

# Genetic characterization of seedling and adult plant resistance against three rust diseases in the 22<sup>nd</sup> SAWSN CIMMYT Nursery

Singh B, Bansal UK and Bariana HS

University of Sydney PBI-Cobbitty, Faculty of Agriculture and Natural Resources, PMB11, Camden, NSW 2570, Australia

## INTRODUCTION

The three rusts of wheat, namely; stem rust (*Puccinia graminis* f. sp. *tritici*), leaf rust (*Puccinia recondita*) and stripe rust (*Puccinia striiformis* f. sp. *tritici*) constitute the major group of diseases that cause yield losses worldwide<sup>1</sup>. Resistance to diseases can be controlled by seedling or adult plant resistance (APR) genes. Combination of both types of genes may be present in a single genotype. Seventy six entries belonging to the 22<sup>nd</sup> Semi-Arid Wheat Screening Nursery (SAWSN) were screened to determine the presence of genetically characterised genes and to identify new sources of resistance. Results presented in this paper also reflect on the extent of genetic diversity of genotype for rust resistance.

## METHODOLOGY

The 22<sup>nd</sup> SAWSN was tested in the greenhouse against an array of pathotypes for postulation of seedling resistance genes and evaluated in the field against the commercially important pathotypes of three rust pathogens. Stripe rust assessment in the greenhouse was made on a 0-4 scale<sup>2</sup>. For stem rust and leaf rust assessments were made on a scale described by Stakman *et al.*<sup>3</sup> with some modifications proposed by Luig<sup>4</sup>. Field assessment of adult plant responses to three rust diseases was based on a 1-9 scale<sup>5</sup>. The adult plant resistance genes *Lr34* and *Sr2* were identified by using molecular markers *swm10<sub>211</sub>*<sup>6</sup> and *stm560.3<sub>171</sub>*<sup>7</sup>, respectively. Marker (Gb)<sub>130</sub><sup>8</sup> was used to confirm the postulation of *Lr19*.

## RESULTS

### SEEDLING RESISTANCE

#### Stripe rust

Twelve entries (16%) carried the 1BL.1RS (wheat-rye) translocation (Fig. 1). This translocation carries rust resistance genes *Yr9*, *Lr26* and *Sr31* (9). Stripe rust resistance genes, *Yr6*, *Yr7*, *Yr9*, *Yr17* and *Yr27* were present either singly or in combinations in 27 entries (Table 1). *Yr9* was postulated alone in six entries and in combination with *Yr27* in five entries. In one entry *Yr9* was postulated with some unknown resistance. Resistance genes in 14 entries could not be postulated

with the current set of pathotypes. Thirty five entries did not carry any seedling stripe rust resistance gene.

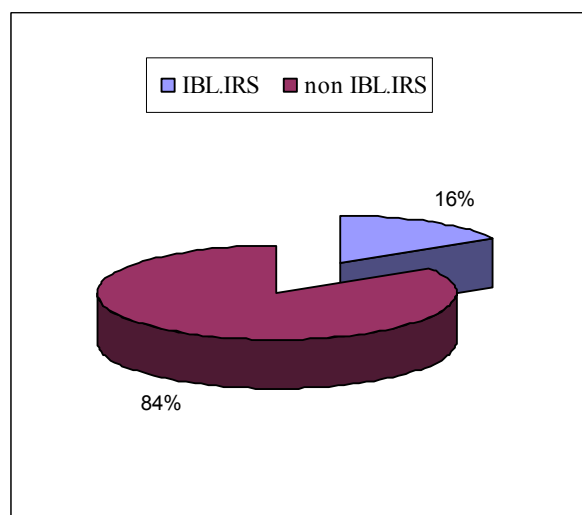


Fig 1. Occurrence of IBL.1RS translocation in 22<sup>nd</sup> SAWSN

Table 1. Frequency of stripe rust resistance genes in the 22<sup>nd</sup> SAWSN

Postulated gene	No. of entries
<i>Yr9</i>	12
<i>Yr27</i>	8
<i>Yr7</i>	4
<i>Yr6</i>	2
<i>Yr17</i>	1
Unknown	14
Nil	35

## Leaf rust

Leaf rust resistance genes *Lr1*, *Lr3a*, *Lr13*, *Lr16*, *Lr19*, *Lr23*, *Lr24*, *Lr26*, *Lr27+Lr31*, and *Lr37* were postulated in the 22<sup>nd</sup> SAWSN (Table 2). Genes *Lr13*, *Lr27+Lr31* and *Lr3a* were present in different combinations in 22, 20 and 16 entries, respectively. Resistance in two entries could not be explained with the pathotypes used in this study. Two entries did not carry any seedling leaf rust resistance. The presence of *Lr19* was confirmed by using molecular marker (Gb)<sub>130</sub><sup>8</sup>.

**Table 2.** Frequency of leaf rust resistance genes in the 22<sup>nd</sup> SAWSN

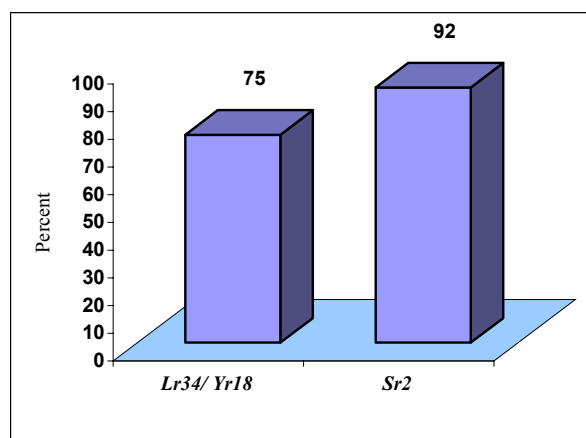
Gene	No. of Entries
<i>Lr13</i>	22
<i>Lr27+Lr31</i>	20
<i>Lr3a</i>	16
<i>Lr26</i>	12
<i>Lr23</i>	7
<i>Lr16</i>	5
<i>Lr37</i>	5
<i>Lr24</i>	4
<i>Lr19</i>	1
<i>Lr1</i>	2
Unknown	2
Nil	2

## Stem rust

Stem rust resistance genes *Sr6*, *Sr8a*, *Sr9g*, *Sr12*, *Sr17*, *Sr23*, *Sr24*, *Sr30*, *Sr31* and *Sr38* were postulated either singly or in different combinations. *Sr24* was postulated alone in three entries. Resistance genes *Sr30*, *Sr17* and *Sr8a* were postulated in 43, 38 and 31 entries, respectively, in different combinations. Resistance carried by four entries did not display differential responses against pathotypes used in this study (Table 3).

### Molecular detection of *Lr34/Yr18* and *Sr2*

Of 76 entries *Lr34/Yr18* and *Sr2*-linked markers indicated the presence of these genes in 75% (57), and 92% (70) of entries, respectively (Fig. 2).



**Fig. 2.** Percentage occurrence of *Lr34/Yr18* and *Sr2* across 22<sup>nd</sup> SAWSN

**Table 3.** Frequency of stem rust resistance genes in the 22<sup>nd</sup> SAWSN

Postulated gene	No. of entries
<i>Sr30</i>	43
<i>Sr17</i>	38
<i>Sr8a</i>	31
<i>Sr31</i>	12
<i>Sr23</i>	10
<i>Sr38</i>	7
<i>Sr24</i>	6
<i>Sr9g</i>	2
<i>Sr12</i>	1
<i>Sr6</i>	1
Unknown	4

## Adult plant responses

A relatively higher proportion (93%) of the entries exhibited low adult plant stripe rust responses (rust score 2, 3 and 4) under field conditions. This was followed by leaf rust (86%) and stem rust (68%). No entry was susceptible to stripe rust in field, whereas, some entries were moderately susceptible and susceptible to leaf rust and stem rust in the field, respectively (Table 4).

**Table 4.** Frequency of adult plant resistance responses against the three rust diseases across the 22<sup>nd</sup> SAWSN

Score	No. of Entries		
	Stripe rust	Leaf rust	Stem rust
2	64	35	31
3	6	21	11
4	1	9	10
5	5	6	11
6	-	3	10
7	-	2	2
8	-	-	1

## CONCLUSIONS

- Seedling resistance genes , *Yr6*, *Yr7*, *Yr9*, *Yr17*, *Yr27*, *Lr1*, *Lr3a*, *Lr13*, *Lr16*, *Lr19*, *Lr23*, *Lr24*, *Lr26*, *Lr27+Lr31*, *Lr37*, *Sr6*, *Sr8a*, *Sr9g*, *Sr12*, *Sr17*, *Sr23* *Sr24*, *Sr30*, *Sr31* and *Sr38* were postulated.
- Genetic diversity in term of effective seedling resistance genes appeared to be narrow.
- Adult plant resistance genes *Lr34/Yr18* and *Sr2* were present in very high proportions of entries.

## REFERENCES

- 1 Kolmer, J. A. (2005). Tracking wheat rust on a continental scale. *Current Opinion in Plant Biology*, **8**, 441-449.
- 2 McIntosh, R.A., Wellings C.R. and Park, R.F. (1995). *Wheat Rust- An Atlas of Rust Resistance Genes*, CSIRO Publications, Canberra, Australia.
- 3 Stakman, E.C., Stewart, D.M. and Loegering, W.Q. (1962). Identification of physiological races of *Puccinia graminis* var. *tritici*. United States Department of Agriculture Research Series. E617.
- 4 Luig, N.H. (1983). A survey of virulence genes in wheat stem rust, *Puccinia graminis* f.sp. *tritici*. In: Horn W, Robbelen G, eds. *Advances in Plant Breeding. Journal of Plant Breeding, Supplement*, **11**. Berlin.
- 5 Bariana, H.S., Miah, H., Brown, G.N., Willey, N and Lehmensiek, A. (2007). Molecular mapping of durable rust resistance in wheat and its implication in breeding. *Developments in Plant Breeding* **12**, 723-728.

6. Bossolini, E., Krattinger, S. G. and Keller, B. (2006). Development of simple sequence repeat markers specific for the *Lr34* resistance region of wheat using sequence information from rice and *Aegilops tauschii*. *Theoretical and Applied Genetics*, **113**, 1049-1062.
7. Hayden, M.J., Stephenson, P., Logojan, A. M., 7. Khatkar, D., Rogers, C., Elsdon, J., Koebner, R. M. D., Snape, J. W. and Sharp, P. J. (2006) Development and genetic mapping of sequence-tagged microsatellites (STMs) in bread wheat (*Triticum aestivum* L.) *Theoretical and Applied Genetics*, **113**, 1271-1281.
8. Prins, R, Groenwald, J.Z., Marias, G.F., Snape, J.W. and Koebner, R.M.D. (2001). AFLP and STS tagging of *Lr19*, a gene conferring resistance to leaf rust in wheat. *Theoretical and Applied Genetics*, **103**, 618-624.
9. Hsam, S.L.K., Mohler, V., Hartl, L.,Wenzel, G. and Zeller, F.J. (2000) Mapping of powdery mildew and rust resistance on the wheat rye translocation chromosome TIBL-IRS using molecular and biochemical markers. *Plant Breeding*, **119**, 87-89.

## ACKNOWLEDGEMENTS

We thank GRDC Australia for financial assistance through the Australian Cereal Control Program. The first author thanks the University of Sydney for the award of International Postgraduate Research Scholarship.