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## Serum uric acid plays a protective role for bone loss in peri- and postmenopausal women: A longitudinal study<sup>☆</sup>

Joanna Makovey<sup>a,\*</sup>, Monique Macara<sup>a</sup>, Jian Sheng Chen<sup>a</sup>, Christopher S. Hayward<sup>b</sup>, Lyn March<sup>a</sup>, Markus J. Seibel<sup>c</sup>, Philip N. Sambrook<sup>a</sup>

<sup>a</sup> Institute of Bone and Joint Research, Kolling Institute, Royal North Shore Hospital, University of Sydney, Sydney, Australia

<sup>b</sup> Department of Cardiology, St Vincent's Hospital, Sydney, Australia, Victor Chang Cardiac Research Institute, University of New South Wales, Sydney Australia

<sup>c</sup> Bone Research Program, ANZAC Research Institute, The University of Sydney at Concord Campus, Sydney, Australia

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## ABSTRACT

**Objective:** Oxidative stress has been linked to osteoporosis. Serum uric acid (UA), a strong endogenous anti-oxidant, has been associated with higher bone mineral density (BMD), lower bone turnover and lower prevalence of fractures in a large cross-sectional study of men. Whether this relationship is present in women and how UA relates to changes in BMD longitudinally has not been examined.

**Methods:** A sample of 356 peri- and postmenopausal women, mean age 60.5 years was studied. Each individual had baseline BMD and body composition measurements by dual energy x-ray absorptiometry (DXA) and at least one repeat measure, on average 9.7 years later. Annual rate of change in BMD (A%ΔBMD) was calculated. UA was measured at each DXA visit. Calcitropic hormones and bone turnover markers were measured at the final visit only.

**Results:** Cross-sectional data analyses revealed that women with higher UA levels had significantly higher absolute BMD measures at all skeletal sites. These women also had higher measures of body weight and its components such as lean mass (LM) and fat mass (FM). Results of multiple regression analyses showed a positive association between UA and BMD that remained significant even after accounting for possible confounders including LM and FM. Regression analyses of the longitudinal BMD data demonstrated significant associations between serum UA levels and annual rates of change in BMD at all skeletal sites. After adjustment associations remained significant for lumbar spine, forearm and whole body BMD but not for hip BMD.

**Conclusion:** Higher serum UA levels appear to be protective for bone loss in peri- and postmenopausal women and this relationship is not affected by changes in body composition measures.

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## Introduction

Soluble uric acid (UA) is present principally as monosodium urate at physiological pH values and is the final breakdown product of purine metabolism. Historically, UA has been viewed as a waste byproduct, which in excess may cause gouty arthritis and renal stones [1,2]. While it is well recognised that UA in its crystalline state is pro-inflammatory [3], there has been controversy as to the biological roles of soluble UA. Although soluble UA was considered biologically relatively inert, it is now thought that higher serum UA levels within normal physiologic levels (0.15–0.4 mmol/L) [4] may have conferred a selection advantage because of their antioxidant effects [3,5–7].

Indeed, UA accounts for approximately half of the antioxidant properties of human plasma [3]. Evidence from observational and

epidemiological studies has linked oxidative stress or low circulating levels of anti-oxidants to reduced bone mineral density (BMD) and osteoporosis [8–11]. On the other hand, increased body weight has been reported as one of the major predictors of elevated levels of serum UA [3,12]. Supranormal serum UA levels (hyperuricaemia) have been associated with presence of the metabolic syndrome [7,12–15] and its components such as diabetes mellitus [16,17], obesity [18–20], hyperlipidemia [21–23] and hypertension [19,24]. Body weight has been related to BMD [25–27]. Numerous previous studies have also reported positive associations between body composition components such as lean body mass and fat body mass and BMD at different skeletal sites [27–30].

In a large population-based study of older men (the CHAMP Study), the CHAMP collaborative recently reported that higher serum UA levels were significantly associated with higher BMD at various skeletal sites after adjusting for covariates [31]. Moreover, higher serum UA levels were associated with a lower prevalence of osteoporosis as determined by either BMD or prevalent non-vertebral fracture status. Whether this relationship is present in women and how UA relates to changes in BMD longitudinally has not been examined.

<sup>☆</sup> All authors state that they have no conflicts of interest.

\* Corresponding author at: Department of Rheumatology, Royal North Shore Hospital, Building 35, Level 4, St Leonards, NSW, 2065, Australia. Fax: +61 2 99061859.

E-mail address: [jmakovey@med.usyd.edu.au](mailto:jmakovey@med.usyd.edu.au) (J. Makovey).

## 82 Methods

### 83 Subjects

84 Study subjects were female twins over 45 years, recruited as part  
85 of the Northern Sydney Twin Study, which has been running at the  
86 Department of Rheumatology, Royal North Shore Hospital, since  
87 1996. The twins were recruited through the Australian National  
88 Health and Medical Research Council (NHMRC) Twin Registry and  
89 from local media campaigns. Twins were invited to participate in an  
90 investigation into the genetic and environmental determinants of  
91 various diseases including osteoarthritis, cardiovascular disease, asthma,  
92 and osteoporosis on several occasions. The hospital's Human  
93 Research Ethics Committee approved the study. After providing written  
94 informed consent, each twin was interviewed separately in accordance  
95 with a standard questionnaire to collect demographic, lifestyle and  
96 medical history data. The baseline visit was completed by 1980  
97 twins (1997–2006), and 864 of these participants attended at least  
98 one follow-up visit (2009–2010).

99 Except for hormone therapy, twins who used medications or who  
100 had medical conditions that could interfere with bone metabolism  
101 were excluded from the analysis. Individuals with conditions that  
102 might compromise the accuracy of DXA measurements such as severe  
103 obesity, the presence of artificial objects such as pacemaker or gallstones,  
104 or significant degenerative spine changes were also excluded.  
105 Hormone therapy use was recorded and included as a covariate in the  
106 statistical analyses. Zygosity in same-sex twins was determined from  
107 the twins' self-report using questions from a validated questionnaire  
108 [32]. DNA fingerprinting was used to determine zygosity in twin pairs  
109 in which their zygosity was either unknown or disputed.

### 110 Baseline characteristics and laboratory measurements

111 Demographic characteristics of the study cohort included age  
112 (years), height (m), weight (kg), BMI ( $\text{kg}/\text{m}^2$ ), menopausal status  
113 (MS), hormone replacement therapy (HRT), physical activity (PA),  
114 alcohol intake and smoking history. Menopausal status was  
115 categorised as 1 – premenopausal (i.e. having regular menstrual  
116 cycles), 2 – perimenopausal (i.e. experiencing changes in frequency  
117 of their menses or amenorrhoea of at least 3 but less than 12 months)  
118 and 3 – postmenopausal (amenorrhoea for 12 consecutive months).  
119 Hormone replacement therapy was recorded and accounted for if  
120 taken regularly for more than 3 months within the last 12 months. PA  
121 was categorised based on time spent on leisure exercise for >  
122 30 minutes per day (0 – none, 1 – <30 min/day, 2 –  $\geq 30$  min/day).  
123 Alcohol intake was recorded as standard drinks per week and  
124 categorised as 0 – none; 1 –  $\leq 1$  drink per week (social occasions  
125 only); 2 – 2–13 drinks per week (moderate) and 3 –  $\geq 14$  drinks  
126 per week (excessive). Smoking habits were recorded as 0 – never;  
127 1 – current smoker; 2 – ex-smoker (not smoked in the last 3 months).  
128 Self-reported fractures that occurred between baseline and the final  
129 visits of the study were also recorded.

130 Fasting blood samples used in this study were collected at each  
131 subject's visit and kept as aliquots at  $-80^\circ\text{C}$  until analysis. Serum  
132 UA was measured from baseline and last visit blood samples. Other  
133 biochemical parameters such as creatinine, calcium, albumin and  
134 phosphorus and bone markers were measured from the last visit  
135 samples only. These tests were performed using standard techniques  
136 on a Roche Modular Analytics <P> module (Roche Diagnostics,  
137 Germany). The UA assay had a detection limit of 0.01 mmol/L, female  
138 reference range of 0.18–0.38 mmol/L and combined measurement  
139 of uncertainty of 1.1% at 0.18 and 0.44 mmol/L. Serum calcium was  
140 measured by colorimetric assay using p-cresolphthalein. Values  
141 were adjusted for circulating albumin levels with a reference  
142 range of 2.15–2.5 mmol/L. Glomerular filtration rate (GFR) was calculated  
143 using the Cockcroft–Gault formula [33,34]. Serum levels of

144 aminoterminal procollagen type I propeptide (PINP) were deter-  
145 mined by Electrochemiluminescence immunoassay on a Roche Mod-  
146 ular Analytics E170 module (Roche Diagnostics GmbH, Germany).  
147 The assay for serum PINP, a marker of bone formation, detects both  
148 trimeric and monomeric fractions of PINP. The detection limit was  
149 5 ng/mL with total precision coefficients of variation (CVs) of be-  
150 tween 3.8% and 4.2%. Serum concentrations of the aminoterminal  
151 cross-linked telopeptide of collagen type I (Serum CTX-I) were mea-  
152 sured using a manual immunoassay (Osteomark, Ostex, USA).

### Bone Mineral Density and Body Composition Measurements

153  
154 Lumbar spine (LS), total hip, forearm and whole body scans were  
155 performed on a fan beam dual-energy X-ray absorptiometry (DXA)  
156 bone densitometer (QDR 4500W, Hologic, Waltham, MA USA) at  
157 baseline and follow-up visits. Measurements of bone mineral density  
158 (BMD) ( $\text{g}/\text{cm}^2$ ) and body composition such as fat mass (FM) (kg) and  
159 lean body mass (LM) (kg) were obtained using standard protocols as  
160 previously described [29,35]. The same densitometer was used  
161 throughout the entire study. Performance of the DXA scanner has  
162 been monitored throughout the study. Routine daily QC scans of the  
163 Spine Phantom were performed and the coefficient of variation for  
164 QC BMD measures in our unit was 0.98%. In vivo reproducibility has  
165 been estimated from duplicate scans (155 patients with repositioning  
166 between scans) as coefficients of variation (CV) and intraclass correlation  
167 (ICC) for BMD and body composition measures. CV and ICC  
168 for LS, total hip, femoral neck BMD were 0.74/0.998; 1.23/0.994  
169 and 1.27/0.994 correspondingly. CV and ICC for Total LM were 1.07/  
170 0.997 and for Total FM – 1.83/0.997.

171 Baseline and last visit measurements were used to calculate an  
172 annual rate of change in BMD. Commonly accepted annual % change  
173 in BMD (%/year  $\Delta\text{BMD}$ ) was selected as a longitudinal BMD measure  
174 to adjust for difference in time between two end-point visits in  
175 study participants [36–41].

### Statistical analysis

176  
177 For comparison between groups of UA tertiles, ANOVA analysis for  
178 continuous variables and chi-square tests for categorical variables  
179 was performed. Adjusted means across tertiles of uric acid were also  
180 reported for bone-related and body composition measures at the  
181 final visit. In addition, generalised linear regression models were  
182 used to assess the association between UA and BMD at the final visit  
183 or annual rate of change in BMD ( $\text{A}\%\Delta\text{BMD}$ ). Lack of independence  
184 of BMD measures between dizygotic (DZ) pairs was taken into  
185 account using generalised estimating equations. The annual rate  
186 of BMD change ( $\text{A}\%\Delta\text{BMD}$ ) was calculated as  $100 \times [\text{BMD at final}$   
187  $\text{visit} - \text{BMD at baseline}] / \text{BMD at baseline} / \text{time interval between}$   
188  $\text{the two measurements}$ , and was used to account for differences in  
189 the intervals among the study participants. The selection of this  
190 common parameter as the outcome variable for the longitudinal  
191 data was due to the fact that the vast majority of the participants  
192 had only two measurements.

193 In multivariate regression analysis, BMD or  $\text{A}\%\Delta\text{BMD}$  were treated  
194 as dependent variables, and log UA or  $\text{A}\%\Delta\text{UA}$  as independent variables.  
195 Models were adjusted for known and potential confounders,  
196 including GFR, serum calcium and CTX-I levels, age, history of  
197 smoking, alcohol intake, HRT use and physical activity. We did not include  
198 weight or BMI in models because weight is made up of BMC, lean  
199 mass and fat mass. We included both lean mass and fat mass  
200 and height as a correction for body size in final models. Regression  
201 analyses for relationships between UA and body composition mea-  
202 sures were done in a similar manner by treating one of the body  
203 composition measures as dependent variable in the regression models.  
204 Longitudinal data was also analysed by time dependent mixed regression  
205 models.

## 206 Results

207 There were 460 female twin participants who had completed their  
 208 last follow-up visit: 96 monozygotic (MZ) and 134 dizygotic (DZ)  
 209 pairs. After randomly excluding one member of each MZ twin pair  
 210 ( $n=96$ ) and subjects taking allopurinol ( $n=4$ ), thiazides ( $n=2$ ) or  
 211 loop diuretics ( $n=2$ ), 356 women with a mean age of 60.4 (range  
 212 45–83) years remained for analysis.

## 213 Cross-sectional analyses

214 The main anthropometric, biochemical and lifestyle characteristics  
 215 of these subjects at the final visit, stratified by tertiles of UA levels are  
 216 presented in Table 1. There were 26 hyperuricemic women (serum  
 217 UA levels  $\geq 0.41$  mmol/L) in the highest tertile of UA. As expected  
 218 final visit UA levels were higher than baseline levels. Women with  
 219 higher serum UA levels were older, heavier and correspondingly  
 220 had higher BMI than those with lower UA concentrations. Total  
 221 serum cholesterol, LDLC, triglyceride, calcium and creatinine levels  
 222 were all significantly higher in the higher UA tertiles. Serum CTX-I  
 223 levels were higher in the highest UA tertile, but no between-tertile  
 224 differences were seen for any of the other bone-related biochemical  
 225 parameters. There was no significant difference in history of smoking,  
 226 alcohol intake or physical activity between groups. There were 40  
 227 incident self-reported fractures during the follow-up period with no  
 228 apparent difference in fracture rates between UA tertiles.

Crude bone density measures of the study cohort at the final visit  
 are shown in Table 2a. When analysed by tertile of serum UA, women  
 with higher UA levels had significantly higher absolute BMD at all  
 skeletal sites at baseline (data not shown) and follow-up visits. Ad-  
 justed means for final visit bone density measures at different skeletal  
 sites across tertiles of UA are presented in Fig. 1. Unadjusted body  
 composition measures of the study cohort at the final visit are  
 shown in Table 2b. Both body fat and lean mass measures as well as  
 the body fat to lean mass ratio were significantly higher in the medi-  
 um and high UA group. Similar results were obtained for baseline  
 body composition characteristics (data not shown). Adjusted means  
 for the final visit body weight, lean mass and fat mass measures  
 across tertiles of UA are presented in Fig. 2.

Multiple regression analysis was performed to examine the  
 cross-sectional associations between serum uric acid levels and final  
 visit BMD measures at different skeletal sites. Estimates of the fully  
 adjusted regression models are presented in Table 3. Serum UA levels  
 were positively associated with baseline and final visit cross-sectional  
 bone density measures at all skeletal sites after adjustment for GFR,  
 serum Ca and CTX-I levels, age, FM/LM/Ht<sup>2</sup>, smoking, alcohol, HRT  
 use and physical activity.

## 250 Longitudinal analyses

Mean duration of follow-up was  $9.7 \pm 1.8$  years and did not differ  
 between UA tertiles. Longitudinal bone density and body composition

t1.1 Table 1

t1.2 Demographic, biochemical and lifestyle characteristics of the study subjects at final visit stratified by tertiles of serum uric acid levels (Final Visit).<sup>a</sup>

t1.3	Tertiles of uric acid levels				Sig.	
	All	1	2	3		
	(N=356)	(N=122)	(N=106)	(N=128)		
t1.6	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD		
t1.7	Age (years)	60.44 $\pm$ 7.89	59.23 $\pm$ 7.88	60.47 $\pm$ 8.30	61.57 $\pm$ 7.43	0.019*
t1.8	Duration of Follow-Up (years)	9.68 $\pm$ 1.84	9.87 $\pm$ 1.51	9.45 $\pm$ 2.09	9.69 $\pm$ 1.89	0.450
t1.9	Weight (kg)	69.44 $\pm$ 12.36	63.58 $\pm$ 9.61	70.63 $\pm$ 13.06	74.00 $\pm$ 11.88	<0.000 <sup>§</sup>
t1.10	Height (m)	1.61 $\pm$ 0.06	1.61 $\pm$ 0.06	1.62 $\pm$ 0.06	1.60 $\pm$ 0.06	0.190
t1.11	BMI (kg/m <sup>2</sup> )	26.83 $\pm$ 4.92	24.48 $\pm$ 3.83	27.09 $\pm$ 5.19	28.85 $\pm$ 4.68	<0.000 <sup>§</sup>
t1.12	<b>Biochemistry measures:</b>					
t1.13	Uric Acid Baseline Visit (mmol/L)	0.26 $\pm$ 0.06	0.22 $\pm$ 0.05	0.25 $\pm$ 0.05	0.30 $\pm$ 0.05	<0.000 <sup>§</sup>
t1.14	Uric Acid Final Visit (mmol/L)	0.29 $\pm$ 0.07	0.21 $\pm$ 0.04	0.28 $\pm$ 0.01	0.36 $\pm$ 0.05	<0.000 <sup>§</sup>
t1.15	A% $\Delta$ Change in Uric Acid (% per yr)	1.36 $\pm$ 3.16	0.26 $\pm$ 3.84	1.59 $\pm$ 2.37	2.36 $\pm$ 2.67	<0.000 <sup>§</sup>
t1.16	Total Cholesterol (mmol/L)	5.47 $\pm$ 0.96	5.41 $\pm$ 0.92	5.33 $\pm$ 0.93	5.63 $\pm$ 1.01	0.060
t1.17	HDLc (mmol/L)	1.45 $\pm$ 0.38	1.50 $\pm$ 0.42	1.45 $\pm$ 0.36	1.41 $\pm$ 0.36	0.053
t1.18	LDLc (mmol/L)	3.39 $\pm$ 1.02	3.21 $\pm$ 1.17	3.30 $\pm$ 0.87	3.63 $\pm$ 0.94	0.001 <sup>±</sup>
t1.19	Triglycerides	1.20 $\pm$ 0.60	1.08 $\pm$ 0.41	1.19 $\pm$ 0.72	1.31 $\pm$ 0.63	0.003 <sup>±</sup>
t1.20	Calcium (mmol/L)	2.30 $\pm$ 0.09	2.28 $\pm$ 0.09	2.30 $\pm$ 0.09	2.32 $\pm$ 0.09	<0.000 <sup>§</sup>
t1.21	Creatinine (micromol/L)	68.99 $\pm$ 11.06	65.48 $\pm$ 9.14	68.45 $\pm$ 9.29	72.79 $\pm$ 12.82	<0.000 <sup>§</sup>
t1.22	C-Reactive Protein (nmol/L)	2.80 $\pm$ 3.65	2.28 $\pm$ 3.94	2.79 $\pm$ 3.74	3.30 $\pm$ 3.22	0.027*
t1.23	Serum CTX-I ( $\mu$ g/L)	283.97 $\pm$ 157.86	260.71 $\pm$ 142.58	290.35 $\pm$ 157.08	300.67 $\pm$ 170.37	0.047*
t1.24	PINP ( $\mu$ g/L)	46.14 $\pm$ 20.20	46.82 $\pm$ 20.39	47.10 $\pm$ 20.13	44.73 $\pm$ 20.17	0.410
t1.25	GFR	85.53 $\pm$ 22.84	83.26 $\pm$ 18.58	88.26 $\pm$ 27.44	85.44 $\pm$ 22.26	0.466
t1.26	Fractures:	39 (11.0%)	12 (9.8%)	16 (15.1%)	11 (8.6%)	0.200
t1.27	<b>Lifestyle characteristics:</b>					
t1.28	Smoking History: (N (%))					0.620
t1.29	Never	224 (62.9%)	72 (59.0%)	72 (67.9%)	80 (62.5%)	
t1.30	Current	21 (6.9%)	7 (5.7%)	7 (6.6%)	7 (5.5%)	
t1.31	Ex-smoker	111 (31.2%)	43 (35.2%)	27 (25.5%)	41 (32.0%)	
t1.32	Alcohol intake (N(%))					0.411
t1.33	$\leq 1$ drink per week	142 (39.9%)	50 (41.0%)	43 (40.6%)	49 (38.3%)	
t1.34	2–14 drinks per week	207 (58.1%)	71 (58.2%)	62 (58.5%)	74 (57.8%)	
t1.35	$\geq$ Drinks 14 per week	7 (2.0%)	1 (0.8%)	1 (0.9%)	5 (3.9%)	
t1.36	Physical activity (N (%))					0.788
t1.37	None	18 (5.1%)	4 (3.3%)	7 (6.6%)	7 (5.5%)	
t1.38	< 30 min per day	167 (46.9%)	58 (47.5%)	47 (44.3%)	62 (48.4%)	
t1.39	$\geq 30$ min per day	171 (48.0%)	62 (50.8%)	50 (47.2%)	59 (46.1%)	

t1.40 <sup>a</sup> ANOVA and test of linearity were performed.t1.41 \*  $p < 0.05$ .t1.42 <sup>±</sup>  $p < 0.01$ .t1.43 <sup>§</sup>  $p < 0.001$ .

**Table 2a**Cross-sectional (final visit) and longitudinal bone mineral density measures of the study subjects, stratified by tertiles of serum uric acid levels.<sup>a</sup>

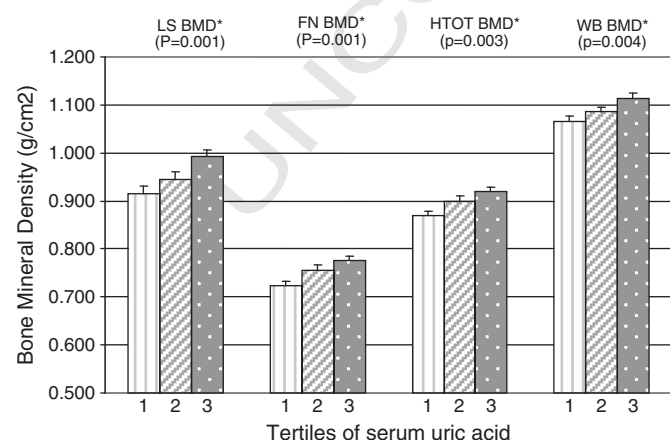
	Tertiles of uric acid				Sig.
	All	1	2	3	
	(N= 356)	(N= 122)	(N= 106)	(N= 128)	
	Mean ± SD	Mean ± SD	Mean ± SD	Mean ± SD	
<i>Cross-sectional BMD (g/cm<sup>2</sup>)</i>					
Lumbar spine	0.952 ± 0.158	0.923 ± 0.157	0.949 ± 0.148	0.982 ± 0.164	0.003 <sup>±</sup>
Femoral neck	0.751 ± 0.120	0.726 ± 0.111	0.761 ± 0.112	0.767 ± 0.132	0.007 <sup>±</sup>
Hip total	0.896 ± 0.125	0.869 ± 0.123	0.907 ± 0.119	0.914 ± 0.129	0.005 <sup>±</sup>
Forearm total	0.522 ± 0.058	0.511 ± 0.061	0.527 ± 0.054	0.528 ± 0.057	0.022 <sup>*</sup>
Whole body total	1.089 ± 0.116	1.071 ± 0.116	1.091 ± 0.120	1.105 ± 0.113	0.021 <sup>*</sup>
<i>A%ΔChange in BMD (% per year)</i>					
Lumbar spine	-0.50 ± 0.83	-0.58 ± 0.85	-0.57 ± 0.76	-0.37 ± 0.86	0.044 <sup>*</sup>
Femoral neck	-0.45 ± 0.75	-0.46 ± 0.76	-0.45 ± 0.78	-0.44 ± 0.72	0.794 <sup>§</sup>
Hip total	-0.38 ± 0.61	-0.39 ± 0.67	-0.39 ± 0.52	-0.36 ± 0.61	0.636
Forearm total	-0.62 ± 0.58	-0.62 ± 0.62	-0.66 ± 0.61	-0.59 ± 0.52	0.630
Whole body total	-0.21 ± 0.65	-0.25 ± 0.67	-0.24 ± 0.62	-0.14 ± 0.65	0.200

<sup>a</sup> ANOVA and test of linearity were performed.<sup>\*</sup> *p* < 0.05.<sup>±</sup> *p* < 0.01.<sup>§</sup> *p* < 0.001.

measures of the study cohort stratified by tertiles of UA are shown in Tables 2a and 2b. Annual rates of increase in body weight and lean body mass (LM) over the preceding 9 years were significantly related to higher serum UA levels and these associations remained after adjusting for potential confounders: age, height, history of smoking, alcohol intake, HRT use and physical activity (data not shown). There was a general trend for women in the highest tertile of UA to be losing bone at a slower rate than those with lower UA level. However this relationship was statistically significant only for A%ΔLSBMD. Women in the highest UA tertile had gained more weight and more lean mass over time, but changes in fat mass were not significantly different by UA tertiles (Fig. 2).

Multiple regression analyses were performed to examine the associations between serum UA and longitudinal BMD measures at different skeletal sites (Table 4). Higher rates of annual change in UA levels were associated with slower rate of decline in BMD at all skeletal sites. When the regression models were adjusted for A%ΔFM (model 2) or A%ΔLM (model 3), the associations between change in UA levels and change in BMD measures remained significant at the spine only.

The results of the time dependent mixed model regression analyses confirmed these findings (not shown).



**Fig. 1.** Adjusted means of Lumbar Spine, Femoral Neck, Total Hip and Whole Body cross-sectional BMD measures (final visit) across tertiles of Serum Uric Acid (N = 356)\*. \* BMD means adjusted for GFR, serum Ca and CTX-I levels, age, height; history of smoking, history alcohol intake, history HRT use and physical activity.

## Discussion

274

In a previous cross-sectional study [31] the CHAMP consortium has reported that higher serum UA levels are associated with greater BMD at all skeletal sites in an older male population. In the present study, we have confirmed, for the first time, that a similar relationship exists between serum UA and BMD in peri- and postmenopausal women. In addition, we have shown that serum UA is also associated with the rate of change in BMD over time in women. In the lumbar spine, forearm and total body, those with higher UA levels were relatively protected from bone loss compared to those with lower levels. However the protective effect of UA on longitudinal BMD appeared to be weaker at hip sites.

Body weight is well known to be related to BMD [27]. However, there has been considerable controversy about the association between fat body mass and lean body mass and their relationship to BMD [27,28,30]. Gender and age are likely to be important factors in modifying this relationship given we showed previously in a cross-sectional study of opposite sex twins that lean mass had stronger relationships with most bone variables than fat mass in both genders at all ages, but fat mass had a positive relationship with total body and hip BMD in women under 50 and men over 50 years of age [29]. Body weight and in particular obesity are also associated with serum UA levels [12,20,42]. In the present study, UA was also associated with cross-sectional weight, FM and LM measures and the rate of change over time in lean body mass. Those women with high UA levels gained more lean body mass without much change in fat mass. However after adjusting for lean mass or fat mass, the relationship between UA and BMD remained.

As noted above, UA is the end product of purine metabolism which in turn is related to lean body mass. However our analyses of changes in body composition do not explain the relationship observed between serum UA and cross-sectional or longitudinal BMD. We found modestly higher serum CTX-I values in women in the highest tertile of UA, which might be expected to be associated with increased bone loss over time rather than what was actually observed. Taken together, these longitudinal data suggest that the relationship between serum UA and lumbar spine BMD in women, both cross-sectionally and longitudinally, is not mediated to any great extent by the relationship between body composition and UA or by direct effects on bone remodeling. Evidence from observational and epidemiological studies has linked oxidative stress to reduced BMD and osteoporosis [9–11] and since UA accounts for approximately

**Table 2b**  
Cross-sectional (final visit) and Longitudinal Body composition measures of the study subjects, stratified by tertiles of serum uric acid levels.<sup>a</sup>

		Tertiles of uric acid				
		All	1	2	3	Sig.
		(N=356)	(N=122)	(N=106)	(N=128)	
		Mean ± SD	Mean ± SD	Mean ± SD	Mean ± SD	
<b>Cross-sectional (kg):</b>						
t3.7	Whole body fat mass	23.84 ± 7.88	20.11 ± 6.05	24.44 ± 8.30	26.87 ± 7.68	<0.000 <sup>§</sup>
t3.8	Whole body lean mass	40.07 ± 5.55	38.30 ± 4.30	40.34 ± 6.75	41.51 ± 5.06	<0.000 <sup>§</sup>
t3.9	Fat mass/lean mass	0.59 ± 0.16	0.52 ± 0.14	0.59 ± 0.15	0.64 ± 0.16	<0.000 <sup>§</sup>
t3.10	<b>Longitudinal (% per year)</b>					
t3.11	A%Δ Whole body fat mass	0.09 ± 2.09	0.09 ± 2.06	0.07 ± 2.20	0.11 ± 2.03	0.947 <sup>±</sup>
t3.12	A%Δ Whole body lean mass	0.72 ± 0.69	0.49 ± 0.60	0.70 ± 0.70	0.95 ± 0.70	<0.000 <sup>§</sup>
t3.13	A%Δ Fat mass /lean mass	-0.57 ± 2.00	-0.36 ± 1.98	-0.58 ± 2.08	-0.75 ± 1.97	0.131 <sup>*</sup>

t3.15 <sup>a</sup> ANOVA and test of linearity were performed.

t3.16 \* *p* < 0.05.

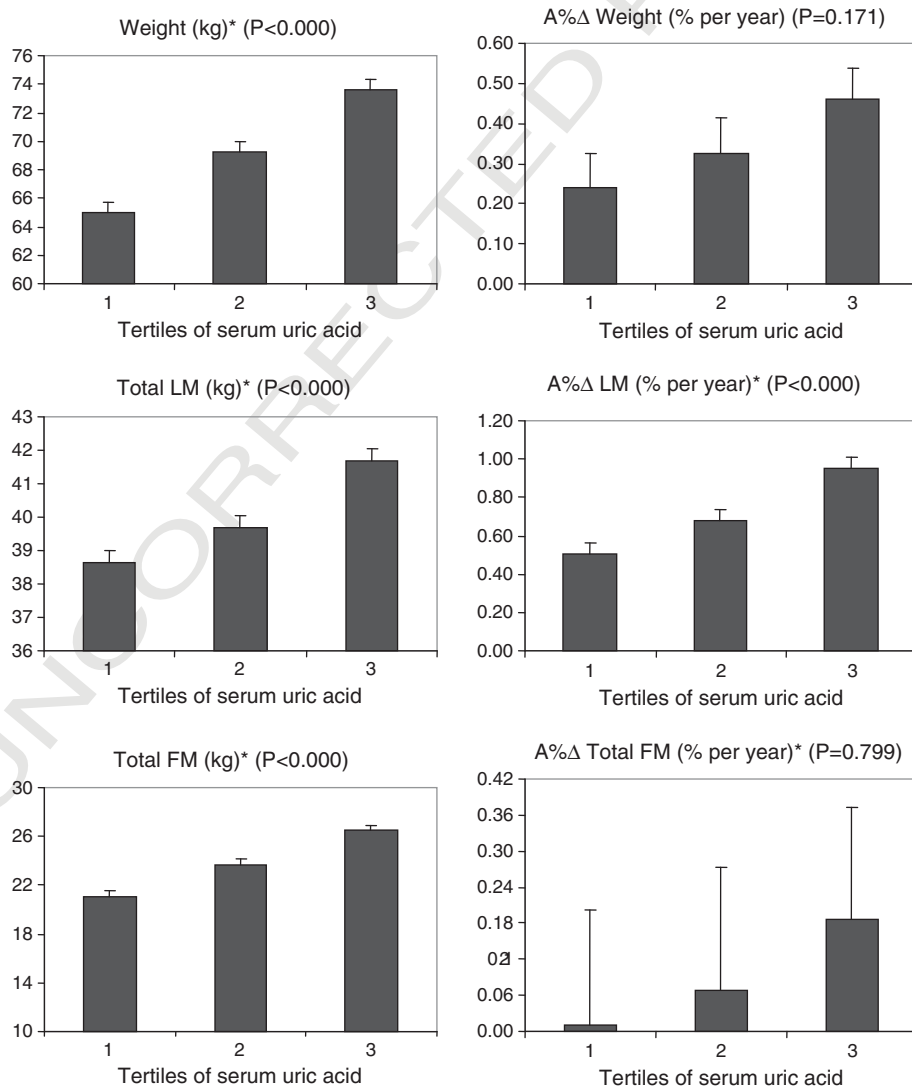
t3.17 ± *p* < 0.01.

t3.18 § *p* < 0.001.

315 half of the antioxidant properties of human plasma [3], this mecha-  
316 nism of action as an explanation of our findings requires further  
317 investigation. The lesser protective effect of UA on hip BMD in our  
318 longitudinal analyses may reflect that hip BMD is more influenced

by body composition measures [43,44] and local environmental 319  
factors such as weight bearing and also deserves further study. 320

Although we have demonstrated that higher serum UA values are 321  
associated with higher BMD and lower rates of bone loss in women, 322



**Fig. 2.** Adjusted means of body weight and body composition measures across tertiles of uric acid\*. \*Means adjusted for UA, GFR, serum Ca and CTX-I levels, age, height; history of smoking, history alcohol intake, history of HRT use and physical activity.

**Table 3**

Multiple regression analysis of the association between serum uric acid and cross-sectional BMD measures at different skeletal sites.

Regression models	Baseline visit*		Final Follow up visit**	
	$\beta^*$	Sig.	$\beta^*$	Sig.
BMD (g/cm <sup>2</sup> )				
Lumbar spine	0.190	0.000 <sup>§</sup>	0.243	0.000 <sup>§</sup>
Femoral neck	0.169	0.002 <sup>±</sup>	0.221	0.000 <sup>§</sup>
Total hip	0.167	0.002 <sup>±</sup>	0.195	0.000 <sup>§</sup>
Total forearm	0.136	0.012 <sup>*</sup>	0.191	0.000 <sup>§</sup>
Whole body	0.170	0.001 <sup>±</sup>	0.230	0.000 <sup>§</sup>

Each regression model included one BMD measure as dependent variable.

Independent variables included in the regression analyses were: Bsl UA, Bsl: age, LM/FM/height<sup>2</sup>; history of smoking, history alcohol intake, history HRT use, and physical activity.

Final Visit UA, Final visit: age, LM/FM/height<sup>2</sup>; history of smoking, history alcohol intake, history HRT use, and physical activity.

\* $p < 0.05$ .

<sup>±</sup> $p < 0.01$ .

<sup>§</sup> $p < 0.001$ .

those with higher UA values also had a more adverse lipid profile with higher total cholesterol, lower LDLC and higher triglyceride values and the clinical significance of these opposing effects need further evaluation. The existence of a possible link between bone, fat and atherogenic pathways has been recognised for some time and serum UA should now be added to consideration of any such interactions. Previous studies in this regard are conflicting. Early postmenopausal women with an atherogenic lipid profile have been reported to have lower lumbar and femoral neck BMD and an increased risk of osteopaenia than those with a normal lipid profile [45], which differs from our findings. In a longitudinal study in postmenopausal women aged 50–75 years, those with the largest increases in serum cholesterol showed the greatest decreases in spine BMD independently of change in the body mass index [46]. We have also previously reported a modest inverse relationship between total serum cholesterol and spine but not hip BMD in perimenopausal women [35], but none of these studies considered the influence of serum UA.

Our study has a number of strengths. We measured change in BMD and body composition measures in women over almost 10 years and UA levels at baseline and final visits. For the first time the associations between serum UA and BMD and body composition measured by DXA were studied on a relatively healthy population of peri- and postmenopausal women. Our study also has some limitations. Only UA levels were measured at the two end points of the study. Bone markers or other biochemical characteristics were only

**Table 4**

Multiple regression analysis of the association between longitudinal measures of uric acid and BMD at different skeletal sites.

Regression models	Model 1		Model 2		Model 3	
	$\beta^*$	Sig.	$\beta^*$	Sig.	$\beta^*$	Sig.
<b>A%<math>\Delta</math>BMD (% per year):</b>						
Lumbar Spine	0.170	0.006 <sup>±</sup>	0.161	0.009 <sup>±</sup>	0.134	0.036 <sup>*</sup>
Femoral Neck	0.121	0.054	0.112	0.073	0.072	0.264
Total Hip	0.121	0.050 <sup>*</sup>	0.108	0.076	0.052	0.405
Total Forearm	0.077	0.207	0.073	0.231	0.038	0.550
Whole Body	0.145	0.022 <sup>*</sup>	0.138	0.029 <sup>*</sup>	0.082	0.202

Each regression model included one BMD measure as dependent variable.

Independent variables included in the regression analyses were:

Model 1: A% $\Delta$ UA, Bsl UA, Bsl BMD, GFR, serum Ca, Cholesterol and CTX-I levels, age, height; history of smoking, history alcohol intake, history HRT use, physical activity and  $\Delta$ BMI.

Model 2: as Model 1 + A% $\Delta$  total body fat mass.

Model 3: as Model 1 + A% $\Delta$  total body lean mass.

\* $p < 0.05$ .

<sup>±</sup> $p < 0.01$ .

<sup>§</sup> $p < 0.001$ .

measured at the final visit. Whereas DXA is regarded by majority as a reference technique for the measurement of the bone mineral, fat and fat-free soft tissue compartments of the body, it is not without limitations. Several studies suggest that long term DXA precision results may be affected by substantial weight gain [47–50]. In our study subjects with severe obesity that affected the quality of DXA scans were excluded and regression analyses of the longitudinal BMD measures were adjusted for rates of changes in BMI.

The variability in rates of change in BMD and body composition was high, although we measured change over almost 10 years and the changes we observed are consistent with annual rates of BMD change reported by others [51,52]. With only 40 incident fractures during the follow-up period we lacked power to examine the effect of UA on fractures.

We recently reported that serum UA levels were significantly associated with BMD at various skeletal sites after adjusting for covariates in a large population-based study of older men [30] and have now confirmed a similar relationship exists in peri- and postmenopausal women.

For decades it has been hypothesised that the antioxidant properties of uric acid might be protective against aging, oxidative stress, and oxidative injury of cells, including cardiac, vascular, and neural cells. However, recent epidemiological and clinical evidences suggest that hyperuricaemia might be a risk factor for cardiovascular disease, where enhanced oxidative stress plays an important pathophysiological role. It has also been hypothesised that hyperuricaemia might be involved in chronic heart failure and metabolic syndrome [7,53,54]. The apparent paradox between protective and toxic effects of UA is supported by clinical evidence that antioxidant compounds may become pro-oxidant compounds in certain situations, particularly when they are present in blood at abnormally high levels [7].

The present study suggests that serum UA, when present at higher physiological concentrations, may have protective effects on BMD, most likely through its antioxidant properties. However, further studies are needed to establish the precise mechanism of action and whether serum UA plays a role in antagonising oxidative stress-induced bone loss.

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## References

- Edwards NL. The role of hyperuricemia and gout in kidney and cardiovascular disease. *Cleve Clin J Med* 2008;75:S13–6.
- Mandell BF. Clinical manifestations of hyperuricemia and gout. *Cleve Clin J Med* 2008;75:S5–8.
- Masseoud D, Rott K, Liu-Bryan R, Agudelo C. Overview of hyperuricaemia and gout. *Curr Pharm Des* 2005;11:4117–24.
- Rossi SE. Australian medicines handbook 2012. Adelaide: Australian Medicines Handbook Pty Ltd.; 2012.
- Glantzounis GK, Tsimoyiannis EC, Kappas AM, Galaris DA. Uric acid and oxidative stress. *Curr Pharm Des* 2005;11:4145–51.
- Dalbeth N, Merriman T. Crystal ball gazing: new therapeutic targets for hyperuricaemia and gout. *Rheumatology* 2009;48:222–6.
- Lippi G, Montagnana M, Franchini M, Favaloro EJ, Targher G. The paradoxical relationship between serum uric acid and cardiovascular disease. *Clin Chim Acta* 2008;392:1–7.
- Sanchez-Rodriguez M, Ruiz-Ramos M, Correa-Munoz E, Mendoza-Nunez V. Oxidative stress as a risk factor for osteoporosis in elderly Mexicans as characterized by antioxidant enzymes. *BMC Musculoskelet Disord* 2007;8:124.
- Sugiura M, Nakamura M, Ogawa K, Ikoma Y, Ando F, Yano M. Bone mineral density in post-menopausal female subjects is associated with serum antioxidant carotenoids. *Osteoporos Int* 2008;19:211–9.
- Sahni S, Hannan MT, Blumberg J, Cupples LA, Kiel DP, Tucker KL. Inverse association of carotenoid intakes with 4-y change in bone mineral density in elderly men and women: the Framingham Osteoporosis Study. *Am J Clin Nutr* 2009;89:416–24.

- 415 [11] Sendur OF, Turan Y, Tastaban E, Serter M. Antioxidant status in patients with  
416 osteoporosis: a controlled study. *Joint Bone Spine* 2009;76:514–8.
- 417 [12] Lee HJ, Park HT, Cho GJ, Yi KW, Ahn KH, Shin JH, et al. Relationship between uric acid  
418 and metabolic syndrome according to menopausal status. *Gynecol Endocrinol* 2010;  
419 1–6.
- 420 [13] Ishizaka N, Ishizaka Y, Toda EI, Nagai R, Yamakado M. Association between serum  
421 uric acid, metabolic syndrome, and carotid atherosclerosis in Japanese individ-  
422 uals. *Arterioscler Thromb Vasc Biol* 2005;25:1038–44.
- 423 [14] Puig JG, Martinez MA. Hyperuricemia, gout and the metabolic syndrome. *Curr*  
424 *Opin Rheumatol* 2008;20:187–91.
- 425 [15] Oda E, Kawai R, Sukumaran V, Watanabe K. Uric acid is positively associated with  
426 metabolic syndrome but negatively associated with diabetes in Japanese men.  
427 *Intern Med* 2009;48:1785–91.
- 428 [16] Dehghan A, van Hoek M, Sijbrands EJG, Hofman A, Witteman JCM. High serum  
429 uric acid as a novel risk factor for type 2 diabetes. *Diabetes Care* 2008;31:361–2.
- 430 [17] Kramer CK, Von Muhlen D, Jassal SK, Barrett-Connor E. Serum uric acid levels  
431 improve prediction of incident type 2 diabetes in individuals with impaired  
432 fasting glucose. *Diabetes Care* 2009;32:1272–3.
- 433 [18] Lee J, Sparrow D, Vokonas PS, Landsberg L, Weiss ST. Uric acid and coronary heart  
434 disease risk: evidence for a role of uric acid in the obesity-insulin resistance  
435 syndrome. *The Normative Aging Study*. *Am J Epidemiol* 1995;142:288–94.
- 436 [19] Iribarren C, Folsom AR, Eckfeldt JH, McGovern PG, Nieto FJ. Correlates of uric acid  
437 and its association with asymptomatic carotid atherosclerosis: the ARIC Study.  
438 *Atherosclerosis Risk in Communities*. *Ann Epidemiol* 1996;6:331–40.
- 439 [20] Bonora E, Targher G, Zenere MB, Saggiani F, Cacciatori V, Tosi F, et al. Relationship  
440 of uric acid concentration to cardiovascular risk factors in young men. Role of  
441 obesity and central fat distribution. *The Verona Young Men Atherosclerosis Risk*  
442 *Factors Study*. *Int J Obes Relat Metab Disord* 1996;20:975–80.
- 443 [21] Russo C, Olivieri O, Girelli D, Guarini P, Corrocher R. Relationships between serum  
444 uric acid and lipids in healthy subjects. *Prev Med* 1996;25:611–6.
- 445 [22] Alderman MH. Serum uric acid as a cardiovascular risk factor for heart disease.  
446 *Curr Hypertens Rep* 2001;3:184–9.
- 447 [23] Ahmed N, Anwar W, Waqas H. Obesity, hyperlipidemia, and hyperuracemia in  
448 young and old hypertensive patients. *J Ayub Med Coll Abbottabad* 2009;21:53–6.
- 449 [24] Mazzali M, Kanbay M, Segal M, Shafiq M, Jalal D, Feig D, et al. Uric acid and hyper-  
450 tension: cause or effect? *Curr Rheumatol Rep* 2010;12:108–17.
- 451 [25] Kirchengast S, Peterson B, Hauser G, Knogler W. Body composition characteristics  
452 are associated with the bone density of the proximal femur end in middle- and  
453 old-aged women and men. *Maturitas* 2001;39:133–45.
- 454 [26] Pongchaiyakul C, Nguyen T, Kosulwat V, Rojroongwasinkul N, Charoenkiatkul S,  
455 Eisman J, et al. Contribution of lean tissue mass to the urban–rural difference in  
456 bone mineral density. *Osteoporos Int* 2005;16:1761–8.
- 457 [27] Dytfield J, Ignaszak-Szczepaniak M, Gowin E, Michalak M, Horst-Sikorska W. Influe-  
458 nce of lean and fat mass on bone mineral density (BMD) in postmenopausal  
459 women with osteoporosis. *Arch Gerontol Geriatr* in press;Corrected Proof.
- 460 [28] Gjesdal CG, Halse JI, Eide GE, Brun JG, Tell GS. Impact of lean mass and fat mass on  
461 bone mineral density: the Hordaland Health Study. *Maturitas* 2008;59:191–200.
- 462 [29] Makovey J, Naganathan V, Sambrook P. Gender differences in relationships between  
463 body composition components, their distribution and bone mineral density: a cross-  
464 sectional opposite sex twin study. *Osteoporos Int* 2005;16:1495–505.
- 465 [30] Bogl LH, Latvala A, Kaprio J, Sovijarvi O, Rissanen A, Pietilainen KH. An investiga-  
466 tion into the relationship between soft tissue body composition and bone mineral  
467 density in a young adult twin sample. *J Bone Miner Res* 2011;26:79–87.
- 468 [31] Nabipour I, Sambrook PN, Blyth FM, Janu MR, Waite LM, Naganathan V, et al. Serum  
469 uric acid is associated with bone health in older men: a cross-sectional  
470 population-based study. *J Bone Miner Res* 2011;26:955–64.
- 471 [32] Sarna S, Kaprio J, Sistonen P, Koskenvuo M. Diagnosis of twin zygosity by mailed  
472 questionnaire. *Hum Hered* 1978;28:241–54.
- 473 [33] Cockcroft DW, Gault MH. Prediction of creatinine clearance from serum creatinine.  
474 *Nephron* 1976;16:31–41.
- 475 [34] Gault MH, Longrich LL, Harnett JD, Wesolowski C. Predicting glomerular function  
476 from adjusted serum creatinine. *Nephron* 1992;62:249–56.
- 477 [35] Makovey J, Chen JS, Hayward C, Williams FM, Sambrook PN. Association between  
478 serum cholesterol and bone mineral density. *Bone* 2009;44:208–13.
- 479 [36] Christian JC, Yu PL, Slemenda CW, Johnston Jr CC. Heritability of bone mass: a lon-  
480 gitudinal study in aging male twins. *Am J Hum Genet* 1989;44:429–33.
- 481 [37] Bainbridge KE, Sowers MF, Crutchfield M, Lin X, Jannausch M, Harlow SD. Natural  
482 history of bone loss over 6 years among premenopausal and early postmenopausal  
483 women. *Am J Epidemiol* 2002;156:410–7.
- 484 [38] Ho SC, Chan SG, Yip YB, Chan CS, Woo JL, Sham A. Change in bone mineral density  
485 and its determinants in pre- and perimenopausal Chinese women: the Hong Kong  
486 Perimenopausal Women Osteoporosis Study. *Osteoporos Int* 2008;19:1785–96.
- 487 [39] Kaptoge S, Reid DM, Scheidt-Nave C, Poor G, Pols HA, Khaw KT, et al. Geographic  
488 and other determinants of BMD change in European men and women at the hip  
489 and spine: a population-based study from the Network in Europe for Male Osteo-  
490 porosis (NEMO). *Bone* 2007;40:662–73.
- 491 [40] Hannan MT, Tucker KL, Dawson-Hughes B, Cupples LA, Felson DT, Kiel DP. Effect  
492 of dietary protein on bone loss in elderly men and women: the Framingham Osteo-  
493 porosis Study. *J Bone Miner Res* 2000;15:2504–12.
- 494 [41] Sirola J, Salovaara K, Rikkonen T, Karkkainen M, Tuppurainen M, Jurvelin JS, et al. Bone  
495 health-related factors and the use of bisphosphonates in community setting—15-year  
496 follow-up study. *Osteoporos Int* 2011;22:255–64.
- 497 [42] Loenen HMJA, Eshuis H, Löwik MRH, Schouten EG, Hulshof KFAM, Odink J, et al.  
498 Serum uric acid correlates in elderly men and women with special reference to  
499 body composition and dietary intake (Dutch nutrition surveillance system).  
500 *J Clin Epidemiol* 1990;43:1297–303.
- 501 [43] Abrahamsen B, Stilgren LS, Hermann AP, Tofteng CL, Barenholdt O, Vestergaard P,  
502 et al. Discordance between changes in bone mineral density measured at different  
503 skeletal sites in perimenopausal women—implications for assessment of bone  
504 loss and response to therapy: the Danish Osteoporosis Prevention Study. *J Bone*  
505 *Miner Res* 2001;16:1212–9.
- 506 [44] Dennison E, Eastell R, Fall CH, Kellingray S, Wood PJ, Cooper C. Determinants of  
507 bone loss in elderly men and women: a prospective population-based study.  
508 *Osteoporos Int* 1999;10:384–91.
- 509 [45] Orozco P. Atherogenic lipid profile and elevated lipoprotein (a) are associated  
510 with lower bone mineral density in early postmenopausal overweight women.  
511 *Eur J Epidemiol* 2004;19:1105–12.
- 512 [46] Tanko LB, Bagger YZ, Nielsen SB, Christiansen C. Does serum cholesterol contrib-  
513 ute to vertebral bone loss in postmenopausal women? *Bone* 2003;32:8–14.
- 514 [47] Hangartner TN. A study of the long-term precision of dual-energy X-ray absorpti-  
515 ometry bone densitometers and implications for the validity of the least-  
516 significant-change calculation. *Osteoporos Int* 2007;18:513–23.
- 517 [48] Blake GM, Herd RJ, Patel R, Fogelman I. The effect of weight change on total body  
518 dual-energy X-ray absorptiometry: results from a clinical trial. *Osteoporos Int*  
519 2000;11:832–9.
- 520 [49] Patel R, Blake GM, Rymer J, Fogelman I. Long-term precision of DXA scanning assessed  
521 over seven years in forty postmenopausal women. *Osteoporos Int* 2000;11:68–75.
- 522 [50] Rajamanohara R, Robinson J, Rymer J, Patel R, Fogelman I, Blake GM. The effect  
523 of weight and weight change on the long-term precision of spine and hip DXA  
524 measurements. *Osteoporos Int* 2011;22:1503–12.
- 525 [51] Nguyen TV, Center JR, Eisman JA. Femoral neck bone loss predicts fracture risk  
526 independent of baseline BMD. *J Bone Miner Res* 2005;20:1195–201.
- 527 [52] McLean RR, Jacques PF, Selhub J, Fredman L, Tucker KL, Samelson EJ, et al. Plasma  
528 B vitamins, homocysteine, and their relation with bone loss and hip fracture in  
529 elderly men and women. *J Clin Endocrinol Metab* 2008;93:2206–12.
- 530 [53] Lippi G, Montagnana M, Luca SG, Targher G, Cesare GG. Epidemiological asso-  
531 ciation between uric acid concentration in plasma, lipoprotein(a), and the tradi-  
532 tional lipid profile. *Clin Cardiol* 2010;33:E76–80.
- 533 [54] Lippi G, Montagnana M, Franchini M, Guidi GC, Targher G. Uric acid concentration  
534 in patient with acute coronary syndrome. *Intern Emerg Med* 2008;3:409–11.
- 535