviparity (live birth;  
72 Blackburn 2015), and transitions in sex determination (Sarre *et al.* 2004). The multiple  
73 origins of these traits provide natural replication for robust tests of key evolutionary  
74 hypotheses (Garland and Adolph 1994): do the physiological and genetic functions of these  
75 traits evolve in the same ways? How do their functions change in species that are distributed  
76 across variable environments (Figure 1)? The wide, overlapping distributions of some  
77 Australian lizard taxa (Cogger 2018) exhibiting variation in these traits provides a framework  
78 for answering these questions across species, and sometimes even within single species.  
79 Furthermore, many of these traits are shared with other animals, so the results gained from  
80 lizard research provide a basis for comparison with those other taxa (Tinkle *et al.* 1970).

81

82 In addition to their utility for answering scientific questions, lizards are tractable research  
83 models. Many species are an ideal size to study using both captive and field-based  
84 approaches. They are big enough to carry dataloggers, yet small enough to house in captivity  
85 with relative ease and low costs (McDiarmid *et al.* 2012). Several Australian skinks and  
86 dragons exhibit high population densities, which enables the sample sizes required for robust  
87 ecological analyses. For these reasons, lizards around the world have been important model  
88 organisms for research in ecology, physiology, and life history evolution since the 1950s  
89 (Huey *et al.* 1983; Milstead 1967; Vitt and Pianka 1994). Early research on Australian lizards  
90 contributed particular advances in understanding how placentas and viviparity evolve  
91 (Weekes 1935), and in community ecology (Pianka 1969; Pianka 1989).

92

93 Our objective with this review is to highlight the reproductive traits exhibited by Australian  
94 lizards that are of particular interest to biologists and students focused on broader  
95 evolutionary and ecological questions that apply to all animals. Many Australian lizard  
96 experts contributed their expertise to highlight some important areas where Australian lizards  
97 have, or could, contribute to answering broader questions in reproductive biology. We  
98 structured our review by covering topics in the sequence in which they occur during a  
99 reproductive cycle: behaviours and signalling involved in courtship; mechanisms involved in  
100 mating, egg production, and sperm competition; nesting and gestation; sex determination; and  
101 finally, birth in viviparous species.

## 102 **1: Pair bonds and sociality**

103 Social behaviour can be as simple as an interaction between two individuals that briefly meet,  
104 or complex social interactions among multiple individuals that form large stable groups that  
105 interact over extended periods of time. Such social groups emerge as the result of either

106 family members remaining together (e.g. fraternal societies) or through non-random  
107 associations of unrelated individuals (e.g. egalitarian societies). Both cases can result in  
108 individuals becoming mutually dependent on one another, sometimes forgoing their own  
109 reproduction and instead assisting with the reproduction of others. The origin of complex  
110 social behaviour and societies has been highlighted as a major evolutionary transition  
111 (Maynard Smith and Szathmary 1997). Thus, understanding the factors responsible for the  
112 evolutionary origins of such societies is a key challenge for biologists.

113 Traditionally, the flagship vertebrate taxa for understanding the evolution of complex  
114 sociality have been co-operatively breeding birds and mammals, organisms that live in  
115 complex societies (Rubenstein and Abbot 2017). Lizards, in contrast, have been largely  
116 ignored. However, a new social paradigm has emerged in lizards based on the recently-  
117 recognised presence of stable, long-term, and sometimes complex social associations in  
118 multiple, independent taxa (Doody *et al.* 2013; Doody *et al.* 2021a; Gardner *et al.* 2016;  
119 Halliwell *et al.* 2017b). The best example of such complex social organisation comes from an  
120 Australian group of scincid lizards, the Egerniinae. The Egerniinae include 62 species in 8  
121 genera (*Egernia*, *Liopholis*, *Bellatorias*, *Lissolepis*, *Tiliqua*, *Cyclodomorphus*, *Corucia*,  
122 *Tribolonotus*) (Gardner *et al.* 2008; Uetz 2010). Many species in the Egerniinae are  
123 characterised by the presence of long-term social associations both between adults, and  
124 between adults and their offspring, which result in the formation of stable family groups  
125 (Gardner *et al.* 2016; While *et al.* 2019; Whiting and While 2017). Crucially, the presence of  
126 these stable social associations and the diversity of social organisation (solitary, pair-bonds,  
127 facultative family living, obligate family living) makes the Egerniinae an outstanding model  
128 system for understanding the factors that have facilitated the origin and elaboration of  
129 complex social organisation.

130 The key foundation of social organisation in the Egerniinae is the presence of long-term pair-  
131 bonds that form between males and females. Pair-bonds were first identified in sleepy lizards  
132 (*Tiliqua rugosa*) (Bull 1988; Bull 2000), where they are stable and may be life-long (Bull *et*  
133 *al.* 2017; Leu *et al.* 2015). Similar long-term pairings underpin social organisation across  
134 most social Egerniinae (Chapple 2003; Whiting and While 2017). The nature of these pair-  
135 bonds can be variable across species. Indeed, while the predominant form of pair-bonding  
136 within Egerniinae is monogamy, in some species, males form bonds with multiple females  
137 (Chapple and Keogh 2006; While *et al.* 2019; While *et al.* 2009b; While *et al.* 2011).

138 In other species, social bonding can extend to multiple adults of both sexes (Duffield and Bull  
139 2002; Gardner *et al.* 2002a; Gardner *et al.* 2001), although often only a small number of  
140 individuals within these groups actually mate (Gardner *et al.* 2012). Long-term data on pair  
141 bonding are relatively sparse for most species, except *T. rugosa*. However, pair stability over  
142 multiple breeding seasons has been identified for *Egernia cunninghami* (Stow and Sunnucks  
143 2004), *E. saxatilis* (O'Connor and Shine 2003), *E. stokesii* (Duffield and Bull 2002; Gardner  
144 *et al.* 2002b), and possibly *E. striolata* (Riley unpubl. data) and *Liopholis whitii* (While *et al.*  
145 unpubl. data). While the occurrence of these long-term pair-bonds is well-established, we  
146 know relatively little about their functional significance. The reproductive benefits of  
147 maintaining a stable pair-bond compared to switching partners (but see Bull 2000; Leu *et al.*  
148 2015), the short and long-term trade-offs between maintaining single versus multiple social  
149 bonds, and the factors that initiate pair separation, remain largely unknown. Addressing these  
150 questions has the potential to reveal the factors that promote and maintain the initial  
151 emergence of long-term pair-bonds, which is relevant to other vertebrates in which family life  
152 is based upon the maintenance of social monogamy (e.g. Young *et al.* 2019).

153 Stable social associations between males and females in the Egerniinae vary not only in their  
154 nature but in the extent to which they correspond to patterns of paternity acquisition. On  
155 average, members of the Egerniinae exhibit some of the lowest levels of female polyandry  
156 (i.e. mating with one or more males) in lizards (Uller and Olsson 2008), with genetic  
157 monogamy the most pervasive mating system. Despite the low levels of polyandry, most  
158 species exhibit at least some multiple mating and/or extra-pair mating. In social *Egernia*  
159 species, this ranges from 2.6 % of litters including extra-pair paternity in *E. cunninghami*  
160 (Stow and Sunnucks 2004) to 10 – 30 % for other species (e.g. Bull *et al.* 1998b; Chapple and  
161 Keogh 2005a; Gardner *et al.* 2001; O'Connor and Shine 2003; While *et al.* 2014; While *et al.*  
162 2009b). In contrast, Egerniinae species that do not live in social groups exhibit higher levels  
163 of genetic polyandry. For example, 75 % of *Tiliqua adelaidensis* litters show evidence of  
164 multiple mating (Schofield *et al.* 2014). Differences in the proportions of polyandry among  
165 species, coupled with variation in the extent of social complexity (see below), make the  
166 Egerniinae an excellent model for testing hypotheses about the role that genetic monogamy  
167 has played in the evolution of social organisation (e.g. Cornwallis *et al.* 2010; Hughes *et al.*  
168 2008; Lukas and Clutton-Brock 2012).

169 Social aggregations in the Egerniinae are not just underpinned by prolonged associations  
170 between males and females, but also by prolonged associations between parents and their  
171 offspring. Indeed, offspring often delay dispersal and remain in their parents' refuge,  
172 sometimes for several years. The number and duration of these parent-offspring associations  
173 varies across the Egerniinae - from species in which these associations are absent [e.g. *T.*  
174 *rugosa* (Bull and Baghurst 1998)], to species in which parents exhibit facultative associations  
175 with a single cohort of offspring [e.g. *L. whitii* (Chapple and Keogh 2006; While *et al.*  
176 2009b), *Liopholis slateri* (Fenner *et al.* 2012), *Egernia saxatilis*, (O'Connor and Shine 2003)],  
177 to species that live in large social groups with multiple cohorts of young. For example, *E.*

178 *cunninghami* and *E. stokesii* can have up to five generations of offspring co-occurring in a  
179 single-family group (Gardner *et al.* 2001; Stow *et al.* 2001). Even in the mostly solitary *T.*  
180 *adelaidensis*, offspring can stay in natal burrows for several days before dispersing (Pearson  
181 *et al.* 2016). The associations between parents and offspring within the Egerniinae have  
182 several functional consequences that may constitute simple forms of parental care. For  
183 example, offspring gain increased access to basking sites, foraging opportunities, and retreat  
184 sites (Bull and Baghurst 1998; Munch *et al.* 2018; O'Connor and Shine 2004), which may  
185 result in an increase in early growth and survival (Botterill-James *et al.* 2016). Offspring also  
186 benefit from extended parent–offspring associations via a reduction in the risk of conspecific  
187 aggression and infanticide (O'Connor and Shine 2004; Sinn *et al.* 2008) and/or predation  
188 (Masters and Shine 2002; Watson *et al.* 2020). For example, the presence of a parent *E.*  
189 *saxatilis* eliminates aggression displayed toward offspring by unrelated adults (O'Connor and  
190 Shine 2004), and female *E. cunninghamii* actively chase off predatory snakes in the presence  
191 of their offspring (Watson *et al.* 2020). Finally, offspring may inherit territories via prolonged  
192 parent–offspring associations, as suggested by the high levels of genetic relatedness within  
193 social groups of *E. stokesii* (Gardner *et al.* 2001) as well as the long term residency of  
194 individuals in groups (Pearson *et al.* 2016).

195 Results from the Egerniinae clearly highlight the diversity in social traits that exist within the  
196 group. Future research needs to combine our understanding of diversity in social complexity  
197 with experimental tests that target the factors that mediate the expression and nature of these  
198 simple forms of social behaviour (e.g. Halliwell *et al.* 2017a; Halliwell *et al.* 2017b) in a  
199 phylogenetic context. This combination will allow us to connect plasticity in social behaviour  
200 at the individual level with evolutionary divergence of social complexity at the population  
201 and species level. Such research has the potential to generate an understanding of how  
202 different social entities (males, females, offspring, siblings) initially come together, how such

203 social associations are maintained in the face of inevitable conflicts that arise, and ultimately,  
204 how the stabilisation of these associations provides a foundation for the elaboration of social  
205 behaviour. The growing appreciation that simple social behaviour, based around family life,  
206 has emerged independently in other lizard families (e.g. Cordylidae, Xantusiidae) (Gardner *et al.*  
207 *al.* 2016; Halliwell *et al.* 2017b) means that we can apply our understanding of social  
208 evolution in the Egerniinae to identify commonalities across transitions to social life in other  
209 social lizards. Furthermore, the Egerniinae is ideally placed to integrate a detailed  
210 understanding of the phenotypic convergence in social evolution with neurological, genomic  
211 and developmental approaches that tell us how social traits converge at a mechanistic level.  
212 Combined, this approach has the potential to place the Egerniinae alongside other emerging  
213 socio-genomic/neuro-molecular vertebrate models, such as cichlids, frogs, butterfly fishes,  
214 birds, and mammals (e.g. Bukhari *et al.* 2019; Fischer *et al.* 2019; Nowicki *et al.* 2020;  
215 O'Connell *et al.* 2012) as a major future contributor to our understanding of what it takes to  
216 make an organism social.

## 217 **2: Pheromones and reproduction in Australian squamates**

218 Pheromones are chemicals that elicit a behavioural or physiological response in others of the  
219 same species (Karlson and Lüscher 1959; although see distinctions of Wyatt 2010), and are  
220 important to the social behaviour of squamates, including behaviour tied to reproduction.  
221 Pheromones influence behaviours such as locating, recognising, and choosing mates;  
222 antagonistic interactions that can determine the availability of mates; and parent–offspring  
223 recognition (reviewed by Houck 2009; Martín and López 2011; Mason and Parker 2010).  
224 Production of pheromones involved in reproductive signalling is controlled by reproductive  
225 hormones in squamate reptiles (Parker and Mason 2012; Parker and Mason 2014).  
226 Conceivably, pheromones could influence cryptic female choice after mating has already

227 happened (e.g. choosing to use or reject sperm from certain males). Additionally, pheromones  
228 could act as ‘primers’ that influence hormonal responses in the receiver (Bradbury and  
229 Vehrencamp 2011); for example, pheromones could induce or suppress sexual receptivity in  
230 the receiver (Mason and Parker 2010). However, many of the details of pheromones and  
231 reproductive behaviour remain to be tested in squamates.

232 Squamates produce pheromones from various regions of the body. Pheromones can be  
233 secreted from the skin (e.g. Mouton *et al.* 2014; Mouton *et al.* 2010; Van Wyk and Mouton  
234 1992), including from specialised epidermal glands, such as generation glands and follicular  
235 pores (Cole 1966; Maderson 1972; Mayerl *et al.* 2015), or from specialised glands within the  
236 cloaca (Cooper and Grastka 1987; Cooper and Trauth 1992; Siegel *et al.* 2014). Pheromones  
237 identified from the scats of several Australian Egeiinae (Bull *et al.* 1999a; Bull *et al.* 2000;  
238 Bull *et al.* 1999b) are likely produced from glands in the cloaca and deposited onto the  
239 surface of the faeces, although this hypothesis remains untested. Variation in the presence of  
240 cloacal glands across squamates has not been widely assessed, but epidermal glands, which  
241 are visible to the naked eye and thus better known, vary widely in their presence among  
242 squamate lineages (García-Roa *et al.* 2017). Epidermal glands can vary between sexes,  
243 typically with glands being better developed or only present in males (Mayerl *et al.* 2015). In  
244 some Australian lizards, such as *Amalosia* and *Nactus* geckos, the presence and absence of  
245 male epidermal pores vary even among closely related populations (Zozaya and Hoskin pers.  
246 obs.). Variation in femoral pore size and extent covaries with environmental variation in the  
247 *Ctenophorus maculatus* species complex, (Edwards *et al.* 2015), suggesting ecological  
248 factors may influence the evolution of epidermal glands in these species. Evolutionary trade-  
249 offs with other signalling modalities, such as visual displays, could also explain some  
250 variation in chemical signalling investment and the presence and extent of epidermal pores  
251 (e.g., *Sceloporus*; Campos *et al.* 2020; Ossip-Klein *et al.* 2013) but has yet to be explored in

252 Australian lizards. The drivers of anatomical variation in pheromone production, and its  
253 consequences for behaviour, are poorly understood areas ripe for further study, with agamids  
254 and *Amalosia* and *Nactus* geckos being potentially good models.

255 While most research on pheromones in squamates has focused on European and New World  
256 species, there are a number of studies on Australasian taxa that have focused on pheromone-  
257 mediated discrimination (Table 1). These studies do not conclusively demonstrate a  
258 pheromonal role in reproduction, but they do suggest the potential for influencing mate  
259 choice and antagonistic behaviours that could lead to assortative mating. The link between  
260 pheromone variation and reproductive behaviours remains to be explicitly tested in most  
261 Australian taxa, except for in the sea krait genus *Laticauda* (Shine *et al.* 2002c). Assessing  
262 female receptivity (Head *et al.* 2005), and recognising and trailing mates (Bull *et al.* 1993a;  
263 Bull and Lindle 2002; Olsson and Shine 1998) are key for identifying, finding, and guarding  
264 potential or actual mates. Additionally, the capacity for reciprocal mother–offspring  
265 recognition (Head *et al.* 2008) is important to mediate post-natal conflict and care. Beyond  
266 these areas, more in-depth studies are needed to determine the precise ways in which  
267 pheromones influence behaviour and reproduction.

268 Pheromonal research in Australia has focussed heavily on skinks, particularly those in the  
269 subfamily Egerniinae (Table 1), which exhibit complex sociality (Whiting and While 2017).  
270 However, pheromones are likely important to many squamate taxa. In fossorial taxa (e.g.  
271 *Anomalopus*, *Lerista*, *Aprasia*, blind snakes), pheromones are probably the most important, if  
272 not the only, signalling trait facilitating the finding and choosing of mates. Furthermore,  
273 recent work showing divergence in the chemical blends of epidermal pore secretions among  
274 morphologically similar, but genetically divergent, gecko lineages suggests pheromones may  
275 be important for mediating reproductive isolation in ‘cryptic species’ complexes (Zozaya *et*

276 *al.* 2019). Continued research into the form and function of squamate pheromone systems is  
277 needed to better understand their influence on reproduction and their evolutionary  
278 consequences (e.g. speciation, evolution of sociality). Australia’s extraordinary squamate  
279 diversity — with its corresponding diversity in morphology (for example in epidermal pores),  
280 ecology, social systems, and behaviour, and the presence of several clades with worldwide  
281 distributions — present multiple potential model systems for pursuing lizard pheromone  
282 research.

### 283 **3: Colour polymorphisms and alternative reproduction tactics in *Ctenophorus* spp.**

#### 284 *Intraspecific variation in male colour and reproductive traits*

285 Colour polymorphism is the co-existence of two or more distinct, heritable morphs within an  
286 interbreeding population, with the rarest occurring too frequently to be solely the result of  
287 recurrent mutation (Huxley 1955). Colour polymorphic species are valuable for examining  
288 the relationships between traits under both natural and sexual selection because the colour  
289 morphs are a convenient visual indicator of specific reproductive behaviours and often carry  
290 physiological costs (Stuart-Fox *et al.* 2021). Trade-offs between body size, colouration,  
291 aggressiveness, and postcopulatory sexual selection often occur in polymorphic species in  
292 which males compete for reproductive success using alternative mating tactics, for example,  
293 where one morph is most successful in monopolising females through territoriality and  
294 another morph is more successful in fertilization through subterfuge (Sinervo and Lively  
295 1996). Australia has two lizard species that have been used as model systems for such  
296 research: *Ctenophorus pictus* (painted dragon) and *C. modestus* (swift dragon). Male *C.*  
297 *pictus* are polymorphic in two aspects: head colour [(red, orange, yellow, or blue (Olsson *et*  
298 *al.* 2007b)], and the presence or absence of a yellow gular bib (Olsson *et al.* 2009a).

299 Similarly, male *C. modestus* are polymorphic for throat colour [orange, yellow, yellow with  
300 an orange centre, and grey (Teasdale *et al.* 2013)]. Colour traits in both species correspond to  
301 alternative behavioural, physiological, and life-history traits. Both species have been used to  
302 study different aspects of alternative reproductive tactics and sexual selection, with *C. pictus*  
303 as a model system for demonstrating the physiological costs associated with each morph, and  
304 *C. modestus* as a model for behaviour and aspects of coloration (e.g. visual conspicuousness,  
305 genetic and biochemical basis).

306 Colour traits in *C. pictus* are associated with trade-offs between investments in male  
307 aggressiveness, metabolism, condition loss, postcopulatory success, and telomere  
308 maintenance (a biomarker of stress and ageing) (Olsson *et al.* 2018; Olsson *et al.* 2017),  
309 which are mediated by oxidative stress (Friesen *et al.* 2017a; Friesen *et al.* 2020b; Friesen *et*  
310 *al.* 2021; Friesen *et al.* 2017b; Healey and Olsson 2009; Healey *et al.* 2007; McDiarmid *et al.*  
311 2017; Olsson *et al.* 2007a; Olsson *et al.* 2009a; Rollings *et al.* 2017). Bibbed and red-headed  
312 males invest more in traits that aid in precopulatory sexual selection, whereas yellow and  
313 non-bibbed males invest more in traits that aid in postcopulatory sexual selection, including  
314 sperm competition (Friesen *et al.* 2020a) (see Section 6). Red-headed and/or bibbed males  
315 tend to be more aggressive, and more likely to win male-male contests (Healey *et al.* 2007;  
316 McDiarmid *et al.* 2017). Bibbed males are also more attractive to females (McDiarmid *et al.*  
317 2017). In contrast, yellow and non-bibbed males share similar reproductive tactics that  
318 include having larger testes (Olsson *et al.* 2009b) that produce more and faster sperm (Friesen  
319 *et al.* 2020c), but with significantly shorter copulation durations than either red or bibbed  
320 males (Friesen *et al.* 2020c; Olsson *et al.* 2009b). Although overall rates of multiple paternity  
321 are relatively low (~15 % in the wild), yellow males have three times greater paternity  
322 success in head-to-head sperm competition trials over red males (Olsson *et al.* 2009b).

323

324 Bibbed male *C. pictus* do not lose paternity to neighbouring males in the field, probably  
325 because of effective territorial defence and mate guarding (Healey and Olsson 2009; Olsson  
326 *et al.* 2009a). Their investment in precopulatory traits carries a cost: both red-headed and  
327 bibbed males have significantly shorter blood cell telomeres (a potential marker of ageing  
328 and stress) than yellow-headed males and non-bibbed males (Friesen *et al.* 2019; Friesen *et*  
329 *al.* 2017a; Olsson *et al.* 2007a). In contrast, their sperm telomeres are longer (Friesen *et al.*  
330 2020c). Sperm telomere length is negatively related to sperm swimming velocity (Friesen *et*  
331 *al.* 2020c). The potential for sperm telomere length to influence sperm competitiveness or  
332 biases in female sperm storage are exciting ideas that remain to be tested (Friesen *et al.*  
333 2020c; Olsson *et al.* 2018; Olsson *et al.* 2017), as they may have implications for sperm  
334 competition across animals.

335

336 The male morphs of *C. modestus* exhibit different reproductive behavioural strategies, with  
337 orange as the most aggressive and grey as the least aggressive. The aggression level of yellow  
338 and orange-yellow morphs is conditional on the competitor's morph, where they are more  
339 aggressive to yellow and orange than to grey. The grey morph is the least bold, with similar  
340 boldness across the remaining morphs (Yewers *et al.* 2016). Differences in morph  
341 behavioural strategy correspond to differences in circulating baseline concentrations of  
342 androgen hormones. The aggressive orange morph has higher concentrations of androgens  
343 than either grey or yellow morphs (Yewers *et al.* 2017).

344

345 Although both *C. modestus* and *C. pictus* exhibit colour polymorphisms, the two species  
346 differ in their geographic distributions of morphs. All populations of *C. modestus* contain the  
347 same four morph types, but morph frequencies vary greatly among populations. There is  
348 evidence that spatial variation in selection shapes morph frequency patterns in *C. modestus*,

349 given strong relationships between morph frequencies and mean annual aridity index and  
350 vegetation cover, with no relationships between genetic similarity or geographic distance  
351 (McLean *et al.* 2015). In contrast, colour traits in *C. pictus* vary in abundance geographically.  
352 It is unclear if the apparent costs and benefits of alternative strategies influence fluctuations  
353 in morph frequency across years. From research on *Uta stansburiana* in the USA,  
354 fluctuations of morphs may be driven by female preference (Alonzo and Sinervo 2001) from  
355 year to year, but they also vary across geographic space from monomorphic to trimorphic  
356 (Corl *et al.* 2010b). Explanations for this variation centre on sexual selection as the cause of  
357 variance in morph number, given a positive relationship between sexual size dimorphism and  
358 morph number in *U. stansburiana* (Corl *et al.* 2010a). However, this relationship may not be  
359 indicative of variation in sexual selection (Chelini *et al.* in review). Instead, it may be driven  
360 by common environmental drivers, which each independently influence the proportion of  
361 sexual size dimorphism and the number of morphs in a population (Chelini *et al.* in review).  
362 Environmental variables similarly influence the proportion of morphs in *C. modestus* (McLean  
363 *et al.* 2015), but this has not yet been explored in *C. pictus*. By what mechanisms the  
364 environment regulates morph frequencies in polymorphic species is yet to be determined. It is  
365 possible that alternate numbers of morphs represent alternate resolutions of sexual conflict  
366 over mating dictated by environmental conditions (Svensson *et al.* 2020).

### 367 *Intraspecific variation in female reproductive traits*

368 Lizard species also exhibit variation in reproductive output, including egg size, egg number,  
369 and reproductive frequency, which collectively determine total per-year fecundity.

370 *Ctenophorus modestus* and *C. decresii* (tawny dragon) are sister species within the *C.*  
371 *decresii* species complex (Dong *et al.* 2021; McLean *et al.* 2014) which differ in maternal  
372 reproductive strategy. *Ctenophorus modestus* females exhibit higher fecundity over a

373 breeding season and produce larger clutches with lower individual egg masses and more  
374 clutches per season (Dong CM 2021). In contrast, *C. decresii* females produce smaller  
375 clutches of larger eggs, and seldom more than one clutch per season. *Ctenophorus modestus*  
376 clutches also have longer incubation periods than do *C. decresii*.

377

378 The differing reproductive strategies of the species may be attributable to environment-  
379 mediated selection. Both species are endemic to South Australia: *Ctenophorus modestus*  
380 occurs in the Flinders and Olary Ranges, whereas *C. decresii* occurs in the Mount Lofty  
381 Ranges, on the Fleurieu Peninsula, and on Kangaroo Island. The two species were long  
382 considered ‘lineages’ of *C. decresii sensu lato*, but were recently elevated to separate species  
383 on the basis of morphological and genetic differentiation (Dong *et al.* 2021). Lineage  
384 divergence may have resulted from local adaptation to separate refugia during glacial-  
385 interglacial Pleistocene cycles (Byrne 2008; McLean *et al.* 2014). The evolution of  
386 differences in reproductive characteristics between the two species presents an opportunity to  
387 investigate the potential influence of environment-mediated selection on reproductive and  
388 other life history traits.

389

390 Longer active seasons may allow for a higher laying frequency in *C. modestus*, and  
391 conversely, shorter active seasons constrain the breeding season of *C. decresii*. A shorter  
392 active season also restricts the time for embryonic development and selects for a higher  
393 investment in the first clutch. This difference in reproductive output aligns with predictions  
394 that cold environments at high latitudes favour the production of fewer and larger eggs to  
395 increase offspring survival (Fischer *et al.* 2003; Morrison and Hero 2003; Yampolsky and  
396 Scheiner 1996). However, patterns of reproductive traits in lizards are not always globally  
397 congruent (Meiri *et al.* 2020; Sun *et al.* 2013). Counter-gradient variation in patterns of

398 reproductive biology may be driven by many additional abiotic and biotic factors beyond  
399 latitude and climate (e.g. resource availability, predation risk, microhabitat preferences;  
400 Angilletta 2009; Mesquita *et al.* 2016). Furthermore, lizards may compensate for  
401 environmental factors in ways such as adjusting seasonal timing of reproduction (Warner and  
402 Shine 2007) or developmental mechanisms such as embryonic development rates  
403 (Niewiarowski and Angilletta 2008; Oufiero and Angilletta Jr. 2006), physiological pathways  
404 (Sun *et al.* 2013), and switching between semelparity and iteroparity (Eckhardt *et al.* 2017).  
405 Australia's diversity of lizard species and geography presents an ideal model for testing  
406 hypotheses about how lizard reproductive traits evolve to cope with the constraints caused by  
407 different environments, because some species are distributed across a variety of environment  
408 types (Cogger, 2018; Wilson and Swan, 2021). The examples of geographic differences in  
409 reproductive output we present for *Ctenophorus* species are likely to also occur in other  
410 species with large distributions across varied environments, such as *Lialis burtoni*, *Varanus*  
411 *gouldi*, and *Tiliqua scincoides*.

#### 412 **4: Sexual selection on the neural control of reproduction**

413 One of the few areas in neuroscience where lizards have made significant contributions is the  
414 neural control of reproductive behaviour (Lovern *et al.* 2004; Wade 2011). Across all  
415 vertebrates, two of the key brain regions controlling reproductive behaviour are the medial  
416 preoptic nucleus and the ventromedial hypothalamic nucleus, both located in an area called  
417 the diencephalon. The medial preoptic nucleus drives male sexual behaviour while the  
418 ventromedial hypothalamic nucleus drives female behaviour (Numan 2014), and the  
419 interaction between hormones and these two brain regions largely governs reproductive  
420 behaviour. Environmental influences, such as time of year and presence of conspecifics,  
421 affect reproductive behaviour in part by driving temporary changes to the size and the activity

422 of these brain regions (Beck *et al.* 2008; Wade *et al.* 1993). In Australian agamid lizards,  
423 selection can drive more permanent changes in these brain regions over evolutionary time  
424 (Hoops *et al.* 2017).

425 Agamids are a particularly attractive lineage in which to study evolutionary changes in brain  
426 structure. They have the lowest known coefficient of brain allometry amongst lizards, and the  
427 highest known encephalisation quotient (Black 1983). Brain allometry is a measure of how  
428 closely changes in body size are paralleled by changes in brain size, while the encephalisation  
429 quotient estimates how “enlarged” a brain is relative to a standard brain for a given body size  
430 (Platel 1979). The net result is that agamids have unusually large brains (for lizards) and  
431 relatively little variation in brain volume based on body size, which provides an opportunity  
432 for research.

433 Sexual selection drives both an increase in the volume of the medial preoptic nucleus and a  
434 decrease in the volume of the ventromedial hypothalamic nucleus in male *Ctenophorus* spp.  
435 dragons (Hoops *et al.* 2017). Thus, sexual selection possibly increases motivation and drive  
436 to perform male reproductive behaviours. However, brain region volumes do not differ across  
437 female *Ctenophorus* species, suggesting that sexual selection does not drive changes in their  
438 female reproductive behaviour. This suggests a passive role for females in reproduction, and  
439 is consistent with behavioural studies in some species of *Ctenophorus* (Jansson *et al.* 2005;  
440 Lebas 2001; Olsson 2001). In other lizard species, however, females can play very active  
441 roles in reproduction, including female-specific evolutionary changes in behaviour and  
442 motivation. For example, the female Lake Eyre dragon (*Ctenophorus maculosus*) displays  
443 unique reproductive colouration and behaviour that signals to males whether she is receptive  
444 to a male’s advances (Olsson 1995). However, the brain structure of *C. maculosus* has not  
445 been compared to those of species with more passive reproductive behaviour. A behavioural

446 innovation like that in female *C. maculosus* is presumably the product of neuroevolutionary  
447 innovation, and presents an opportunity to further understand how sexual selection can alter  
448 the brain.

449 Additional studies of brain anatomy across Australian lizards have great potential to develop  
450 our understanding of brain evolution. Approaches examining similar patterns of brain  
451 evolution with respect to sexual selection could reveal how generalizable the findings in the  
452 study of *Ctenophorus* species are to other lizards, and tease apart specific behaviours and  
453 ecological factors (such as territoriality, seasonality, or life history) that may influence how  
454 sexual selection shapes brain structure. There are additional topics, such as pair bonding  
455 (Section 1) and parental care, where Australian lizards have the potential to make seminal  
456 contributions to our understanding of the neural underpinnings of reproductive behaviour,  
457 and how these traits evolve. The strength of Australian lizards as models for study of brain  
458 anatomy and function is the breadth of diversity of reproductive strategies, how our intimate  
459 understanding of this diversity can be leveraged towards understanding the underlying neural  
460 control, and what that means for the evolution of diverse reproductive strategies across  
461 vertebrates. The findings presented here represent the first step forward; the potential in this  
462 area is almost limitless.

## 463 **5: Genital variation**

464 Postcopulatory sexual selection is likely the main evolutionary force driving diversification in  
465 genital morphology in most animals (Brennan and Prum 2015; Simmons 2014).

466 Understanding intrasexual and intersexual variation in genital morphology is critical for  
467 understanding mating-system dynamics, sexual conflict, and cryptic female choice. This  
468 knowledge is especially important given the diversity of mating systems within Australian  
469 lizards, each representing different opportunities for conflict and postcopulatory choice.

470 Genital traits may also have an underappreciated role in understanding species diversification  
471 in lizards (Klaczko *et al.* 2015), as these traits are often involved in the development of  
472 reproductive isolation among putative species. Most of our understanding of genitalia in  
473 Australian lizards comes from work describing genital development in relation to sex  
474 determination (Whiteley *et al.* 2017b). In squamates, males have hemipenes, whereas females  
475 may or may not have analagous structures called hemiclitores (Böhme 1995; Martínez-Torres  
476 *et al.* 2015).

477 Research on hemipenal morphology is limited in Australian lizards. In a broadscale study  
478 across *Varanus*, including Australian species, hemipenal morphology is more  
479 phylogenetically informative than non-genital morphological characters; this pattern could  
480 also be indicative of faster evolving genitalia like that seen in *Anolis* species (Klaczko *et al.*  
481 2015). Understanding such variation in Australian lizards may be important for understanding  
482 diversification dynamics in rapidly-radiating Australian lizard lineages. Anecdotally,  
483 Australian agamids have larger hemipenes (relative to SVL) compared to other ecologically-  
484 similar lizard families on other continents (i.e., phrynosomatids; D.L. Edwards pers. obs.).  
485 Greater hemipenal lengths are associated with greater copulation frequencies between species  
486 of *Anolis* lizards (Johnson *et al.* 2011). Differences in relative hemipenal size could therefore  
487 suggest fundamental differences in mating frequencies across lizard families. In *Liolaemus*  
488 species, hemipenal eversion is part of conspecific male-male aggressive displays (Ruiz-  
489 Monachesi *et al.* 2019). The potential for hemipenal morphology to play a similar role in  
490 Australian lizards remains to be investigated.

491 In contrast to males, very little is known about female genital evolution in squamates. Female  
492 genitals in squamates can be present as a rudimentary structure (Neaves *et al.* 2006), as  
493 hemiclitores (Böhme 1995; Martínez-Torres *et al.* 2015), or as miniaturized hemipenes

494 (Telemeco 2015). Even when present, they can also exhibit different coloration from  
495 conspecific male hemipenes (Valdecantos and Lobo 2015). The functional role of female  
496 genitalia in squamates, and its variability, is unknown. *Varanus* spp., which are especially  
497 diverse across Australia, exhibit morphological diversity in hemiclitoros (Böhm *et al.* 2013;  
498 Böhme 1995; Böhme and Ziegler 2009; Ziegler *et al.* 2005; Ziegler *et al.* 2007), yet a  
499 comprehensive exploration of hemiclitoral morphology or even presence/absence in any  
500 Australian lizard genus, including *Varanus* (but see (King and Green 1999)), is currently  
501 lacking. Hemiclitoral morphology may relate to cryptic and post-mating female control of  
502 reproduction. A prevailing paradigm is that precopulatory female choice is rare in lizards  
503 (Olsson 2001; Olsson and Madsen 1995; but see Sullivan and Kwiatkowski 2007). Variation  
504 in the presence/absence of cloacal glands, sensory innervation, epithelial wall thickness, and  
505 presence/absence of sperm crypts in lizards suggests that females have morphologically  
506 variable structures capable of providing them with the opportunity for postcopulatory choice  
507 (Sánchez-Martínez *et al.* 2007). Hemiclitori may have been retained for similar roles, such as  
508 expulsion of sperm. Studies should be undertaken using lizards with different mating systems  
509 and morphologies to determine the extent to which female lizards are able to control  
510 copulation duration and fertilisation in association with hemiclitoral and cloacal-vaginal  
511 morphology, which would determine mating biomechanics, sexual conflict over mating and  
512 the evolution of postcopulatory female choice (Brennan 2016; Firman *et al.* 2017; Friesen *et*  
513 *al.* 2016; Friesen *et al.* 2014).

514 Work stemming from understanding genital development in relation to sex determination  
515 mode have provided some understanding of how lability in hemiclitoral structure develops.  
516 The central hypothesis is that genital variation occurs through differences in developmental  
517 programming among species. *Pogona vitticeps* exhibits temporary pseudohermaphroditism  
518 (TPH), whereby both ovaries and hemipenes are present at hatching (Whiteley *et al.* 2017a;

519 Whiteley *et al.* 2018). Females exhibit more lability in phenotype than males, which is not  
520 associated with temperature-dependent sex determination (Whiteley *et al.* 2017a).  
521 Thermostability of sex differentiation may play a role in an extended period of TPH, because  
522 other species with temperature-dependent or thermal influenced sex determination also show  
523 extended periods of TPH (Whiteley *et al.* 2018). While TPH complicates sex assignment  
524 based on hemipene eversion at birth in *P. vitticeps* (Whiteley *et al.* 2018), other Australian  
525 lizards (*Carinascincus* [formerly *Niveoscincus*] *ocellatus*) show clear genital differentiation  
526 at birth (Neaves *et al.* 2006). Resolving these differences among species is important for  
527 efforts to use genitalia to identify sex at birth. Given the diversity of sex determining  
528 mechanisms present within Australian lizards (Section 10), especially agamids, it is important  
529 to understand how genitalia develop in these different systems.

## 530 **6: Multiple paternity, sperm competition, and postcopulatory sexual selection**

531 Polyandrous mating systems are widespread throughout animal and plant taxa (Jennions and  
532 Petrie 2000; Pizzari and Wedell 2013; Taylor *et al.* 2014). Squamate reptiles are no exception  
533 (Friesen *et al.* 2020a; Uller and Olsson 2008), and are excellent models to test critical  
534 questions about female promiscuity. Except for sperm, male lizards and snakes do not  
535 directly provide females with resources during courtship, mating, or after offspring are  
536 hatched/born, although females of territorial species may receive benefits from residing on a  
537 resource-rich site (Uller *et al.* 2010). Thus, the evolution of polyandry in lizards and snakes is  
538 simplified compared to other vertebrates with parental care (mammals and birds), so the  
539 fundamental costs and benefits of polyandry can be isolated (Kvarnemo and Simmons 2013;  
540 Uller and Olsson 2008). One benefit of polyandry is to ensure that a female has enough  
541 sperm to fertilize her ova. Sperm limitation may drive multiple mating in common lizards  
542 (*Zootoca vivipara*) (Uller and Olsson 2005). Multiple mating is associated with higher

543 fecundity in lizards (LaDage *et al.* 2008; Noble *et al.* 2013; Uller and Olsson 2005; York and  
544 Baird 2019), but more studies specifically designed to tease apart the effects of sperm  
545 limitation and benefits of polyandry are needed in lizards. Social skinks and agamids with  
546 colour traits that may function in pre-copulatory sexual selection are ideal model systems,  
547 and both occur in Australia (Section 3).

548 A consequence of polyandry is that the battle for reproductive success does not always end  
549 after intrasexual competition for mates (usually males) or intersexual selection of mates  
550 (usually by females)—which together evolve as a result of precopulatory sexual selection  
551 (Andersson 1994; Darwin 1871). Instead, sexual selection continues within the reproductive  
552 tracts of promiscuous females, where the sperm of different males compete to fertilise eggs  
553 (Parker 1970; Parker and Pizzari 2010), and where female traits bias the contest for  
554 fertilisation success of different males (Arnqvist 2014; Eberhard 1996; Thornhill 1983).  
555 Together, these phenomena evolve as a result of postcopulatory sexual selection. Sperm  
556 competition is the post-mating analogue of male-male competition for mates, which occurs  
557 when the ejaculates of more than one male overlap within the reproductive tract of a  
558 polyandrous female (Parker 1998). Sperm competition between multiple males usually results  
559 in multiple paternity when there is no complete bias towards a single competitor. Multiple  
560 paternity is pervasive in squamates studied thus far, with more than 50 % of clutches/litters  
561 exhibiting multiple paternity in the wild (Uller and Olsson 2008). Within lizards, rates of  
562 multiple paternity depend on how well males can monopolise access to females (Uller and  
563 Olsson 2008; Uller *et al.* 2010). Rates of multiple paternity are generally low (4 – 30 %) in  
564 Australian territorial agamids (Table 2), but higher in non-social skinks (43 – 94 %; Uller and  
565 Olsson 2008). The levels of polyandry in agamids (Table 2) contrast with ecologically-  
566 similar families of lizards where multiple paternity is much higher (40 – 80 %), for example,  
567 in the genera *Lacerta* in Europe (Fitze *et al.* 2005), and *Uta*, *Crotaphytus*, and *Sceloporus* in

568 North America (Abell 1997; Haenel *et al.* 2003; Peterson and Husak 2006; Zamudio and  
569 Sinervo 2000).

570 Female sperm storage, where females store sperm for weeks to years after mating, increases  
571 the chance that sperm from different males will compete (Parker 1970). Sperm competition  
572 for fertilisation produces strong selection on sperm and ejaculate traits (Lüpold *et al.* 2020),  
573 which may impose energetic costs and concomitant trade-offs with precopulatory traits (body  
574 size, aggressiveness, colouration) as we find in some Australian lizards (Section 3). Indeed,  
575 females of some Australian dragons store sperm across multiple clutches, creating the  
576 situation in which males may sire young posthumously (Olsson *et al.* 2009b). Female sperm  
577 storage could therefore shift male investment towards early-season mating success (Zamudio  
578 and Sinervo 2000). As a result, the evolution of female sperm storage, sperm longevity, and  
579 male lifespan are likely linked in some species. Females could select for sons that mature  
580 earlier, with longer-lived sperm, but die younger due to investments in early maturity. This  
581 pattern could be common in short-lived annual lizards common in Australia, like  
582 *Ctenophorus pictus* and *C. fordi* but has yet to be investigated.

583 Polyandry also has indirect genetic benefits in lizards and snakes (Wapstra and Olsson 2014),  
584 including positive fitness effects linked to offspring survival (Eizaguirre *et al.* 2007; Madsen  
585 *et al.* 1992), offspring dispersal (Laloi *et al.* 2009), improved heterozygosity for inbreeding  
586 avoidance, and “trading-up” to a better mate (Laloi *et al.* 2011; Olsson *et al.* 1996b; While *et*  
587 *al.* 2014). Post-copulatory processes and conditions within the female reproductive tract may  
588 allow females to select sperm from males less-related to themselves (Olsson *et al.* 1996a;  
589 Olsson *et al.* 2004; Olsson *et al.* 1997), or, in species with XX/XY chromosomal sex  
590 determination, allow females to select sperm to determine the sexes of their offspring in a  
591 sex-biased environment (Cox and Calsbeek 2010; Olsson *et al.* 2007b). Nevertheless, the

592 effects of multiple mating on offspring phenotypes are not always positive in lizards (Keogh  
593 *et al.* 2013; Noble *et al.* 2013), but these effects could also be cryptic in studies that do not  
594 analyse genetic interactions between the individuals participating in staged matings. In future  
595 work involving the assignment of parentage using molecular techniques, we encourage  
596 researchers to include enough markers to allow examination of genetic effects on the  
597 probability of paternity (e.g. relatedness) as well as male traits (genitalia, sperm, ejaculates)  
598 that may be used as cues by females to select paternity. Nearly nothing is known about  
599 female morphology, physiology, or behaviours that generate biases due to relatedness of their  
600 partners. Any polyandrous Australian species of lizard with a moderate length of sperm  
601 storage that demonstrates regular individual male-female interactions would be an ideal  
602 model for teasing apart the mechanisms of cryptic female choice. Dissociated mating and  
603 female sperm storage are relatively common in Australian skinks, including the genera  
604 *Carinascincus*, *Hemiergus*, and *Pseudemoia*, so these taxa might be useful models (Murphy *et*  
605 *al.* 2006).

## 606 **7: Nesting Ecology in Australian Goannas**

607 Most reptiles lack parental care and desert their eggs after depositing them in an excavated  
608 ground nest, burrow, or under objects such as rocks, logs, bark, or vegetation. For example,  
609 only a very small proportion of lizards attend their eggs (Shine 1988; Somma 2003), fuelling  
610 a perception that reproducing reptiles rarely engage in social behaviour after mating (but see  
611 Section 1). Moreover, excavating and backfilling shallow ground nests is conserved across  
612 many lizard families (Doody *et al.* 2009), with eggs predictably deposited in shallow nests in  
613 areas that are warm enough for successful development. In contrast, ‘deep-nesting’  
614 Australian monitor lizards (‘goannas’) highlight that the ecology and evolution of nest site  
615 choice behaviour in lizards is anything but stereotypical or conserved.

616 Where the yellow-spotted monitor, *Varanus panoptes*, and its sister species, Gould's monitor,  
617 *V. gouldii*, lay their eggs has only recently been identified. Aboriginal women suspected that  
618 eggs were laid in 'warrens', areas denuded of vegetation with multiple burrow entrances  
619 (Figure 2) (Christian 2004), which was confirmed by excavation beyond the terminus of open  
620 burrows. These warrens contain communal nests, solitary nests (with mothers returning to the  
621 same nest site year after year), nests with complex structures, and the deepest vertebrate nests  
622 in the world [up to 4 m deep; (Doody *et al.* 2015; Doody *et al.* 2014; Doody *et al.* 2018a;  
623 Doody *et al.* 2018b)]. Moreover, the burrows provide refuges, foraging sites, aestivation sites  
624 and nesting sites for many other animals, which is the first demonstrated case of ecosystem  
625 engineering in lizards (Doody *et al.* 2021b).

626 Each warren contains multiple nests for the present (communal nesting) and previous years'  
627 nesting (which may indicate traditional solitary nesting). For example, one portion of a *V.*  
628 *gouldii* warren contained 97 nests, including 21 nests with eggs (the rest were nests from  
629 previous years) (Doody *et al.* 2018a). Similarly, a *V. panoptes* warren contained 110 nests,  
630 including 11 with incubating eggs (Doody *et al.* 2018b). Extrapolation to the area of the  
631 entire *V. gouldii* warren predicted 53 nests with eggs, which implicates many mothers,  
632 despite the potential for multiple clutches per annum. When well-fed in captivity, *V. panoptes*  
633 mothers can produce multiple clutches (D. Kirschner, unpubl. data), but this is less likely in  
634 nature, especially in a food-limited desert ecosystem (Doody *et al.* 2018b). Communal and  
635 traditional solitary nesting in these species may be related to the ease in constructing deep  
636 nesting burrows in soil that has been loosened by conspecifics within and among years, or  
637 some other benefit to mothers, eggs or hatchlings (Doody *et al.* 2009).

638 Both *V. panoptes* and *V. gouldii* nests are the deepest for any vertebrate: mean nest depths are  
639 2.3-2.5 m (range 1.0 – 3.6 m; N=162) for *V. panoptes* and 3.0 m (range 1.8 – 4.0 m; N=103)

640 for *V. gouldii* (Doody *et al.* 2015; Doody *et al.* 2014; Doody *et al.* 2020; Doody *et al.* 2018a;  
641 Doody *et al.* 2018b). Most ground-nesting reptiles deposit their eggs at depths of 20 – 250  
642 mm, and even the gigantic leatherback sea turtle nests at depths averaging < 1 m (Doody *et*  
643 *al.* 2014). These two goanna species probably nest deeply to maintain moist conditions  
644 during the long (~8 month) incubation period that spans the entire dry season, rather than  
645 depth being related to temperature (Doody *et al.* 2015). *Varanus panoptes* nests are deeper in  
646 a desert site than in a woodland site with higher rainfall (Doody *et al.* 2018b), and *V.*  
647 *panoptes* mothers nest deeper in drier years (Doody *et al.* 2021b). Decoupling of temperature  
648 from moisture in reptile nests (Doody *et al.* 2021b) demonstrates plasticity in depth due to a  
649 soil moisture gradient; in all other (shallower) reptile nests, shallower nesting means warmer  
650 but drier conditions, complicating the isolation of whether temperature or moisture is the cue  
651 for plasticity in nest depth.

652 Nesting is complex in both *V. panoptes* and *V. gouldii*, which construct elaborate helical  
653 burrows consisting of straight sections to a depth of about 1.5 m, followed by 2 – 7 tight  
654 spirals descending straight down and terminating in a slightly enlarged egg chamber. Fossil  
655 helical burrows date back to 255 Mya [*Diictodon*, a mammal-like therapsid; (Smith 1987)]  
656 and more recently from 22 – 34 Mya [*Palaeocastor*, a terrestrial beaver; (Martin and Bennett  
657 1977)]. Extant animal species with helical burrows include pocket gophers, prairie dogs, and  
658 scorpions (reviewed in Doody *et al.* 2015). The adaptive advantages of helical burrows are  
659 untested for any species, but may include an anti-predator function, improved drainage or  
660 hatchling escape, providing a more stable microclimate, an anti-crowding mechanism, or  
661 mechanical advantage (reviewed in Doody *et al.* 2015). Interestingly, hatchling goannas  
662 appear not to use the spirals to emerge from the nest, but rather excavate a straight vertical  
663 tunnel, often through resistant soils (Doody *et al.* 2018a).

664 These varanids serve as ecosystem engineers for small animals, including 28 species of  
665 lizards (geckos, skinks, goannas), snakes, frogs, mammals, and numerous invertebrates. One  
666 warren contained 418 individual frogs (mostly *Uperoleia spp.*) that were aestivating there  
667 during the dry season (Doody *et al.* 2021a). The warrens also provided nesting habitat for  
668 another goanna species (*V. acanthurus*) and the gecko *Strophurus ciliaris* (Böhm *et al.* 2013;  
669 Doody *et al.* 2017; Doody *et al.* 2021b). These two goanna species thus join the burrowing  
670 tortoises of North America (*Gopherus*) and perhaps mound-nesting crocodylians as reptilian  
671 ecosystem engineers (Kinlaw and Grasmueck 2012; Somaweera *et al.* 2020).

672 Deep, complex communal nesting would not be expected to have evolved in desert or dry-  
673 adapted lizard species with shorter incubation periods, because the short incubation period  
674 would preclude the need for incubation over the entire dry season. The evolution of helical  
675 nesting remains enigmatic. Without knowing its value (adaptive or not), we cannot make  
676 predictions about its evolution, or lack thereof, in other lizard species. No other lizard species  
677 constructs such deep, helical burrows, but we predict that a third large, desert-inhabiting  
678 goanna with a long incubation period, the perentie (*V. giganteus*), may also nest at great  
679 depths due to its ecological similarity to its congeners. Deep helical nesting may also occur in  
680 populations of *V. griseus* near the Sahara Desert. As for why these goannas nest communally,  
681 the energy required to construct such deep complex burrows, and for the young to escape the  
682 burrows through resistant soils using their own tunnels, could select for mothers returning to  
683 communal nesting areas to take advantage of loosened soils. Returning to these sites would  
684 reduce the energetic costs of digging for both mothers and offspring, and thus allow both  
685 easier nesting, and easier escape from nests.

## 686 **8: Viviparity and placentation**

687 Australia is one of the best places in the world to study the evolution of viviparity and  
688 placentation, using reptiles as a model. Viviparity has evolved here repeatedly in both skinks  
689 and snakes (Blackburn 2015), and Australia has 20 % of the reliably classified species that  
690 are bimodally reproductive: *Lerista bougainvillii* (Greer 1989; Qualls and Shine 1998), and  
691 *Saiphos equalis* (Smith and Shine 1997). In these species, individuals in some locations lay  
692 eggs, whilst those in other locations give birth to live young, providing a unique opportunity  
693 for comparative research on the evolution, costs, and mechanisms underpinning parity mode  
694 within a single species. Complex placentation and obligate placentotrophy have also evolved  
695 at least twice in Australian skinks, in the genera *Carinascincus* and *Pseudemoia* (Stewart and  
696 Thompson 2004; Weekes 1935). These represent 40 % of the known origins of  
697 placentotrophy in lizards. A third skink species, *Eulamprus quoyii*, also exhibits a modified  
698 chorioallantoic placenta (Murphy *et al.* 2011), though it is not highly placentotrophic  
699 (Thompson 1977; Thompson 1981). Together, viviparity and placentation represent some of  
700 the best opportunities for studying how novel complex traits evolve in vertebrates, because  
701 they have evolved repeatedly so many times (Griffith and Wagner 2017) and provide the  
702 phylogenetic replication necessary for comparisons to be statistically robust. In Australia,  
703 most research on viviparity and placentation has focused on a relatively small group of skink  
704 species: *Sphenomorphus* group (*Eulamprus* spp., *Lerista bougainvillii*, *Saiphos equalis*),  
705 *Eugongylus* group (*Carinascincus* spp., *Pseudemoia* spp.), and Egerniinae (*Egernia* spp.,  
706 *Liopholis* spp., *Tiliqua* spp.). The parity mode of additional species has been assigned based  
707 on observations of reproductive behaviour or dissection of gravid female museum specimens  
708 (Blackburn 2000). It is unlikely that viviparity will be discovered in any lizard previously  
709 thought to be oviparous, but unexpected discoveries always have the potential to develop new  
710 avenues of research. For example, observation of both oviparity and viviparity *within an*

711 *individual* (the first in a vertebrate) in *S. equalis* raises the possibility that reproductive mode  
712 may be plastic in some species (Laird *et al.* 2019).

713 All viviparous lizards studied so far have at least a simple placenta that provides respiratory  
714 gas exchange and a site of embryonic anchoring to the uterus (Blackburn 2015). Some  
715 species have more complex nutritive placentas, in which substantial quantities of organic  
716 nutrients are transported from the mother to embryos during pregnancy (obligate  
717 placentotrophy). Detecting this feature is more difficult than identifying viviparity alone.  
718 Placentotrophic species are identified by examination of the morphology of the placental  
719 tissues and comparisons of the dry masses of freshly-ovulated eggs to those of newborn  
720 offspring (matrotrophy index; Thompson *et al.* 2000). Placentotrophic embryos increase in  
721 dry mass during development as mothers allocate nutrients to them, whereas non-  
722 placentotrophic embryos decrease in dry mass as they catabolise nutrients from their yolk  
723 (Stewart 1989; Swain and Jones 2000; Thompson *et al.* 2000). Placental transport of nutrients  
724 has also been demonstrated in Australian skinks using stable and radioisotopes (Itonaga *et al.*  
725 2012b; Jones and Swain 2006; Swain and Jones 1997; Thompson 1977). Within Australian  
726 lizards, placental structure and function have been most well-studied in the *Eugongylus* group  
727 skinks *Carinascincus* spp. and *Pseudemoia* spp. Even within these genera, the placental  
728 structures in *P. baudinii*, *P. rawlinsoni*, *C. palfreymani*, and *C. orocryptus* remain unstudied,  
729 and the degree of placental nutrient transport in *P. baudinii*, *P. rawlinsoni*, *C. greeni*, *C.*  
730 *palfreymani*, and *C. orocryptus* remains unknown. Furthermore, the amount of nutrients  
731 allocated across the placenta can vary within a species (Itonaga *et al.* 2012a; Thompson and  
732 Speake 2006; Van Dyke *et al.* 2014), so single estimates of matrotrophy index may not be  
733 representative of all individuals. The phylogenies for both *Carinascincus* and *Pseudemoia*  
734 remain poorly-resolved (Brandley *et al.* 2015; Hutchinson and Donnellan 1992). Basic

735 studies of placental structure and function in these taxa, and a robust phylogeny, are needed  
736 to determine how the placenta has evolved in these species.

737 Research on Australian lizards underlies much of our understanding of how viviparity and  
738 placentation evolve and function in reptiles, and this work provides a basis for comparison  
739 with other viviparous taxa. Early in lizard pregnancies, a plasma membrane transformation  
740 (PMT) homologous to that of mammals occurs, which likely allows for embryos to anchor to  
741 the uterus (Murphy *et al.* 2000). This phenomenon was first recognised in Australian skinks.  
742 In *Pseudemoia* spp., the PMT is facilitated partially by changes in the distributions of  
743 desmosomes, tight junctions, occludins and cadherins in the uterine epithelium (Biazik *et al.*  
744 2007; Biazik *et al.* 2008; Biazik *et al.* 2010; Wu *et al.* 2011). Once pregnancy begins,  
745 chorioallantoic and yolk sac placentas develop in all viviparous lizards, which facilitate  
746 contact between maternal and embryonic tissues (Thompson and Speake 2006). The  
747 functions of the two placentas likely vary across species, but the chorioallantoic placenta  
748 always exhibits dense capillary beds that presumably facilitate placental exchange, especially  
749 for respiratory gases. In *Eulamprus tympanum*, angiogenesis in uterine and embryonic  
750 components of the chorioallantoic placenta increases the density and apposition of capillaries  
751 in both tissues (Parker *et al.* 2010a; Parker *et al.* 2010b). In *Saiphos equalis*, angiogenesis is  
752 driven in part by vascular endothelial growth factors (VEGFs; Murphy *et al.* 2010;  
753 Whittington *et al.* 2015). Carbonic anhydrases in both uterine and embryonic components of  
754 the chorioallantoic placenta likely facilitate transport of carbon dioxide in *Pseudemoia*  
755 *entrecasteauxii* (Van Dyke *et al.* 2015). Most viviparous lizards also transport small amounts  
756 of nutrients to their developing embryos (Thompson *et al.* 2000), especially inorganic ions  
757 like calcium (Linville *et al.* 2010), which may replace the role of the eggshell as an ion  
758 reserve in oviparous species (Herbert *et al.* 2006). In Australian lizards, evidence of obligate  
759 placentotrophy of organic nutrients is so far limited to skinks in the genera *Carinascincus* and

760 *Pseudemoia*; candidate nutrient transporters in these taxa include lipoprotein lipase, and  
761 amino acid-transporting solute carriers (SLCs; Griffith *et al.* 2016; Griffith *et al.* 2013).  
762 Gene expression analyses, particularly transcriptomic studies, have identified thousands of  
763 genes that are likely to be involved in pregnancy in Australian lizards (Foster *et al.* 2020;  
764 Griffith *et al.* 2016; Griffith *et al.* 2017; Hendrawan *et al.* 2017). These genes likely  
765 contribute to key pregnancy-related functions including uterine remodelling, nutrient and  
766 respiratory gas transport, and immune regulation, and testing the functions of these genes is  
767 the next step in understanding their role in the evolution of viviparity.

768 Australian lizards also present an excellent opportunity to study the selective pressures that  
769 drive the evolution of viviparity and placentae, although these questions have received much  
770 less attention than the genetic, physiological, and morphological mechanisms that underpin  
771 both traits. Adaptive hypotheses for viviparity and placentation focus on the potential fitness  
772 benefits mothers gain by being able to continuously control the environment that developing  
773 embryos experience, and also on embryos' ability to impact maternal physiology to their own  
774 benefit (Crespi and Semeniuk 2004; Shine 1995; Tinkle and Gibbons 1977). Research  
775 integrating ecophysiological and molecular approaches is needed to test how viviparity and  
776 placentotrophy impact fitness, and whether the fitness benefits of these traits overrides their  
777 concomitant fitness costs. For example, viviparity may allow mothers to improve the fitness  
778 of their individual offspring, but it also reduces the number of reproductive events a single  
779 female can have per year, and increases the physical burden on the mother (Tinkle and  
780 Gibbons 1977). Australian skinks are ideal models to test these adaptive hypotheses given  
781 their diversity of reproductive modes, especially in taxa that overlap in distribution and  
782 ecology, as do the genera *Pseudemoia*, *Carinascincus*, *Bassiana*, and *Lampropholis*. The two  
783 reproductively bimodal species, *Lerista bougainvillii* and *Saiphos equalis*, also present an  
784 excellent opportunity to test how the fitness costs and benefits of reproductive mode vary

785 with environment within single species. Simultaneously, they provide the opportunity to  
786 determine whether gene flow between viviparous and oviparous populations constrains the  
787 evolution of reproductive mode.

788

## 789 **9: Sex Reversal**

790 Sex determination and differentiation are two developmental processes that govern the  
791 dichotomous phenotype, male or female. This variation is constrained to produce either male  
792 or female phenotypes. A sex-determining factor initiates the sexual differentiation cascade,  
793 and results in individuals that possess either ovaries (female) or testes (male), as well as sex-  
794 specific behaviours, morphologies, and physiologies. Lizard sex determining factors could  
795 fall into diverse categories and we still have not identified a single sex determining gene for  
796 lizards. In many lizard species, sex chromosomes determine female or male development,  
797 and these genetic sex determination (GSD) systems can be either female heterogametic  
798 (ZZ/ZW) or male heterogametic (XX/XY) (Bull 1980). Temperature-dependent sex  
799 determination (TSD) is also common among lizards (Harlow 2004), whereby sex is  
800 determined by the temperature at which the embryo develops during a sensitive period of  
801 incubation. Although some lizard lineages possess stable and conserved sex chromosomes,  
802 others exhibit rapid sex chromosome evolution, including transitions between male and  
803 female heterogamety and turnover in the chromosome pair recruited to determine sex  
804 (Pennell *et al.* 2018; Sarre *et al.* 2011). Sex chromosomes are predicted to play a central role  
805 in lizard evolution, adaptation, and speciation; even though the same reproductive phenotypes  
806 are achieved (male or female), episodic turnover in sex chromosomes and sex determining  
807 systems has broad evolutionary consequences (Ezaz *et al.* 2009; Sarre *et al.* 2011). Many

808 transitions have occurred between TSD and GSD across the lizard phylogeny (Janzen and  
809 Krenz 2004) (see also Section 10).

810 Sex determining systems exist as a continuum of genetic and environmental influence, with  
811 many species likely to occupy an intermediate position on the continuum from TSD to GSD  
812 (Sarre *et al.* 2004). Incubation temperatures are suspected of being capable of overriding  
813 genotypic sex-determining signals in ZZ/ZW and XX/XY systems in some species (Holleley  
814 *et al.* 2015; Holleley *et al.* 2016; Quinn *et al.* 2007; Shine *et al.* 2002a) in both oviparous and  
815 viviparous lizards (Cornejo-Páramo *et al.* 2020a; Dissanayake *et al.* 2021; Holleley *et al.*  
816 2015; Holleley *et al.* 2016; Shine *et al.* 2002a; Wiggins *et al.* 2020). Discordance between  
817 genotypic and phenotypic sex is known as sex reversal (Weber and Capel 2018). Naturally  
818 occurring temperature-sex reversal has only been definitively identified in two vertebrates,  
819 both Australian lizards, which possess contrasting systems of GSD that simultaneously  
820 display temperature-dependent influences on sex. *Pogona vitticeps* is an agamid dragon lizard  
821 with a female heterogametic (ZZ/ZW) GSD system, but high incubation temperatures (>  
822 32°C) result in reversal of the ZZ male genotype to a female phenotype (Holleley *et al.* 2015;  
823 Quinn *et al.* 2007). In contrast, *Bassiana duperreyi* has a male heterogametic (XX/XY) GSD  
824 system in which low incubation temperatures (< 20°C) result in reversal of the XX genotype  
825 to a male phenotype (Dissanayake *et al.* 2021; Radder *et al.* 2008; Shine *et al.* 2002a).

826 The occurrence of sex reversal in wild populations of *P. vitticeps* and *B. duperreyi* provide  
827 unique opportunities to study the distribution and dynamics of sex reversal under different  
828 selective pressures in Australia (Figure 3). Theoretical studies show that as the frequency of  
829 sex reversal increases in a population, and provided the reversed individuals are fertile, a  
830 likely response is the reduction and possible elimination of the W or Y chromosome under  
831 Fisher's frequency-dependent selection (Düsing ; Fisher 1930) and a transition to a pure TSD

832 system (Bókonyi *et al.* 2017; Bull 1981; Dissanayake *et al.* 2021; Geffroy and Wedekind  
833 2020; Grossen *et al.* 2011; Holleley *et al.* 2015; Schwanz *et al.* 2020). Alternatively, selection  
834 for the rarer sex may drive the evolution of higher temperature thresholds for sex reversal and  
835 the maintenance of GSD in this species (Schwanz *et al.* 2020). Across the natural range of *P.*  
836 *vitticeps*, sex reversal is spatially constrained, displaying no association with latitude or other  
837 environmental variables, which is contrary to expectation based on laboratory observations  
838 (Castelli *et al.* 2021). In *B. duperreyi*, rates of sex reversal increase with elevation (from zero  
839 to 18.46 % of XX individuals manifesting as phenotypic males; Dissanayake *et al.* 2021).  
840 This observation suggests that *B. duperreyi* has not been subject to selective pressures for  
841 rapid evolution in the threshold for sex reversal, and thus populations at high elevations may  
842 be susceptible to loss of the Y chromosome (Dissanayake *et al.* 2021). Evolution of the  
843 thermal threshold for sex reversal is possible in both species, and there exists some  
844 evolutionary capacity to buffer or moderate the effects of extreme climates (Castelli *et al.*  
845 2021; Dissanayake *et al.* 2021).

846 Few species have been examined for instances of sex reversal in Australia or elsewhere [but  
847 see (Whiteley *et al.* 2021a; Wiggins *et al.* 2020)]. The widespread occurrence of  
848 homomorphic sex chromosomes in lizards means that instances of sex reversal are  
849 challenging to detect. New bioinformatics tools and methods make it possible to identify and  
850 characterize sex chromosomes, leading to unanswered questions in sex determination  
851 mechanisms in lizards (Cornejo-Páramo *et al.* 2020a; Dissanayake *et al.* 2020; Hill *et al.*  
852 2018; Palmer *et al.* 2019). Sex reversal, although not currently confirmed, may occur in  
853 several other Australian lizards; the southern water skink *Eulamprus heatwolei* (Cornejo-  
854 Páramo *et al.* 2020a), spotted snow skink, *Carinascincus ocellatus* (Cornejo-Páramo *et al.*  
855 2020a) (see Section 10), Jacky dragon *Amphibolurus muricatus* (Whiteley *et al.* 2021b), and  
856 several overseas species, including the common collared lizard (*Crotaphytus collaris*)

857 (Wiggins *et al.* 2020), the multi-ocellated racerunner (*Eremias multiocellata*) (Wang *et al.*  
858 2015), and the Japanese gecko (*Gekko japonicus*) (Tokunaga 1985).

859 So far, the molecular mechanisms of sex reversal in lizards are not fully understood (Castelli  
860 *et al.* 2021; Georges and Holleley 2018; Whiteley *et al.* 2021c). The tantalizing possibility  
861 that many traditionally regarded TSD species also have a cryptic underlying chromosomal  
862 component to sex determination is an important area of future research. Evolutionary  
863 modelling has shown that the Y or W chromosome can persist at low frequency despite high  
864 rates of sex reversal (Quinn *et al.* 2007; Schwanz *et al.* 2020). Beyond cryptic Y or W sex  
865 chromosomes, even more complex scenarios may exist that interact with thermal sex-  
866 determining cues, such as polygenic sex determination or *de novo* sex chromosomes (Mork *et*  
867 *al.* 2014). This is likely to be a very fruitful area of research using new genomic techniques.  
868 Finally, there are several evolutionary questions of interest concerning the transitions  
869 between GSD and TSD. For example, once the transition to TSD occurs, is there sufficient  
870 time for optimization of the thresholds for sex determination (pivotal temperature) to avoid  
871 catastrophic sex ratio skew and demographic extinction under climate change? There will no  
872 doubt be many more surprises. The recent media report of sex reversal in an adult Boyd's  
873 forest dragon (*Lophosaurus boydii*) in response to loss of the sole male in its captive  
874 population (Mannix 2020) is one such example. If confirmed, this will provide exciting new  
875 avenues of research and the interaction of cytogenetic and genetic aspects of developmental  
876 programming in lizards.

## 877 **10: Evolution of sex determination systems**

878 Sex determination is so fundamental to reproduction that it is expected to be under strong  
879 purifying selection with highly conserved processes (Uller *et al.* 2007). Indeed, conserved  
880 GSD systems exist in therian mammals (male heterogamety; XX/XY) and birds (female

881 heterogamety; ZZ/ZW)(Bachtrog *et al.* 2014; Vicoso *et al.* 2013). In contrast, reptilian sex  
882 determination mechanisms vary in a phylogenetically complex manner, which suggests  
883 multiple evolutionary transitions (Janzen and Phillips 2006; Pokorna and Kratochvil 2009). In  
884 lizards, there is GSD (both XX/XY and ZZ/ZW are both common), TSD, and interactions  
885 between genes and temperature (GSD+EE or thermosensitive GSD) (Sarre *et al.* 2004). Sex  
886 chromosomes range from highly differentiated to morphologically indistinct pairs in lizards  
887 with GSD. Sex determination in lizards may be dosage dependent, where the homogametic  
888 sex (ZZ or XX) possesses two copies of a sex determining gene and acquires a threshold for  
889 sexual phenotype, whilst the heterogametic sex (XY or ZW) possesses one copy and does not  
890 reach the threshold (Quinn *et al.* 2007). In addition, patterns of TSD are diverse: high  
891 temperatures can produce excess males ('FM' pattern), excess females ('MF' pattern) or an  
892 excess of females at both high and low temperatures with males produced at intermediate  
893 temperatures ('FMF' pattern; Figure 4). Finally, reaction norms to temperature can be steep,  
894 where the switch between male and female producing temperatures can occur over a small  
895 pivotal range (Figure 4a-c), or shallow (Figure 4d), where sex ratios vary gradually over a  
896 broad range of temperatures.

897 Work on Australian lizards has led to the important re-defining of sex determination as  
898 existing as a continuum of states between pure GSD and pure TSD, rather than a  
899 dichotomous trait (Sarre *et al.* 2004). However, questions remain about how and why  
900 transitions between systems of sex determination occur. The drivers and mechanisms acting  
901 at the time of transitions are difficult to determine due to the divergence between lizard  
902 groups with different sex determining systems. Australian lizards are central to understanding  
903 the evolutionary drivers of transitions, the mechanisms that underpin transitions and their  
904 consequences to species and populations. The adaptive benefit of GSD versus TSD has been  
905 the subject of extensive research in Australia (Pen *et al.* 2010; Shine 1999; Shine *et al.* 1995;

906 Wapstra and Warner 2010; Warner and Shine 2005; Warner and Shine 2008). TSD should be  
907 favoured when there are sex-specific advantages resulting from development at temperatures  
908 that produce an excess of the sex that benefits (Charnov and Bull 1977; but see Uller and  
909 Olsson 2006). GSD typically produces 50:50 sex ratios and is therefore favoured when  
910 seasonal temperature fluctuations are high and might otherwise produce sex ratio biases that  
911 are maladaptive for some individuals (Bulmer and Bull 1982; but see Cornejo-Páramo *et al.*  
912 2020b; Dooren and Leimar 2003). Species with TSD may be particularly vulnerable to  
913 climate change because of the effect that highly-skewed sex ratios can have on population  
914 persistence if one sex becomes rare to the point of limiting mating frequency (Boyle *et al.*  
915 2014; Le Galliard *et al.* 2005; Valenzuela *et al.* 2019; Wedekind 2017). While selection can  
916 favour biased sex ratios when sex-specific fitness benefits occur with incubation temperature  
917 (Charnov and Bull 1977), populations with sex ratios that are consistently biased towards one  
918 sex can experience deterioration of genetic diversity and therefore have reduced adaptive  
919 potential, with consequences for species distributions (Le Galliard *et al.* 2005; Mitchell and  
920 Janzen 2010; Mitchell *et al.* 2008). However, such population-wide skews are rare and often  
921 transient, because frequency-dependent selection favours females (or parents) producing the  
922 under-represented sex (Fisher 1958), which concomitantly can select for mechanisms to  
923 balance skews (Uller *et al.* 2007).

924 In reptiles, TSD was first discovered in *Agama agama* more than 50 years ago (Charnier  
925 1966; Steele *et al.* 2018), followed by observations in turtles (Pieau 1972; Yntema 1976), the  
926 leopard gecko (Viets *et al.* 1993), and the tuatara (Cree *et al.* 1995), all of which are  
927 oviparous. Because viviparous females can behaviourally buffer the effects of temperature on  
928 offspring development, TSD was considered incompatible with viviparity (Bull 1980), but  
929 temperature effects on offspring sex have since been demonstrated in two viviparous  
930 Australian lizards (Robert and Thompson 2001; Wapstra *et al.* 2004). These viviparous

931 species differ from typical TSD species because they have distinctly shallow reaction norms  
932 best characterised by Figure 4d. Australia has considerable lizard diversity, scientific  
933 expertise, and exceptional species amenable to advancing our understanding of evolutionary  
934 transitions in sex determination. One such species (*C. ocellatus*) is undergoing an incipient  
935 transition in sex determination, offering a rare opportunity to unravel the causes and  
936 consequences of such transitions.

937 *An Australian model system for transitions in sex determination: Carinascincus ocellatus*

938 The viviparous spotted snow skink, *C. ocellatus* (Figure 5a) is a rare example of a viviparous  
939 species undergoing incipient divergence in sex determination (Cunningham *et al.* 2017; Pen  
940 *et al.* 2010; Wapstra *et al.* 2004). This small Tasmanian-endemic skink has a broad  
941 geographic and climatic distribution across Tasmania, with concomitant variation in life  
942 history (Wapstra and Swain 2001; Wapstra *et al.* 1999; Wapstra *et al.* 2001). In a warm, low  
943 elevation population, maternal basking opportunity (and therefore developmental  
944 temperature) influences the sex of her offspring; warm environmental conditions lead to an  
945 excess of female offspring in the population and cool conditions lead to an excess of males.  
946 In a high elevation population, sex ratios remain at parity regardless of environmental  
947 conditions and maternal basking opportunity (Cunningham *et al.* 2017; Pen *et al.* 2010)  
948 (Figure 5b,c). The sex ratio response to temperature, initially observed in the laboratory in  
949 females from the low elevation population through altered basking opportunity (Wapstra *et*  
950 *al.* 2004), has since been demonstrated in the wild (Wapstra *et al.* 2009). There are  
951 potentially important consequences for this population-specific effect of environmental  
952 temperatures on offspring sex ratios (5c) and for our understanding of the impact of climate  
953 change on species with TSD (especially where they follow a relatively rare shallow reaction  
954 norm) (Cunningham *et al.* 2020). Because warm temperatures produce a female-biased sex

955 ratio in *C. ocellatus*, climate warming may result in population growth rather than decline as  
956 temperatures warm, as long as males do not become limiting (Cunningham *et al.* 2017;  
957 Rankin and Kokko 2007; Wapstra *et al.* 2009).

958 In *C. ocellatus*, divergence in sex determination could be driven by a combination of  
959 population-specific selection on how mothers allocate the sexes of their offspring (resulting  
960 in thermosensitive sex determination at low elevation) along with selection against strongly  
961 biased sex ratios in variable, unpredictable environments [resulting in GSD at high elevation  
962 (Pen *et al.* 2010), Figure 5d]. Specifically, in warm lowland populations, individual females  
963 that produce daughters under warmer conditions potentially have higher fitness because  
964 female offspring born early reach sexual maturity earlier and have a higher lifetime  
965 reproductive output than those born late. This situation selects for a link between gestational  
966 temperature, birth date, and offspring sex. Conversely in the cold highlands, selection does  
967 not occur because cooler conditions lead to slower growth with later sexual maturity, which  
968 creates a dissociation between temperature-dependent development, birth date, size at  
969 maturity, and sex-specific fitness benefits to mothers from sons or daughters. The high annual  
970 fluctuation in temperature in the cold highlands further selects against temperature effects in  
971 this model, because of the resultant frequency-dependent selection against the large skews in  
972 sex ratios (Figure 5d).

973 The theoretical model (Figure 5) established a potential adaptive explanation for the intra-  
974 specific variation in sex determination in *C. ocellatus*, but its mechanistic predictions  
975 (emergence/loss of genes of major effect at high/low elevation) require testing. Sex  
976 chromosomes and sex-linked DNA in *C. ocellatus* differ only slightly between the  
977 populations (Hill *et al.* 2021a; Hill *et al.* 2018) and these populations diverged during the  
978 Pleistocene with negligible gene flow since divergence (Hill *et al.* 2021b). Specifically, and

979 in contrast to some of the model predictions, both populations have XY heterogamety with  
980 morphologically undifferentiated X and Y sex chromosomes (Hill *et al.* 2021a) (Figure 5e),  
981 and both populations largely share sex-linked genetic sequences (Hill *et al.* 2018). In  
982 addition, each population also possesses unique sex-linked genetic sequences. The *C.*  
983 *ocellatus* system highlights that only minor changes to the sex-linked genome are required for  
984 transitions in sex determination to occur: the high elevation population has more repeat and  
985 heterochromatin accumulation on the Y chromosome than the low elevation population (Hill  
986 *et al.* 2021a) and recombination among the sex-linked loci shared among populations is more  
987 suppressed in the high elevation population (Hill *et al.* 2018). This molecular work  
988 demonstrates that rather than loss/gain of genes of major effect as predicted, population-  
989 specific sex ratio responses to temperature are underpinned by sex-linked genes important for  
990 sex determination in both populations. However, the theoretical model (Figure 5) is useful  
991 because it establishes testable hypotheses for how sex determination evolves in viviparous  
992 lizards, and could easily be applied to other Australian viviparous species that exhibit  
993 temperature effects on offspring sex, like *Eulamprus tympanum* (Robert and Thompson  
994 2001). Indeed, the model predictions are consistent for taxa starting with either XY or ZW  
995 chromosomal sex determination. The diversity of viviparous lizards in Australia may provide  
996 several useful models for further research if temperature is found to affect offspring sex in  
997 additional species.

## 998 **11: Control over timing of birth**

999 Most viviparous animals give birth to their offspring synchronously (within minutes or  
1000 hours), although some lizard species complete the act of birth over several days. This  
1001 phenomenon has been termed ‘birthing asynchrony’ and is defined as a spread in births  
1002 separated by a minimum of 12 hours [i.e., not within the same day; (While *et al.* 2007a)].

1003 Birthing asynchrony is analogous to hatching asynchrony in birds, whereby the eggs of a  
1004 clutch are laid and hatch over several days (Amundsen and Slagsvold 1996; Magrath 1990;  
1005 Stenning 1996; Stoleson and Beissinger 1995). However, unlike hatching asynchrony,  
1006 birthing asynchrony appears not to result from developmental asynchrony - birds are  
1007 constrained to produce one embryo at a time and thus, to lay one egg at a time. In contrast,  
1008 lizards that exhibit birthing asynchrony retain fully-developed offspring and give birth to  
1009 them one at a time (While *et al.* 2007b), even when multiple offspring are present within the  
1010 same uterus. This reproductive strategy occurs in multiple species within the Egeriinae, as  
1011 well as four species within the African lizard family Cordylidae (Table 3).

1012 The presence of birthing asynchrony raises several interesting questions relating to the  
1013 proximate and ultimate mechanisms that facilitate this behaviour. Most work to date has  
1014 focussed on the latter. Analogous hatching patterns in birds may be a mechanism that allows  
1015 parents to mediate intra-brood conflict during times of limited resource availability (Lack  
1016 1947; Stienen and Brenninkmeijer 2006). In Egeriinae, birthing asynchrony may provide a  
1017 similar advantage by influencing the competitive environment of the brood and thus  
1018 mediating offspring growth, survival or dispersal (While and Wapstra 2009). Like hatching  
1019 asynchrony in birds, birthing asynchrony results in a size hierarchy in a litter. While this size  
1020 hierarchy does not impact the competitive ability of each offspring, it does alter the level of  
1021 competition present within a single litter (While *et al.* 2009a; While and Wapstra 2009).

1022 Social Egeriinae often live in saturated habitats that include intense competition for  
1023 resources, high levels of conspecific aggression and high juvenile mortality (Chapple 2003).  
1024 Birthing asynchrony may operate as a trade-off between offspring mortality/dispersal and  
1025 offspring mass/growth as a result of size hierarchy (While and Wapstra 2009). Indeed, when  
1026 offspring are kept together in captivity, smaller individuals, which are frequently attacked by  
1027 conspecifics, grow at a slower rate and behavioural development differs between dominant

1028 and submissive individuals (Riley *et al.* 2017). In the wild, such behaviour may facilitate the  
1029 dispersal of the subdominant individual, allowing parents to modify the number of offspring  
1030 they tolerate in their home range during periods of resource limitation. Interestingly, the only  
1031 other family of lizards in which birthing asynchrony has been documented, the African  
1032 Cordylidae, also include group-living species (Mouton 2011). The co-occurrence of these  
1033 traits across potentially two independent transitions to simple family life makes the links  
1034 between the evolution of complex sociality and that of birthing asynchrony hard to ignore.  
1035 However, the functional consequences of birthing asynchrony and the size hierarchies it  
1036 produces remain to be investigated.

1037 While considerable work has attempted to understand the function of birthing asynchrony, we  
1038 know almost nothing about how it is achieved mechanistically in either lizard clade. The  
1039 answer is likely to lie in the co-option of the mechanisms that underpin the birthing process  
1040 itself. These mechanisms span hormonal, neuronal, and embryonic effects, all of which  
1041 interact to initiate and maintain the process of parturition. Lizards with asynchronous birth  
1042 may have co-opted these mechanisms to finely control when birth is initiated for each  
1043 individual offspring while simultaneously preventing the birth of the remainder of the litter.  
1044 For example, hormones actively influence the parturition process within the mother (Chaim  
1045 and Mazor 1998) by way of their presence in the bloodstream and through changes in  
1046 receptor expression in uterine tissue (Blanks and Thornton 2003). Additionally, dynamic  
1047 changes in uterine innervation and neuronal receptor expression could facilitate the isolation  
1048 of a single embryo within the uteri of a female. This type of regionally-specific neuronal  
1049 fluctuation is readily evident in the mammalian cervix at term pregnancy (Boyd *et al.* 2009;  
1050 Chávez-Genaro *et al.* 2006) and exemplifies the capacity for the uterine environment to  
1051 change rapidly and with precision. These mechanisms are further impacted by the presence of  
1052 the developing embryos, and maternal-foetal hormone signalling provides an avenue for each

1053 embryo to influence the timing of its own birth (Challis *et al.* 2000; Liggins 2000). While all  
1054 the above mechanisms present intriguing targets for co-option during the evolutionary  
1055 refinement of live birth, none yet have come under empirical scrutiny.

1056 The integration of physiological, histological, and molecular techniques provides an  
1057 opportunity to tease apart these competing mechanisms and generate a holistic understanding  
1058 of the parturition process, and, more explicitly, how timing of birth is controlled by mothers  
1059 and/or offspring. Specifically, the use of contraction bioassays to measure uterine contractile  
1060 responses to key hormones (Paul *et al.* 2020), in combination with measuring changes in  
1061 neuronal density across regions of the uterus to gauge the capacity for uterine relaxation,  
1062 have the potential to demonstrate how this behaviour is achieved. Furthermore, combining  
1063 contractile assays with transcriptomic data will allow identification of the regions of the  
1064 genome that are associated with the contractile and relaxation responses. Such information  
1065 should be compared across clades to examine whether similar mechanisms have been co-  
1066 opted in independent lineages in a convergent pattern. Combined, this will enhance our  
1067 understanding of how systemic reproductive innovations emerge, and will provide  
1068 fundamental knowledge of how the process of live birth itself evolved. These results would  
1069 have implications for a broad range of disciplines including evolutionary biology,  
1070 conservation biology, and even human health.

## 1071 **Conclusions**

1072 We have identified a number of areas where research on Australian lizards has provided key  
1073 advances in our understanding of animal reproduction, and areas where their potential as  
1074 models for research has been under-utilised. Australian lizards are particularly important  
1075 models for studying the origins and mechanisms underlying sociality and mating systems.  
1076 They exhibit a continuum of social behaviours, from largely solitary to highly social, which

1077 provides a framework for understanding how sociality first evolves. Lizards communicate  
1078 using chemical and/or visual signals, and these signals correlate with their mating behaviours.  
1079 Thus, how their signals and mating behaviours coevolve is a potentially rich field for future  
1080 research, especially given the diversity Australian lizards exhibit between communal sociality  
1081 and solitary life. Furthermore, their mating behaviours feed into a complex arena of sexual  
1082 conflict, where males and females both exhibit potential adaptations to increase their own  
1083 fitness at the expense of their mates'. The fitness consequences of mating systems, as well as  
1084 post-mating mechanisms like cryptic female choice and sperm competition, thus provide a  
1085 basis for understanding how mechanisms of communication might provide a starting point for  
1086 more complex social behaviours to evolve.

1087

1088 Australian lizards are also important models for post-fertilisation components of  
1089 reproduction, especially the evolution of nesting decisions, viviparity, placentation, sex  
1090 determination mechanisms, and asynchronous birth. Each of these traits has important  
1091 consequences for how animal species survive in the face of environmental change. Our  
1092 review shows how nesting decisions, TSD, and viviparity allow females to manipulate and  
1093 potentially optimize their offspring fitness in response to environmental conditions like  
1094 temperature, moisture, and population sex ratio. Placentation may provide a mechanism for  
1095 females to maintain fetal survival in response to changes in food abundance during  
1096 reproductive events. Birthing asynchrony allows the timing of parturition to be modified,  
1097 possibly in response to social or environmental cues, so that all offspring are not born into the  
1098 same environment.

1099

1100 Although we have covered many key topics, many questions and areas of investigation where  
1101 Australian lizards would provide excellent models for study remain. We have highlighted a

1102 list of outstanding key questions that were illuminated by our review in Table 4. It is not an  
1103 exhaustive list, but we highlight questions that emerged after considering the review of all  
1104 our topics. For example, communication, sociality, and environment-dependent variation (or  
1105 fitness) are key topics that appear repeatedly in our review, and have major consequences for  
1106 several areas of investigation. Our review demonstrates the utility of Australian lizards for  
1107 addressing these questions.

1108 In addition to the questions that emerged from our review, we also catalogued several topics  
1109 that our review did not cover (Table 5). These topics were neglected by necessity rather than  
1110 by choice, because no authors responded to our initial invitation to the Australian Society of  
1111 Herpetologists listserv to cover these topics. There are certainly active research projects  
1112 focusing on these topics that we did not review sufficiently here as a result. However, the  
1113 lack of response for these topics may indicate that these fields are areas for which lizards are  
1114 relatively under-utilised. We have suggested possible outstanding questions for which lizards  
1115 would be ideal models to address.

1116 Our reviews clearly highlights that the Australian dragons (Agamidae) and skinks (Scincidae)  
1117 have contributed to understanding broad biological questions, in part because of their large  
1118 number of species, and their special features such as sexual colour dimorphisms and  
1119 reproductive modes. By contrast, we have highlighted only one study on goannas  
1120 (Varanidae), and none on any of the gekkotans (Carphodactylidae, Diplodactylidae,  
1121 Gekkonidae, Pygopodidae). Despite this fact, it seems unlikely that the latter families do not  
1122 offer potential opportunities for significant research on broad reproductive questions. For  
1123 example, the gekkonids are unusual for laying calcareous-shelled eggs, which provides a  
1124 good comparison with the flexible-shelled eggs of species in the other gekkotan families,  
1125 with implications for embryonic physiology and development (Andrews et al, 2013). Apart

1126 from the research on deep-nesting varanid lizards (Section 7), little is known about nests and  
1127 nesting (nest sites, nest structure and environmental features, especially temperatures and  
1128 water potentials) in most Australian lizards. Nest sites, nest structure, and environmental  
1129 features, especially temperatures and water potentials, remain unknown for most species.  
1130 Given the range of potential nest environments in Australia, from extreme aridity to moist  
1131 temperate regions, investigations of nest and nesting behaviours, and their impact on  
1132 embryonic physiology, are likely to be informative for development of the understanding of  
1133 the evolution of reproductive strategies (viviparity, sex determination, incubation periods,  
1134 rates of development and so on), and for predicting future effects of climate change.  
1135 Gekkotans will be equally as important in these investigations as are dragons and skinks.

1136 In summary, Australian lizards are speciose and diverse, providing opportunities for exciting  
1137 research in reproductive biology. We have highlighted key areas of research that are ongoing,  
1138 emerging, and relatively neglected. We hope that new generations of reproductive biologists  
1139 and ecologists will be inspired by our review to consider these topics for their own research  
1140 careers. To encourage students who are excited by the topics in our review, we have provided  
1141 a list of the authors responsible for each section in our supplementary materials  
1142 (Supplementary Table 1). We urge readers to contact the relevant authors for research  
1143 opportunities in the topics that they find interesting. Australian lizards offer excellent  
1144 opportunities to test important hypotheses in vertebrate reproductive biology. Their  
1145 abundance and the diversity of environments they inhabit provide an important resource for  
1146 ongoing ecological and evolutionary research.

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1153 The authors declare no conflicts of interest.

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2205 *V. indicus* and the *V. prasinus* species groups. *Zootaxa* **1472**(1), 1.

2206 Zozaya, S.M., Higgie, M., Moritz, C., and Hoskin, C.J. (2019) Are pheromones key to  
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2208

2209 **Figure Titles**

2210 **Figure 1.** Species richness map for Australian lizards. Colour coding represents the number  
2211 of species in a 50 km x 50 km pixel, with cooler colours indicating fewer species (down to 4)  
2212 and warmer colours indicating more species (up to 88; Reproduced from Cogger 2018 with  
2213 permission from CSIRO Publishing).

2214 **Figure 2.** Gould's monitor lizard (*V. gouldii*) communal nesting warren before excavation.  
2215 Note the multiple entrances (photograph by Stephen Zozaya).

2216 **Figure 3.** Sex reversal in the Australian lizard species *Bassiana duperreyi* and *Pogona*  
2217 *vitticeps*. (A) Distribution of sex reversal across the ranges of *P. vitticeps* (Castelli et al.  
2218 2021) and *B. duperreyi* (Dissanayake et al. 2021). The species range is indicated by the  
2219 shaded areas. (B) Graphical representation of sex reversal characteristics (not to scale); top –  
2220 *P. vitticeps*: sex reversal occurs when an individual with a male genotype (ZZ) is incubated at  
2221 temperatures above 32 °C, causing it to develop as a phenotypic female (ZZf) (Holleley et al.  
2222 2015); bottom – *B. duperreyi*: an individual with a female phenotype (XX) incubated at low  
2223 temperatures will reverse its sex and develop as a phenotypic male (XXm) (Shine et al. 2002;  
2224 Radder et al. 2008). (C) Sex reversal frequency of *B. duperreyi* along an elevational gradient  
2225 (Dissanayake et al. 2021). Underlying map generated using ArcGIS 10.5.1  
2226 (<http://www.esri.com>) and data from the Digital Elevation Model (Geoscience Australia)  
2227 made available under Creative Commons Attribution 3.0 Australia  
2228 (<https://creativecommons.org/licenses/by/3.0/au/legalcode>, last accessed 21-Dec-20). The  
2229 species range is indicated by the shaded areas (Data from the Atlas of Living Australia  
2230 website at <http://www.ala.org.au>. Accessed 3 January 2021).  
2231 (<https://creativecommons.org/licenses/by/3.0/au/legalcode>, last accessed 21-Dec-20).

2232 **Figure 4.** Patterns of temperature dependent sex determination (TSD). FM pattern (a,d)  
2233 where males are produced at high temperatures; MF pattern (b) where females are produced  
2234 at high temperatures; or FMF pattern (c) where males are produced at an intermediate  
2235 temperature and females are produced either side of this temperature. Reaction norms can be  
2236 steep (a-c) or shallow (d).

2237 **Figure 5.** *Carinascincus ocellatus* (a) high (1200 metres above sea level (masl), GSD, blue)  
2238 and low (50 masl, GSD+EE, red) elevation populations (b) with population specific sex-ratio  
2239 responses to incubation temperature (c). A life history model parameterised with long-term  
2240 field data predicts loss of sex determining genes at low elevation and emergence of sex  
2241 determining genes at high elevation (adapted from Pen et al. 2010) (d). Sex chromosomes (e),  
2242 labelled here with *C. ocellatus*-specific Y-linked probe (Hill, et al. 2021a), are  
2243 undifferentiated in both populations.