1 Coral community changes in the Great Barrier Reef in

2 response to major environmental changes over glacial-

interglacial timescales

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20 Abstract

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22The Ribbon Reef 5 borehole offers a unique record of reef growth spanning the entire history 23of the northern Great Barrier Reef (GBR). Previous studies have reported the main 24stratigraphical, lithological and chronological patterns, as well as basic descriptions of the coralgal assemblages, but no detailed coral community analysis was undertaken. We present 2526a quantitative analysis of the nature and distribution of Pleistocene coral communities and 27apply several statistical tools to define recurrent coral associations and compare the eight reef-building cycles recognized throughout the evolution of the GBR. The start of significant 2829reef building occurs at 137 m based on a major change in coral community structure and the inception of the reef cycles (Cy1-8). This revision, along with available stratigraphical and 30 31chronological data, suggests that barrier reef initiation may have occurred prior to MIS 11, earlier than previous reports. The coral assemblages at 137 m reflect the transition from lower 3233mesophotic (60-100 m) to upper mesophotic (30-60 m) settings, while the eight reef cycles 34above are characterized by three recurrent shallow-water reef-coral associations: Porites-Montipora-faviids (Po-Mo-Fa), pocilloporids (Poc), and Acropora-Isopora (Acro-Iso). Typically, 35

36these cycles begin with the Po-Mo-Fa association and end with the Acro-Iso association, reflecting shallowing and a catch-up growth mode. However, the first two cycles are 37characterized by a transitional phase dominated by the Poc association. The dominance of 3839pocilloporids during the early stages of the GBR's history and the long-term shift to an Acropora-Isopora-dominated community may reflect an increase in competitive pressure of 40 acroporids over pocilloporids. Our findings are consistent with the view that reef coral 41community structure is predictable over 100-kyr time scale. However, variations within reef 4243cycles highlight the importance of environmental changes operating at millennial time scales. 44Further studies are needed to better refine the reef chronology and clarify the influence of 45environmental variables (i.e. sea surface temperature, turbidity) on reef coral community 46 structure.

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58 **1. Introduction**

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60 Coral reef ecosystems are in decline worldwide (Bruno and Selig, 2007; Chin et al., 2011; De'ath et al., 2012; Jackson et al., 2014). The impact of local factors such as overfishing 6162and pollution are compounded by the global influence of ocean warming and acidification on coral health and calcification potential (Hoegh-Guldberg, 2011). However, the decline of coral 63 64 reef ecosystems is not uniform (Cinner et al., 2016) and future trends in reef community 65structure will depend on multiple factors, such as the intra- and interspecific adaptation potential of reef organisms, range shifts in coral species, and local anthropogenic factors like 66 67 overfishing and coastal development (Pandolfi et al., 2011). Information on the responses of 68 reef-corals exposed to environmental stressors such as elevated sea surface temperature 69 (SST), reduced pH or high sedimentation rate can be obtained from in vivo experiments 70involving single coral colonies (e.g. Hoadley et al., 2015; Ganase et al., 2016) or, less commonly, a portion of a reef (Dove et al., 2013; Albright et al., 2016). Large-scale reef 7172monitoring informs us on the current impact of global climate change and other factors as well 73as on the recent (few decades) history of coral reef ecosystems (e.g. Connell et al., 1997; Bak 74et al., 2005; De'ath et al., 2012). These approaches are essential to our understanding of the susceptibility of reefs and reef communities to ongoing environmental perturbations. However, 7576coral reef ecosystems are long-lived and crucial information about their long-term history and 77responses under various natural perturbations and environmental boundary conditions may be derived from the fossil record. Valuable information on the responses of coral communities 78to rapid environmental changes has been obtained from the study of Quaternary coral reef 79systems (Pandolfi and Greenstein, 2007; Camoin and Webster, 2015; Braithwaite, 2016). Of 80 particular interest are the periods of rapid warming and sea level rise leading to interglacial 81 82 highstands because these conditions may mimic those which coral reef ecosystems are 83 experiencing today. Studies of fossil reefs provide important insights into the responses of coral reef ecosystems to sea level and climate changes (Woodroffe and Webster, 2014; 84 Kiessling and Pandolfi, 2014). The Holocene and Last Interglacial records are the best 85 86 documented and most extensively studied of Quaternary reef-building episodes, and 87 improving our knowledge of older reef sequences stands as one of the major challenges of 88 fossil reef studies (Camoin and Webster, 2015).

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90 The Great Barrier Reef (GBR) is a passive margin and pre-Holocene reef limestones are not normally exposed at the surface (Hopley et al., 2007). Drilling is therefore necessary 9192to study the history of reef growth and reef communities. Accumulation of reef limestones on 93 the continental shelf occurred during interglacial periods when sea level was high enough to 94flood the shelf (Hopley et al., 2007; Davies, 2011). Reef growth there was consequently interrupted by sea level falls during glacial periods and became restricted to a narrow band 95along the shelf edge during sea level lowstands (Hopley et al., 2007; Davies, 2011). IODP 96 97 Expedition 325 drilling into the submerged terraces and ridges off the modern GBR (Fig 1), 98 combined with site survey data (Hinestrosa et al., 2016), uncovered multigenerational reef sequences developed during glacial sea level lowstands and subsequent deglacial sea level 99 100 rises (Webster et al., 2011).

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102Few boreholes have penetrated the entire section of reef limestones deposited on 103the continental shelf during previous sea level highstands. The earliest, deep drilling operations were undertaken by the Great Barrier Reef Committee in 1926 at Michaelmas Cay 104105(183 m) in the northern GBR and in 1937 at Heron Island (223 m) in the southern GBR. A 106 petroleum exploration well was drilled at Wreck Island in 1958 (575 m) in the southern GBR 107(Davies, 2011). The thickness of the entire reef section in these boreholes is interpreted to be 108< 150 m but core recovery was poor. Two additional long, high quality boreholes were drilled 109 in 1995, one at Ribbon Reef 5 (210 m) and the other at Boulder Reef (86 m), in the northern 110 GBR (Alexander et al., 2001). Ribbon Reef 5 lies near the shelf edge facing the open ocean (outer-shelf reef) whereas Boulder Reef is close to the land and influenced by terrigenous 111

sedimentation (inner-shelf reef). These reefs lie, respectively, at opposite ends of the crossshelf gradient in wave energy and turbidity, and are characterized by distinct coral
communities (Done, 1982) (Fig. 1).

- The relationship between the rate of vertical seafloor displacement (uplift vs. 116subsidence) and eustatic sea level change controls the relative position of successive reef 117sequences (Woodroffe and Webster, 2014). In relatively stable depositional settings such as 118the GBR, the resulting geometry of multigenerational reef sequences is a vertical succession 119120of stacked reefs, each formed during a sea level highstand, bounded by low sea level erosion surfaces (Davies et al., 1988, 1989). The erosion of the windward margin during sea level 121lowstands may result in the retrogradation of successive highstand reef sequences (Davies et 122123al., 1988, 1989; Webster, 1999). The highstand stack model of reef development explains the 124recurrence of shallow coralgal assemblages in the Ribbon Reef 5 cores (Webster and Davies, 1252003). Similar successions of stacked Pleistocene reef units have been described in various 126other locations, for example, at Eniwetok Atoll (Szabo et al., 1985), Mururoa Atoll (Camoin et al., 2001), the Ryukyu Islands (Sagawa et al., 2001), New Caledonia (Cabioch et al., 2008), 127128the Florida Keys (Multer et al., 2002) and Belize (Gischler, 2007).
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130Previously, only two basic coral assemblages were described in the Ribbon Reef 5 borehole (Webster, 1999; Webster and Davies, 2003). One characterized by Isopora and 131132robust branching Acropora spp. (humilis gr. and robusta gr.), the other composed of massive Porites and faviids associated with encrusting Porites and Montipora. Webster and Davies 133(2003) interpreted these coral assemblages as representative of higher- and lower-energy 134reef settings, respectively. The recurrence of apparently similar coral assemblages in the 135136Ribbon Reef borehole over at least 400 ka is consistent with detailed paleoecological studies of uplifted Pleistocene reef terraces at Barbados (Jackson, 1992; Pandolfi and Jackson, 2006), 137Huon Peninsula (Pandolfi, 1996, 1999), and, more recently, Egypt and Vanuatu (Mewis, 2016) 138139reporting the repetition of similar coral assemblages over multiple sea level highstands. These 140observations suggest a remarkable persistence in coral community structure over geological 141timescales of 10s to 100s kyr despite repeated sea level falls and major changes in SST (Pandolfi, 1996; Pandolfi, 2002). The predictable nature of reef coral assemblages may result 142from the high degree of ecological dependence of reef organisms due to their complex 143interactions (e.g. competition, predation, and symbiosis), use of available resources, and 144occupation (and re-occupation) of distinct habitats (Pandolfi, 1996; Bode et al., 2012). 145However, variability in coral species composition and diversity over glacial-interglacial 146147timescales may arise from species migration related to elevated SST. Several studies have 148presented evidence for latitudinal variations in coral species distribution related to elevated SST occurring during the last interglacial (western Australia, Greenstein and Pandolfi, 2008; 149

150at a global scale, Kiessling et al., 2012), the mid-Holocene (Florida, Precht and Aronson, 2004), and in recent times (Japan, Yamano et al., 2011; Florida, Vargas-Angel et al., 2003; and 151eastern Australia, Baird et al., 2012). Variability in reef-coral community structure also occurred 152153at a broader timescale during the Quaternary, with the abundance of Acropora increasing markedly after the Pliocene to become the dominant reef-building coral worldwide during the 154Middle Pleistocene (Renema et al., 2016). Fast growth rates and asexual reproduction via 155fragmentation are attributes of the genus Acropora which make this coral particularly suitable 156to track rapid Quaternary sea level changes (Renema et al., 2016). In addition, changes in 157158ocean surface circulation affecting larval dispersion and genetic connectivity of coral populations during the Quaternary have been proposed as a key factor controlling the 159distribution of reef-coral species (Veron, 1995). 160

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162The Ribbon Reef 5 borehole represents a rare archive of the long-term evolution of 163coral assemblages in the GBR during successive sea level highstands since the inception of 164reef growth in the Middle Pleistocene (for a recent review, see Davies, 2011). Until now the published taxonomic information was largely qualitative and intended only to highlight basic 165trends in taxonomic composition (Webster and Davies, 2003). A more quantitative analysis is 166167needed to assess the taxonomic similarity of successive coral assemblages and verify the 168hypothesis that coral communities have highly predictable taxonomic compositions during 169successive interglacial cycles. In this study we present a detailed analysis of the nature and 170distribution of coral taxa in the Ribbon Reef 5 borehole based on a new and more comprehensive taxonomic census. Several statistical approaches are used to identify 171172objectively the nature of recurrent coral assemblages with similar taxonomic compositions and reconstruct the history of coral community changes at Ribbon Reef 5 since the inception of 173174the reef system. We discuss our new results within the context of previously published trends in taxonomic composition and depositional units described in Webster and Davies (2003) and 175the depositional units described in Braithwaite et al. (2004) and Braithwaite and Montaggioni 176177(2009). Finally, we discuss the implications of our results for understanding the response of 178coral communities to individual glacio-eustatic cycles, from the colonization of bare substrate 179during shelf inundation to reef demise caused by sea level fall, and coral assembly rules at the scale of the whole history of the GBR during multiple glacio-eustatic cycles. 180

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- 182 **2. Material and methods**
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184 Ribbon Reef 5 is about 6.5 km long and 0.7–1 km wide (Fig. 1). It is one of many 185 ribbon reefs in the northern Great Barrier Reef that form a 700 km long ridge interrupted only 186 by narrow channels and forming an impressive shelf-edge barrier reef directly exposed to the 187 swells of the Pacific Ocean (Hopley et al., 2007).

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The Ribbon Reef 5 borehole (15° 22.40' S, 145° 47.149' E) is 210 m long and the recovery is 76% on average, although recoveries of 95% were common in the lower part of the main reef section (Webster, 1999; Webster and Davies, 2003). The maximum length of a core section recovered during one drill is 3 m. A total of 125 core sections (B1-B125) were drilled with an average length of 1.68 m (Fig. 2A, B).

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Three main sections have been recognized in previous publications (Fig. 2C): a basal 195196 lower section composed of grainstones and packstones (210–158 m), a middle section rich in 197 rhodoliths (158–96 m), and an upper section composed of in situ reef framework (96–0 m) (Alexander et al., 2001; boundaries revised by Webster and Davies, 2003). Webster and 198199Davies (2003) defined 6 reef units in the upper section (R1-R6) based on variations in coralgal assemblages, the occurrence of soil horizons and stable isotope (O, C) data (Fig. 2D-F). 200201Two additional reef units (R7-R8) were identified within the middle section. Braithwaite et al. 202(2004) divided the same borehole into 9 depositional units based on changes in lithology, diagenetic features, and stable isotope data (Fig. 2G-J); evidence of emergence (erosional 203surface, paleosoil) was reported between unit 6, 7, 8 and 9 in the upper section of the borehole. 204205The unit boundaries defined by Braithwaite et al. (2004) do not match those of Webster and 206Davies (2003) except for the base of the uppermost Holocene unit at 16 m (R1 in Webster and 207Davies, 2003; unit 9 in Braithwaite et al., 2004). The criteria used to define stratigraphic units 208in each publication are detailed in Table 1. The diagenetic history of the borehole was reconstructed by Braithwaite and Montaggioni (2009). Their study confirmed the correlation 209between the nature and diversity of diagenetic features and the unit boundaries defined by 210Braithwaite et al. (2004). Moreover, they found evidence for an additional erosional surface 211212within unit 6 (after Braithwaite et al. (2004)) and therefore defined two new units, i.e. 6a and 6b. Despite considerable age uncertainties, radiometric dating suggests that the main reef 213section was probably accreted during the Middle Pleistocene <500 ka (MIS 13) and the lower 214215section could be as old as 700 ka (MIS 17) (Webster and Davies, 2003; Braithwaite et al., 2162004; Braithwaite and Montaggioni, 2009; Fig. 2K). Paleomagnetic data constrain the age of 217the entire borehole to within the Bruhnes chron (<780 ka, Braithwaite et al., 2004).

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In the present study we reexamined the distribution, taxonomy and morphology of reef corals (Scleractinia) in the borehole. A total of 1332 corals were recorded. Each was categorized as either in situ (IS), probably in situ (IS?), not in situ (ISX), or context unknown (ISN) (Webster al., 2011). A combination of criteria was used to determine the context of fossil corals: (1) the overall orientation of the corallum (orientation of surface projections, branches), (2) the orientation of corallites, (3) the presence of thicker algal crusts on the upper surface of the corallum, (4) the absence of evidence of transport (abraded or broken surfaces), (5) the

226orientation of geopetal surfaces, and (6) the depositional setting (slumps, rubble) (Webster et al., 2011). Of all the corals examined, about 60% were tagged as ISX. The taxonomy of corals 227228follows Veron (2000) except for Isopora which is treated as a genus (Wallace et al., 2007). 229Corals were identified at the species level whenever possible, but the identification was more often achieved at the genus level due to the obscuring of surface features by sediments or 230encrusters, or the lack of preservation. Species groups were defined for corals belonging to 231different species within the same genus that cannot be differentiated because they display 232close morphological affinities (e.g. Favia gr. pallida includes F. pallida, F. speciosa, and F. 233234matthaii). For 17% of the corals examined, no taxonomic identification was possible; 84% of 235these unidentified corals are ISX corals. The following morphological traits were quantified: colony thickness, corallite width and branch width. Colonies attached to the substrate less 236237than 5 cm in thickness are categorized as encrusting; the others are described as massive. 238Branch width is used to classify branching corals into the following three categories: fine-239branching (BF, <1 cm), medium-branching (BM, 1-1.5 cm), and robust-branching (BR, >1.5 240cm). Additionally the maximum thickness of coralline algal crusts was also measured.

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242In order to investigate the variations of coral taxonomic composition in the borehole 243and identify recurrent coral associations, we compared the taxonomic composition of the core 244sections using cluster analysis of the Bray-Curtis similarity (BC) coefficient calculated for each pair of core sections. The analysis was performed on presence-absence data and on 245246abundance data for both the in situ (corals tagged IS and IS?) and non in situ (corals tagged 247ISX) fraction. The few corals for which the context was described as uncertain (ISN) were considered likely to be not in situ and were added into the list of non in situ corals (ISX). For 248the analysis involving abundance data, the BC coefficient was calculated using the square 249250root of abundance values in order to mitigate the influence of taxa with high count numbers (Perry et al., 2008; Roche et al., 2011). The analysis performed on in situ corals involved the 251comparison of the distribution of 37 taxa across 75 core sections. For the analysis of ISX 252253corals, the data matrix is composed of 21 taxa and 114 core sections. The lower number of 254taxa in the analysis of the ISX fraction results from the overall lower resolution of the taxonomic 255identification of allochtonous corals due to their poorer state of preservation. The results of 256these analyses enable us to identify any regularity in the sequence of corals observed in the 257borehole and highlight stratigraphic intervals displaying recurrent coral associations. 2-D MDS 258ordination of BC coefficients and analyses of similarities (ANOSIM test) are used to assess differences in taxonomic composition between these stratigraphic intervals. A similarity 259260percentage analysis (SIMPER) is conducted to constrain the identity of taxa that are 261characteristic of the recurrent coral associations. Statistical analyses were performed using 262the computer software PRIMER v6 (Clarke and Gorley, 2006). 263

3. Results

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The reexamination of the Ribbon Reef 5 borehole led to the identification of 58 266species or species groups of scleractinian corals under 34 genera (Table 2, S1). Three 267additional genera of non-scleractinian corals have also been identified (Tubipora, Millepora 268and Sinularia?). The majority of in situ corals identified belong to Acropora, Isopora, Montipora, 269270Porites, faviids and pocilloporids. There is a clear inflection point in the cumulative number of in situ corals identified in the borehole as a function of depth at around 120-125 m (Fig. 3). 271272The thickness of in situ coral colonies and coralline algal crusts increases steadily in the lower 273half of the borehole, reaching a first maximum around 117 m and 132 m, respectively, and then shows large fluctuations (Fig. 4A,C). Upcore trends in colony thickness vary among coral 274275genera. Faviids display a broadly decreasing trend in colony thickness at the scale of the 276borehole (Fig. 4A). The thickness of Porites and Montipora colonies is highly variable unlike 277that of Acropora and Isopora colonies (Fig. 4A). In addition, acroporids with robust-size 278branches become common above 130 m and increase in frequency toward the borehole top 279(Fig. 4B).

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3.1. Analysis of in situ (IS, IS?) corals

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283 3.1.1 Comparison of core sections

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Cluster analysis of the BC coefficient calculated between each pair of core sections 285286using presence-absence data produces 15 clusters (G1-G15) and two isolated core sections (54, 106) (Fig. 5A). These groups can be characterized by the systematic occurrence of one 287288or a combination of the following taxa: Acropora, Isopora, encrusting and massive Porites, encrusting and massive Montipora, pocilloporids, and faviids (Fig. 5B). A large cluster defined 289290by the association of encrusting *Porites* and *Montipora* (G12) can be further subdivided into 5 291groups of core sections (a-e) and 1 isolated core section (41). Similar results are obtained 292using the same analysis based on abundance data (Figs S2 and S3).

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There is a dramatic change in coral assemblage composition at around 137 m (Fig. 2942956A). Below that limit, the proportion of in situ corals is low and the coral assemblages are 296characterized by Galaxea (106), Goniastrea (G9), foliaceous pectiniids (most likely 297Echinophyllia, G2), encrusting Porites and/or Montipora (G11, G14, G7). Above that limit, the number of in situ corals increases sharply, and so does the diversity of coral assemblages. 298299The sequence of coral assemblages above 137 m is complex and characterized by a high 300 variability in composition at the scale of the core section. However, there is a cyclic pattern 301 that is overprinted on this apparent complexity (Fig. 6B). This cycle is defined by repeated

302intervals devoid of Acropora and characterized by encrusting and massive Porites, encrusting Montipora, and diverse faviids (mainly G12e). These intervals are systematically followed by 303 304 intervals enriched in Acropora and/or Isopora (G5, G3, G12b, G12c and G12d). The transition 305is in places marked by the occurrence of a pocilloporid-rich interval (mainly G4). We therefore conclude that one complete cycle is defined by the following succession of coral associations: 306 a Porites-Montipora-faviids association (Po-Mo-Fa), a pocilloporid association (Poc), and an 307308Acropora-Isopora association (Acro-Iso) (Fig. 6A, B). We identify 8 cycles in total based on these recurrent shifts in composition (Cy1-Cy8) overlying the lower unit (Lw) (Fig. 6B). Two of 309 310 these cycles (Cy8 and Cy7) are composed of all three coral associations. The Poc association is missing in 4 cycles (Cy6, Cy5, Cy4 and Cy1) and its presence at the base of Cy2 is uncertain 311due to the sparse occurrences of pocilloporids in this interval. The Po-Mo-Fa association is 312313only absent in Cy3 (and perhaps Cy2), and is poorly developed in Cy8. Interestingly, the Po-Mo-Fa interval is thickest in Cy7 and becomes progressively thinner in subsequent cycles. 314315Leptoseris occurs at the base of the Po-Mo-Fa interval in Cv7 and Cv6 and coexists with 316 Pachyseris speciosa in Cy7. The Po-Mo-Fa interval in Cy6 also contains several colonies of 317Galaxea astreata. Massive Isopora occurs in the Po-Mo-Fa interval in Cy4 only. The Poc intervals in Cy7 and Cy6 are well developed and characterized mostly by Stylophora, in 318319association with Seriatopora and an Echinopora-rich assemblage (G13) in Cy6. The Poc 320association at the base of Cy3 is also composed of Stylophora. The species composition of 321Acropora in the Acro-Iso association is variable from one cycle to another and includes the 322following major taxonomic groups: A. gr. humilis, A. gr robusta?, A. gr. formosa and various 323fine-branching morphological groups (corymbose, digitate, arborescent and platy). Isopora colonies display encrusting, massive or robust-branching growth forms. Acropora and Isopora 324may coexist or else may appear in sequence. In two cycles, Cy4 and Cy6, the shift from 325326Isopora- to Acropora-dominated assemblages is particularly clear. The Acro-Iso association includes Porites and Montipora as well as diverse faviids in Cy6, Cy4 and Cy1. 327

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In summary, comparison of the taxonomic compositions of core sections shows (1) high variability in taxonomic composition from one core to the next, (2) a regular pattern of taxonomic composition at the scale of several core sections characterized by the repetition of three main coral associations (Po-Mo-Fa, Poc and Acro-Iso), and (3) two main units at the scale of the whole borehole – a non-reefal unit with low density, mostly encrusting or foliaceous coral assemblages changing at around 137 m to a reefal unit with high-density, high-diversity coral assemblages displaying cyclicities in taxonomic composition.

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337 3.1.2. Comparison of reef cycles (Cy1-Cy8) and the lower unit (Lw)

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339 Cluster analysis of the BC coefficient calculated between each pair of cycles using

340 presence-absence data reveals two clusters: cluster 1 composed of cycles Cy8, Cy3 and Cy2, and cluster 2 composed of cycles Cy4, Cy5, Cy6 and Cy7 (Fig. 7A). The 2D MDS plot clearly 341shows the two groups of data points occupying distinct regions of space (Fig. 7B). The 342343difference in taxonomic composition between clusters 1 and 2 is confirmed by the ANOSIM test which gives an R value of +0.80 (p=2.9%). The average BC similarities of the cycles in 344clusters 1 and 2 are 61.9% and 66.5%, respectively (Fig. 7C). The taxonomic diversity of the 345346cycles in cluster 1 is lower than that of those in cluster 2. The SIMPER analysis shows that cluster 1 is characterized by robust-branching Acropora, encrusting and massive Porites, and 347348 Stylophora, whereas cluster 2 is defined by medium-branching Acropora, encrusting to 349 massive Isopora, encrusting to massive Porites and Montipora, Stylophora, Favia, and pectiniids (mainly Echinophyllia) (Table 3A). Cy1 is more closely related to cluster 2 than 350cluster 1 and the lower unit of the borehole (Lw) is clearly distinct from the reef cycles based 351352on presence-absence data.

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354 Cluster analysis based on the square root of abundance data provides a slightly 355 different result. The same cluster 2 is produced but Cy2 is grouped with Cy1 instead of with 356 Cy3 and Cy8. Cycles Cy1 and Cy2 have in common the following taxa: fine-branching 357 *Acropora*, encrusting to massive *Porites* and *Montipora*, and *Favites*. The lower unit (Lw) is 358 grouped with Cy1 and Cy2 due to similarities in abundances of encrusting *Montipora*, and 359 encrusting and massive *Porites*.

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361 3.1.3. Comparison of coral associations (Po-Mo-Fa, Poc and Acro-Iso)

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Cluster analysis and 2D MDS ordination of BC similarities were also used to compare 363364 the taxonomic compositions of the three coral associations (Po-Mo-Fa, Poc and Acro-Iso) among reef cycles and with the lower unit (Lw). The clustering reflects the separation between 365the three previously identified coral associations (Fig. 8A). Cluster 1 is composed of two of the 366 367 four Poc intervals, and clusters 2 and 3 mostly include Po-Mo-Fa and Acro-Iso intervals, 368 respectively. However, there is some overlap as the clustering is not entirely determined by 369 the type of association to which each interval is assigned. The 2D MDS plot also shows that the three associations define three overlapping domains (Fig. 8B). The ANOSIM test suggests 370 371there is a significant difference in taxonomic composition between the three coral associations 372(R = +0.34). The SIMPER analysis highlights the corals that are the most representative of 373each association and the results are entirely consistent with the description presented above (Table 3B). Massive to encrusting Porites and encrusting Montipora are characteristic of both 374375associations Po-Mo-Fa and Acro-Iso but the latter is distinguished primarily by the presence 376 of diverse Acropora species. Stylophora is indicative of the Poc association. The average BC similarities of the intervals within each association are widely different, i.e. 50.3%, 37.7% and 377

23.2% for associations Acro-Iso, Po-Mo-Fa and Poc, respectively (Fig. 8C). This variability is
illustrated by the spread of the data points associated with each domain in the 2D MDS plot.
The association Acro-Iso displays the compositions that are the most consistent among
different stratigraphic intervals. The composition of the lower unit (Lw) is most similar to that
of the Po-Mo-Fa association of Cy6.

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The separation between the three coral associations is less clear when the square root of abundance data is used for cluster analysis. This is mostly due to the influence of encrusting *Porites* and *Montipora* which can be equally abundant in intervals assigned to different coral associations so that these intervals are more closely related in the cluster diagram when abundance data are used.

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390 3.2. Analysis of non-in situ (ISX) corals

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The number and proportion of non-in situ corals is largest in the lower unit of the borehole below 137 m. The proportion of non-in situ corals that could be identified is lower than in the in situ fraction because non-in situ corals have commonly undergone some degree of physical alteration that tends to reduce the size of individual specimens and erase morphological details. The taxonomic composition of the non-in situ fraction is clearly dominated by *Acropora* and pocilloporids. Together these taxa represent 37-84% of the number of ISX corals identified in each cycle.

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400 Cluster analysis of the BC coefficient calculated between each pair of core sections using presence-absence data for the ISX fraction leads to the segregation of 12 groups (Fig. 4014029). The occurrence of Acropora or pocilloporids together or separately clearly determines the clustering of most of the core sections. For example the large cluster G7 is defined primarily 403 by the association of Acropora and pocilloporids whereas G11 is defined on the basis of the 404405absence of all taxa but Acropora. Like the in situ corals, the composition of the lower detrital 406 unit is distinct from that of the overlying reefal unit. The ISX coral fraction in the lower unit is 407 characterized by pocilloporids, encrusting Porites and Montipora, and lesser agariciids (mostly Leptoseris) and pectiniids (mostly Echinophyllia) (Fig. 10). The main difference with the in situ 408 fraction is the abundance of pocilloporids and agariciids. Interestingly, there is a difference in 409 410 the generic composition of pocilloporids in the ISX fraction between the lower and upper unit: Seriatopora occurs more consistently in core sections of the lower unit whereas Stylophora 411 and Pocillopora are generally dominant in the overlying reefal unit (with the notable exception 412413of Cy7) (Fig. 10). There is also a significant increase in the frequency of occurrence and 414relative abundance of Acropora and pocilloporids in the reefal unit and most of the ISX coral assemblages defined by cluster analysis occurring in the main reefal unit are characterized by 415

416 a combination of pocilloporids and *Acropora* (Figs. 9, 10).

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The cyclic pattern observed in the reefal unit above 137 m is not clearly reflected in the ISX fraction. However, there are variations in the ISX fraction that coincide with changes in the compositions of in situ corals. The transition between the Po-Mo-Fa and Poc associations in Cy8 and Cy7 coincides with an increase in the abundance of pocilloporids in the ISX fraction (Fig. 10). In addition, the Po-Mo-Fa intervals in Cy7 and Cy6 correlate with a marked increase in the abundance of non-in situ *Porites* and/or *Montipora* (Fig. 10).

- 424
- 425 **4. Discussion**
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427 4.1. Long-term trends in coral community structure

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429Variations in the abundance, taxonomy and morphology of in situ corals in the Ribbon Reef 5 borehole indicate a general shallowing upward trend and a transition from a relatively 430 431deep, non-reefal setting to cycles of reef growth and demise, in agreement with previously published works (Alexander et al., 2001; Webster and Davies, 2003; Braithwaite et al., 2004; 432433Braga and Aguirre, 2004). We identify the start of the major reef-building processes at 137 m 434based on: (1) a significant change in coral community structure and, (2) the inception of a cyclic pattern in coral succession which characterizes the upper part of the borehole. This 435boundary corresponds with the base of the first reef unit (i.e. R8) originally described in 436437Webster and Davies (2003).

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The in situ coral assemblage below 162 m is dominated by thin laminar Porites, 439440 Montipora and Echinophyllia, also occurring in the non in situ fraction with Leptoseris. The taxonomy and morphology of these corals and their association with Melobesioid algae (Braga 441 442and Aguirre, 2004) suggest a depositional setting deeper than 60 m (Abbey et al., 2013). Fine 443branching Seriatopora is common among coral clasts and could be derived from the upper 444mesophotic zone (30-60 m). Seriatopora is currently found down to 55 m in the GBR (Bridge 445et al., 2012) and forms dense populations in the upper mesophotic zone in the Ryukyu Islands (Sinniger et al., 2013; Sinniger, 2016). The transition at 162 m to recurrent Lithophylloid-rich 446 447intervals coincides with a marked increase in coral colony thickness (Fig. 4). Abbey et al. 448 (2013) reported a similar shift in coral thickness at 60 m in the GBR with modern and postglacial mesophotic corals primarily thin platy to encrusting (less than 2 cm in thickness) 449450below 60 m and more massive above 60 m. In addition, faviids, particularly Echinopora, become more common in the ISX fraction above 162 m in the Ribbon Reef 5 borehole. Faviids, 451452in particular *Echinopora*, were reported in the mesophotic zone from depths <60 m by Abbey et al. (2013). In conclusion, the coralgal assemblages in the lower section of the Ribbon Reef 453

5 borehole are similar to the mesophotic coralgal communities of the modern GBR and the
evolution in taxonomic and morphological composition indicates the transition from a lower
mesophotic (60-100 m) to an upper mesophotic (30-60 m) coralgal assemblage.

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Above 137 m the thicknesses of corals and algae and the numbers and diversity of 458corals increase sharply. The 137 m boundary also marks a sharp increase in the abundance 459460 of branching Isopora and Acropora species (Figs. 4, 11), two major components of modern shallow exposed reef crest communities of outer shelf reefs in the GBR (Done, 1982). The 461462 evolution of coralgal assemblages in the Ribbon Reef 5 borehole clearly indicates a general 463shallowing trend initiated in the lower section (Webster and Davies, 2003; Braga and Aguirre, 2004). The Ribbon Reef 5 record suggests that Isopora and Acropora were important 464contributors to the reef-building process from the beginning of the reef growth history since 465the Acro-Iso association is present in the initial cycle (Cy8) and was reestablished during each 466 467subsequent cycle. This observation is in line with a recent compilation showing that Acropora became a dominant reef builder during the Middle Pleistocene, and this success has been 468469attributed to the ability of Acropora to cope with rapid sea level rise (Renema et al., 2016). In the modern GBR Isopora is closely associated with Acropora in wave-exposed habitats (Done, 4704711982) and our record shows that this was also the case during the past 3-5 interglacial 472highstands (Webster, 1999; Braithwaite and Montaggioni, 2009), indicating that these two taxa 473have played a key role in not only the initiation but also the long-term maintenance of the 474barrier reef ecosystem during successive rapid post-glacial sea level rises.

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476 4.2. Barrier reef initiation

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478Drilling into submerged fossil reefs along the shelf edge of the modern GBR has demonstrated that lowstand and early deglacial reef communities may have been important 479sources of coral propagules to support the early colonization of the newly inundated shelf 480 481 (Webster et al., 2011). In the absence of a precursor lowstand reef ecosystem coral 482propagules may have been initially supplied by nearby non-reefal shallow communities. The 483 morphology and taxonomy of the ISX corals in the lower section of the Ribbon Reef 5 borehole, mostly laminar Porites, Montipora, agariciids and fine-branching Seriatopora, suggests that 484485the majority of coral clasts were derived from nearby deep mesophotic communities and no 486 clear signature of shallow water coral reef communities is recognized. Another possibility involves a more distant source of propagules, either from locations farther north, i.e. the Torres 487488 Shelf or the Gulf of Papua, where reefs may have already been established (Davies et al., 1991), or from the Queensland Plateau (McKenzie et al., 1993). The latter seems more likely 489 490 given the present-day configuration of ocean currents with the westward-flowing South Equatorial Current dividing around Cairns into the northward-flowing North Queensland 491

492 Current and the southward-flowing East Australian Current (Choukroun et al., 2010). However,
493 simulations of larval dispersal trajectories show that the connectivity between Papua New
494 Guinea and the GBR may be enhanced during La Niña events (Treml et al., 2008).

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The timing of barrier reef initiation is still poorly constrained. The warm SST's and 496high-amplitude sea level during the MIS 11 interglacial have been proposed as triggers for the 497initiation of extensive barrier reef growth worldwide, including the GBR (Droxler et al., 2003; 498Montaggioni et al., 2011; Droxler and Jory, 2013). This view has been challenged by 499500sedimentological studies of distal boreholes from the upper slope of the Marion Plateau 501placing the onset of the southern central GBR earlier, during MIS 15 (Dubois et al., 2008) or MIS 17 (Obrochta, MS 2004). Given that alkenone SST reconstructions in the region indicate 502503MIS 11 was not particularly warm (Lawrence and Herbert, 2005), the claim that barrier reef initiation on the Eastern Australian Shelf occurred at MIS 11 needs to be reassessed. A similar 504505pre-MIS 11 age (MIS 15 or MIS 13) has been suggested for reef initiation on the Western 506Australian Shelf and linked to the strengthening of the Leeuwin Current as well as increased aridity and alkalinity (Gallagher et al., 2014). Higher-amplitude sea level oscillations after the 507Mid-Pleistocene Transition (Elderfield et al., 2012) have also been proposed as a possible 508509trigger for extensive barrier reef growth on the Eastern Australian Shelf through prolonged 510shelf flooding (Dubois et al., 2008) and the creation of a cyclone corridor reducing terrigenous sediment input on the outer shelf (Larcombe and Carter, 2004). Our analysis of the coral 511512succession in the Ribbon Reef 5 borehole, combined with previously published data on 513coralline algae (Braga and Aguirre, 2004) and the currently available age data, suggest the emergence of a cyclic pattern of reef growth and demise took place prior to MIS 11. Based on 514the chronology of the Ribbon Reef 5 borehole presented in Montaggioni et al. (2011), reef 515516initiation would therefore have occurred during MIS 15, in agreement with Dubois et al. (2008). 517

- 518 4.3. Environmental interpretation of the coral associations
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520The Ribbon Reef 5 borehole above 137 m is characterized by a cyclic pattern in the 521coral community structure. Each cycle is composed of up to three coral associations (Po-Mo-Fa, Poc and Acro-Iso). The Po-Mo-Fa association, dominated by encrusting to massive 522523Porites and encrusting Montipora associated with faviids and agariciids, indicates a relatively 524deep, mid to lower reef slope habitat or a turbid environment (Done, 1982; Veron, 2000). 525Porites and Montipora are typical of highly turbid carbonate environments (Sanders and 526Baron-Szabo, 2005). The occurrence of Lithophylloid and Melobesioid algae in Cy7, with this 527coral association may indicate more temperate climatic conditions (Braga and Aguirre, 2004). 528The pioneer Po-Mo-Fa association is replaced by the Poc association in Cy8 and Cy7 which 529consists primarily of Stylophora pistillata, associated with Seriatopora hystrix and faviids

530(mostly branching Echinopora) only in Cy7. Stylophora pistillata and Seriatopora hystrix have a broad range of distribution, from shallow to deep (>30 m) reef habitats with Stylophora 531pistillata locally dominant on exposed reef fronts and Seriatopora hystrix typical of reef flats 532(Done, 1982; Veron, 2000; Bridge et al., 2012; Abbey et al., 2013). Dense communities of 533Seriatopora hystrix have been reported from the upper mesophotic zone in the Ryukyu Islands 534(Sinniger et al., 2013; Sinniger, 2016). The association of Echinopora (probably E. 535mammiformis) and Seriatopora hystrix in Cy7 indicates a low-energy upper reef slope or 536lagoonal setting (Done, 1982; Veron, 2000). The lithology in this interval is best described as 537538a floatstone with bioclasts heavily encrusted by Lithophylloid algae (Rhodolith unit 1 of 539Webster and Davies, 2003), in concordance with a hydrodynamic regime characterized with low-wave exposure. The Acro-Iso association, composed of Isopora and medium-size to 540robust branching Acropora gr. humilis and Acropora gr. robusta, indicates a shallow exposed 541(<10 m) reef setting in the GBR (Done, 1982) and other Indo-Pacific reefs (Cabioch et al., 5425431999; Hongo and Kayanne, 2010; Hongo, 2012). In contrast, the assemblage characterized by arborescent Acropora gr. formosa between 90-95 m in Cy7 is typically found in the modern 544545GBR in lower-energy habitats on reef slopes or in lagoons (Done, 1982; Oliver et al., 1983; Wallace, 1999). 546

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548Notable shifts in taxonomic composition occur within the Acro-Iso association and consist of transitions between Isopora-dominated and Acropora-dominated intervals. In Cv7, 549the transition from arborescent Acropora gr. formosa to Isopora was discussed above and 550interpreted as a shallowing-upward trend. In Cy6 and Cy4, the transition from Isopora-551dominated to Acropora-dominated intervals does not seem to reflect any major change in 552paleowater depth as both assemblages are associated with shallow Mastophoroid algae. 553554Interestingly, the shift in these two cycles coincides with unit boundaries defined in Braithwaite et al. (2004). Transitions between Acropora-dominated and Isopora dominated intervals are 555556observed in Holocene core sections from the southern GBR (Dechnik et al., 2015). In particular, 557the replacement of an Acropora assemblage by an Isopora assemblage on the windward margin of the Fitzroy reef was interpreted as reflecting a minor shallowing within the 0-10 m 558559depth range. There is however no clear evidence, based on associated species and algal crust 560thicknesses, that the two assemblages in the Ribbon Reef 5 borehole occupied distinct reef 561zones.

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563 4.4 Interpretation of reef cycles

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565 The repeating cycles observed in the Ribbon Reef 5 borehole begins with the Po-Mo-566 Fa association and terminates with the Acro-Iso association with or without a transitional 567 phase characterized by the Poc association. The repeated cycles of coral succession 568observed in Ribbon Reef 5 are broadly similar to Holocene sequences described in the GBR, characterized by a pioneer assemblage of massive Porites and faviids replaced by a high-569energy Acropora-Isopora assemblage (Montaggioni, 2005; Dechnik et al., 2015). This type of 570571coral succession is common in the Holocene and interpreted as a shallowing-upward sequence reflecting a catch-up mode of reef growth (Montaggioni and Braithwaite, 2009). In 572a similar fashion, the Po-Mo-Fa association in the successive reef-building episodes observed 573574at Ribbon Reef 5 likely represents the first stage of colonization of the shelf after inundation during each post-glacial sea level rise, perhaps also influenced by lower SSTs based on the 575576occurrence of Lithophylloid and Melobesioid algae (Braga and Aguirre, 2004). The first of 577these pioneer corals in the Ribbon Reef 5 are colonies of Goniastrea (probably G. edwardsi) at the base of Cv8. Several species of Goniastrea, including G. edwardsi, are stress-tolerant, 578579particularly to sedimentation and turbidity (Perry and Smithers, 2009; Darling et al., 2012), and 580therefore may be particularly tolerant to sediments resuspended during shelf inundation. 581Subsequent Po-Mo-Fa intervals are stratigraphically thicker and more diverse, characteristics 582which may reflect the increase in diversity of the regional species pool and/or higher amplitudes of sea level rise. Greater accommodation space created by higher-amplitude sea 583level rise could have led to thicker vertical accumulations of the initial Po-Mo-Fa assemblage 584585but this remains to be explored.

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The Poc association precedes the Acro-Iso association in three reef cycles, i.e. Cv8, 587Cy7 and Cy3. In Cy8, a Stylophora-Mastophoroid assemblage is overlain by an Acropora gr. 588humilis-Mastophoroid assemblage. Both indicate a shallow exposed (<10 m) reef setting. A 589similar interpretation can be proposed for the coral succession in Cy3. In Cy7, a Stylophora-590Seriatopora and Lithophylloid assemblage is replaced by an Acropora gr. formosa and 591592Lithophylloid assemblage, which is in turn replaced by an Isopora and Mastophoroid assemblage. The coralgal succession in Cy7 suggests a general shallowing-upward sequence. 593The transition to Mastophoroid algae could also reflect higher SST's (Braga and Aguirre, 2004). 594595However, it is not clear whether the transition between the first two assemblages results from 596decreasing paleowater depths as their distribution range overlaps. In conclusion, the transition 597from the Poc association to the Acro-Iso association in Cy8, Cy7 and Cy3 does not necessarily 598reflect any major change in depositional setting or environmental conditions and may result 599from variations in community structure within the same habitat caused by lateral reef accretion 600 (Dechnik et al., 2016; Webb et al., 2016; Webster and Davies, 2003). However, this hypothesis 601 does not explain the dominance of the Poc association in the first two reef cycles and the 602 subsequent long-term shift to acroporid-dominated branching coral assemblages. An 603 alternative explanation is that this transition reflects an ecological succession driven by 604 intrinsic ecological factors. Two reasons may explain the success of Stylophora and 605Seriatopora during the early phase of the GBR evolution. First, the pool of Acropora and

606 Isopora colonies may have been less extensive during the early phase of the GBR history. The newly formed barrier reef may not have been as extensive as it is today due to the 607 608 absence of an antecedent barrier reef to support further reef growth (Larcombe and Carter, 609 2004). The presence of well-developed Rhodolith facies (Rd1 and Rd2; Webster and Davies, 610 2003) in Cy7 supports this interpretation. Furthermore, the less abundant and patchier 611 populations of fast-growing acroporids may have given an opportunity to pocilloporids to 612 colonize the substrate first. This hypothesis is supported by the general upward increase in 613 Acropora abundance at Ribbon Reef 5 (Fig. 11). Both Stylophora pistillata and Seriatopora 614 hystrix are fast-growing corals (Pratchett et al., 2015), release fertilized larvae (Hugues et al., 6151999), and have a broad depth distribution range in the GBR (Done, 1982, Bridge et al., 2012). They are successful early colonizers of denuded substrates after disturbance (Loya, 1976; 616 617 Fan and Dai, 1996) and have been described as "weedy" species (Darling et al., 2012). 618 Stylophora pistillata and Seriatopora hystrix are therefore ideal candidates to rapidly colonize 619 available substrates during rapid environmental and sea level changes, in exposed and 620 protected settings, respectively. The second reason is that environmental conditions early in 621 the GBR history may have favored opportunistic species like S. pistillata and S. hystrix. The 622 lack of a fully developed barrier reef system may have exposed the outer shelf reefs to more 623storm-induced disturbances and terrigenous influence (Larcombe and Carter, 2004), favoring 624opportunistic species like S. pistillata and S. hystrix. This would also explain the paucity of reef 625framework below 85 m despite the inception of the reef cycles much earlier (Webster et al., 6262003; Braithwaite et al., 2004).

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628 4.5. Comparison of reef cycles

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630 The remarkable cycles of coral community structure observed in the Ribbon Reef 5 borehole and the repetition of similar types of coral associations in successive generations of 631 632reefs is consistent with records of high persistence in coral community structure during successive highstands reported elsewhere (Jackson, 1992; Pandolfi, 1996; Pandolfi and 633 634 Jackson, 2006). Our analysis shows high similarity in taxonomic compositions between 635successive cycles, particularly between Cy4, Cy5, Cy6 and Cy7 that are also the cycles with the highest taxonomic diversity. However, the Ribbon Reef 5 record also shows that the 636 637competition between pocilloporids and acroporids may have caused variations in the relative 638abundance of these taxa during the history of the GBR. In addition, some of the cycles 639 identified, specifically Cy4, Cy5 and Cy6, may represent more than one highstand episode 640 based on the previous interpretation of the reef units in the same borehole (Webster, 1999). 641 Braithwaite and Montaggioni (2009) also identified two subaerial erosion surfaces at 36 m and 64257 m, respectively. We do not see any persistent change in reef setting associated with these 643events. However, it is possible that the initial coral association after the reflooding of the shelf was an Acro-Iso association indicating a shallow exposed reef setting similar to that of theAcro-Iso association terminating the preceding cycle.

646

647The similarities among intervals characterized by the same types of coral association 648 (Po-Mo-Fa, Poc or Acro-Iso) are variable. Intervals displaying the highest similarities are characterized by the Acro-Iso association. Except for the interval in Cy7, the Acro-Iso 649 650association probably represents an aggrading phase of reef growth in a shallow exposed 651environment under relatively stable hydrodynamic conditions that may have promoted higher 652similarities in taxonomic composition. Conversely, the Po-Mo-Fa association represents the 653initial phase of reef development during sea level rise and initial conditions on the submerged shelf may have been more unpredictable (Dechnik et al., 2015). Furthermore the composition 654of the initial coral association depends on the pool of coral species available to recolonize the 655shelf that is, in turn, controlled by environmental conditions during the preceding lowstand and 656657subsequent sea level rise. The higher variability in taxonomic composition among the Poc intervals may reflect the contrasting paleoenvironmental interpretations assigned to the Poc 658659interval in Cy7 (low-energy) and those of Cy8 and Cy3 (high-energy).

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661 Trends in colony thickness across reef cycles also vary from one taxon to another. 662 Some trends underline similarities in community structure between intervals from different reef 663 cycles. For example, in most cycles thicker *Porites* colonies are systematically associated with 664the first phase of reef growth, i.e. the Po-Mo-Fa intervals (Fig. 4A). Massive Porites associated 665with faviids typically occurs in a semi-exposed or sheltered environment in Indo-Pacific reefs consistent with our interpretation of the Po-Mo-Fa association (Montaggioni, 2005). In addition, 666 peaks in Montipora colony thickness coincide with Acro-Iso intervals (Fig 4A). Massive 667 668 Montipora is a major component of some shallow (<10 m) reef facies and agrees well with our interpretation of the Acro-Iso association (Camoin et al., 2012). Conversely, the colony 669 670 thickness of faviids does not seem to be determined by the coral association, but rather shows 671 a long-term decline across multiple reef cycles (Fig. 4A). This is correlated with a notable 672 increase in the frequency of acroporids with robust branches toward the top of the borehole 673 (Fig. 4B). One possible explanation for this would be an incremental increase in wave exposure due to the progressive retrogradation of reef sequences over multiple sea level 674675highstands (Davies et al., 1988, 1989; Webster, 1999). In addition, the fast-growing acroporids 676 may have had a long-term competitive advantage over slow-growing faviids (Renema et al., 677 2016).

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Based on our analysis of the nature and distribution of coral taxa in the Ribbon Reef
5 borehole, we have identified patterns of coral community changes at varied time scales (Fig.
12). Superimposed on the long-term cycles of reef initiation and demise are variations in

682 community structure within each cycle, represented by successive stratigraphic intervals with distinct coral associations as well as short-term variations in community structure within these 683 intervals. Coral succession in each cycle is clearly influenced by glacial-interglacial sea level 684 685changes that ultimately control accommodation space and reef growth. In addition, millennialscale environmental changes, such as variations in SST and turbidity, associated with 686 deglacial conditions and shelf inundation are likely to influence coral community structure. 687 688 However, the importance of these factors is still poorly constrained. A key objective of future studies will therefore be to quantify these environmental variables and analyze their 689 690 relationship with the observed patterns of coral community changes. Promising techniques 691 include laser abrasion ICP-MS U/Th measurements to improve reef chronologies and novel geochemical proxies (REE, lithium isotopes) to reconstruct terrigenous sediment influx. 692

693

694 **5. Conclusions**

695

696 Our detailed study of the taxonomy and distribution of coral taxa in the Ribbon Reef 5 697 borehole, combined with previously published records, leads to the following conclusions:

- The pattern of repeated reef growth and demise starts at 137 m. Available
 stratigraphical and chronological data suggest that barrier reef initiation may have
 occurred prior to MIS 11. The non-reefal section, below 137 m, shows a transition from
 a lower to an upper mesophotic coral assemblage, replaced in turn by the initial coral
 association of the first reef cycle at 137 m, indicating a shallowing-upward trend.
- Three coral associations are identified in the reefal section: (1) encrusting to massive *Porites*, encrusting *Montipora* and faviids (Po-Mo-Fa) mid to lower reef slope habitat
 or turbid environment, (2) pocilloporids (Poc) shallow exposed reef fronts (*Stylophora pistillata*) and protected settings (*Seriatopora hystrix*), and (3) massive or branching *Isopora* and medium to robust-branching *Acropora* (Acro-Iso) shallow exposed (<10
 m) reef settings (*Isopora-Acropora* gr. *humilis-Acropora* gr. *robusta*) and lower-energy
 reef-slope or lagoonal settings (*Acropora* gr. *formosa*).
- 3. Eight reef cycles are identified. Each typically starts with the Po-Mo-Fa association
 (pioneer) and ends with the Acro-Iso association. We interpret this as a shallowingupward sequence following shelf inundation during each successive post-glacial sea
 level rise and highstand. Poor water quality due to sediment resuspension and/or lower
 SST's may also have influenced the pioneer association.
- The Poc association is prominent in the first two reef cycles (Cy8, Cy7) and represents
 a transitional phase between the Po-Mo-Fa and Acro-Iso associations. The singular
 predominance of pocilloporids during the early stages (Cy8, Cy7) of reef history and
 the long-term shift to an *Acropora-Isopora*-dominated shallow reef community may be
 attributed to an increase in competitive pressure of acroporids over pocilloporids within

the same reef habitat.

- 5. The recurrence of broadly similar coral associations throughout multiple interglacial highstands is consistent with the view that reef coral community structure is predictable over 100-kyr time scales. However, variations within reef units indicate that other important processes are also operating on millennial scales, particularly during initial stages of shelf reflooding, causing differences in water clarity, SST, or the pool of coral species available to reseed the newly inundated substrate.
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1007 Figure and Table captions

1008

1009Figure 1: (A) Map showing the location of Ribbon Reef 5 and Boulder Reef in the northern 1010 Great Barrier Reef and (B) detailed bathymetric map of Ribbon Reef 5 based on Laser 1011 Airborne Depth Sounder (LADS) (0-40 m) and multibeam echo sounder (40-250 m) data 1012 (Beaman et al., 2008). The yellow and white sticks show the approximate location of the deep 1013 Ribbon Reef 5 bore hole and the shorter (<20 m) Holocene cores (Davies and Montaggioni, 1014 1985). (C) Schematic profile of the continental shelf showing the broad geologic setting of the 1015 onshore boreholes Ribbon Reef 5 and Boulder Reef and the offshore IODP 325 boreholes 1016 (Webster et al., 2011).

1017

Figure 2: (A) Representation of the core sections (1-125) of the Ribbon Reef 5 borehole with
(B) core recovery. The basic subdivisions of the borehole defined in previous publications are
presented in the other columns. (C) The three main sections of the borehole with boundaries
from Webster and Davies (2003). (D-F) Depositional reef units (R1-R8) and corresponding
variations in lithology and coralgal assemblages as described in Webster and Davies (2003).
(G) Depositional units (U1-U9) and lithological variations (H), reported in Braithwaite et al.

- 1024(2004) (with U6 divided into U6a and U6b in Braithwaite and Montaggioni, 2009), δ^{18} O and1025 δ^{13} C profiles (I, J) and age data (K) published in Braithwaite et al. (2004).
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Figure 3: Cumulative numbers of corals as a function of depth (time). Note that the cumulativenumber of non in situ corals (ISX) is an underestimation.

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Figure 4: Bar charts showing the variations in some key morphological characteristics of corals and coralline algae with depth: (A) thickness of coral colonies, (B) branch width of *Acropora* and *Isopora* and (C) thickness of coralline algal crusts.

1033

Figure 5: (A) Group-averaged hierarchical clustering of the BC coefficient calculated between
 core sections based on taxonomic presence-absence data for in situ corals; (B) two-way table
 showing the taxa indicative of each cluster (coral assemblage).

1037

Figure 6: (A) Column showing the succession of in situ (IS, IS?) coral assemblages defined by cluster analysis (using the same colour code as in Figure 5) and distribution of major taxonomic groups in the borehole; (B) succession of coral associations (Po-Mo-Fa, Poc and Acro-Iso) and reef cycles (Cy1-Cy8) defined on the basis of the sequence of coral taxa presented in this study; (C) comparison of the depositional units of Webster and Davies (2003) and of Braithwaite et al. (2004) (D) (modified by Braithwaite and Montaggioni, 2009).

1044

Figure 7: (A) Group-averaged hierarchical clustering of the Bray-Curtis similarity coefficient calculated between each reef cycles (Cy1-Cy8) and the lower unit (Lw); (B) the same analysis but represented as a 2D MDS plot of the BC coefficient with the two clusters identified in (A) circled in black; (C) the average similarity between reef cycles in each cluster and the results of the ANOSIM test.

1050

Figure 8: (A) Group-averaged hierarchical clustering of the Bray-Curtis similarity coefficient calculated between borehole intervals assigned to one of the three coral associations (Po-Mo-Fa, Poc and Acro-Iso) and the lower unit (Lw); (B) the same analysis but represented as a 2D MDS plot of the BC coefficient with colours identifying each interval as belonging to the association Po-Mo-Fa (black), Poc (orange), Acro-Iso (blue) and the lower unit (purple); (C) the average similarity within each group of intervals assigned to a coral association and the results of the ANOSIM test.

1058

1059Figure 9: (A) Group-averaged hierarchical clustering of the BC coefficient calculated between1060core sections based on taxonomic presence-absence data for non-in situ (ISX) corals; (B) two-

1061 way table showing the taxa indicative of each cluster (coral assemblage).

1062

Figure 10: Columns showing (A) the succession of non in situ (ISX) coral assemblages defined by cluster analysis (using the same colour code as in Figure 8), (B) the distribution of coral associations described in this paper and (C) the succession of reef cycles against stacked area charts showing the variations in the number of non-in situ corals identified in each core section for various major coral taxa.

1068

Figure 11: Bar chart of the number of *Acropora* recorded in each core section for in situ (IS) and non in situ (ISX) specimens. The asterisks indicate core sections in which the number of non-in situ *Acropora* was likely underestimated. (A) Above the bar chart are displayed the available chronologic data for the Ribbon Reef 5 borehole, (B) the reef cycles and coral associations defined in this paper, (C) the depositional units published in Braithwaite and Montaggioni (2009), and (D) those defined by Webster and Davies (2003).

1075

1076 Figure 12: Schematic representation of the coral associations and their successions in the 1077reef cycles identified in the Ribbon Reef 5 borehole, and selected images of coral colonies typical of each association. Corals displayed are considered in situ unless stated otherwise in 1078 1079this caption. Acro-Iso association, 1. Acropora gr. humilis (core 70-1), 2. Acropora gr. formosa 1080(core 46-3), 3. Acropora gr. robusta (cores 9-1, 9-3), 4. massive Isopora (core 63-3), 5. 1081 branching Isopora (core 38-2). Po-Mo-Fa association, 6. Platygyra gr. sinensis, 7-8. 1082unidentified corals, 9. Montipora (core 29-7), 10. Porites (core 20-1), 11. Montipora (core 41-1083 3), 12. Goniastrea edwardsi (core 80-1). Poc association, 13. Seriatopora hystrix (core 55-5), 14-15. Stylophora pistillata (cores 56-19, 76-7). Mesophotic association, 16. Echinophyllia 1084 (core 117-5), 17. Porites (core 117-2), 18-19. Seriatopora hystrix (not in situ), 20. Porites (core 10851086 125-5). The scale bar is 5 cm.

1087

1088 **Table 1**: Comparison of criteria used to define depositional unit boundaries.

1089

1090 **Table 2**: List of coral taxa identified in the Ribbon Reef 5 borehole.

1091

1092 **Table 3**: Results of the similarity percentage analysis (SIMPER).

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Supplementary figure S1: Distribution of in situ (IS, IS?) and non in situ (ISX) coral taxa in the Ribbon Reef 5 borehole identified in this study. Each coral is represented by a horizontal bar (solid fill: IS, IS? no fill: ISX). Corals displayed in the column Po/Mo are those for which the distinction between *Porites* and *Montipora* could not be made and include mainly thin encrusting colonies. Bar charts showing the variations in the thickness of in situ corals and algal crusts in the borehole are also shown.

1100

1101 **Supplementary figure S2**: (A) Group-averaged hierarchical clustering of the BC coefficient 1102 calculated between core sections based on the square root of abundance data for in situ 1103 corals; (B) two-way table showing the taxa indicative of each cluster (coral assemblage).

- 1104
- 1105 Supplementary figure S3: Columns showing the succession of in situ (IS, IS?) coral
- assemblages defined by cluster analyses: (A) analysis of presence-absence data and (B)
- 1107 analysis of the square root of abundance data.









Figure 4

















Table 1.

Comparison of criteria used to define depositional unit boundaries.

		Lithology	Diagenetic cements	Dissolution features	¹ Paleosol	Surface	Fossilized plant	Δ color	Δ Corals	∆ Algae	δ¹ ⁸ Ο	$\delta^{^{13}}C$
A.ª	R2-R1 (16 m)*			vugs	brown sedim.	erosion surf.	cells				negative shift	
	R3-R2 (25 m)	reef fmwk <i>→</i> <i>Halimeda</i>		vugs	brown sedim.	erosion surf.				$M \to L$	negative shift	
	R4-R3 (64 m)							pink→brown		$M \to L$	negative shift	
	R5-R4 (85 m)								$A \mathop{\rightarrow} B$	$M \to L$	negative shift	
	R6-R5 (94 m)									$M \to L$	small negative shift	
	Rd1-R6 (96 m)	rhodo Imst → reef fmwk							abund.↑ <u>●</u> compo. shift			
	Rd2-Rd1 (112 m)								abund.↑ <u>●</u> compo. shift	$L \to Me$		
	R7-Rd2 (117 m)	reef fmwk → rhodo Imst							shift in taxon.comp.	$M {\rightarrow} Me$		
	R8-R7 (130 m)	reef fmwk → grst				irregular surf.				$M \to L$	negative shift	
	Rd3-R8 (138 m)	$ rhodo \ Imst \\ \rightarrow \ coral \ Imst $							in situ coral abund.↑ _			
	Rd4-Rd3 (152 m)								in situ coral abund.↓ <u>●</u>			
	Grst-Rd4 (158 m)	grst→ rhodo Imst										
В. ^ь	U8-U9 (15.85 m)	grst/rdst → grst/pckst	agaronite cements 💻	dissolution ↑	brown sedim.	erosion surf.	rootlets cells				positive shift	positive shift
	U7-U8 (28 m)	grst/wckst → grst/rdst		vugs	brown sedim.	erosion surf.	rootlets cells				_	negative peak
	U6b-U7 (36 m)	rdst → grst/wckst		vugs	brown sedim.	erosion surf.	rootlets					decreasing trend
	U6a-U6b [*] (57 m)	**	complex cements	vugs		erosion surf.						variability↑ ●
	U5-U6a (73 m)			vugs					coral density, diversity ↑ ●			variability↓
	U4-U5 (99 m)	wckst/pckst → grst		dissolution ↑							negative shift	
	U3-U4 (131 m)	grst → mdst/wckst				irregular surf.			coral size, diversity ↑_●_			positive shift
	U2-U3 (155 m)	grst/pckst → grst/rhodo		dissolution ↑							negative shift	negative shift
	U1-U2 (184 m)	folded mdst	arag., fibrous calcite								positive peak	positive peak
					Lit	hologies					Coral	line algae
	Major criter Secondary	ria <u>•</u> criteria •	Above bounda Below bounda	ary ∱lno ıry ↓De	crease rd: ecrease gr:	st Rudston st Grainsto	e wcks ne mdst	t Wackestor Mudstone	e rhodo fmwk	Rhodolith Framework	M L	Mastophoroid Lithophylloid
pckst Packstone Imst Limestone Me Melobesioid												

a Criteria used in Webster (1999) and Webster and Davies (2003).

b Criteria used in Braithwaite et al. (2004) and Braithwaite and Montaggioni (2009).

* Depth of the depositional unit boundary (in mbsf).

** Subdivision of U6 into U6a and U6b proposed in Braithwaite and Montaggioni (2009).

Table 1

Table 2.
List of coral taxa identified in the Ribbon Reef 5 borehole.

SCLERACTINIAN CORALS	10. Stylophora	6. Mussidae	NON-SCLERACTINIAN CORALS			
1. Acroporidae	23. S. pistillata	23. Symphyllia	15. Tubiporidae			
1. Acropora	11. Pocillopora	44. S. sp.	35. Tubipora			
1. A. sp. 1 (BRF ARB)	24. P. verrucosa	7. Euphylliidae?	59. <i>T. musica</i>			
2. A. sp. 2 (BRF COR)	5. Faviidae	24. Euphyllia?	16. Milleporidae			
3. A. sp. 3 (BRF DIG)	12. Caulastrea	45. <i>E.</i> sp.?	36. Millepora			
4. A. sp. 4 (BRF TUB) ^{isx}	25. <i>C. furcata</i> ^{ISX}	8. Oculinidae	60. <i>M.</i> sp.			
5. A. sp. 5 (BRM ARB)	13. <i>Favia</i>	25. Galaxea	17. Alcyonidae			
6. A. gr. hyacinthus (PLA	TY) 26. F. ? sp. 1*	46. G. astreata	37. Sinularia			
7. A. gr. formosa (STAG)	27. F. stelligera	47. G. fascicularis ^{ISX}	61. S. sp.			
8. A. gr. humilis (DIG, CO	PR) 28. F. laxa	9. Siderastreidae				
9. A. gr. robusta?	29. F. gr. pallida	26. Coscinarea				
2. Isopora	14. Favites	48. C. gr. columna				
10. <i>I.</i> sp. 1 (BR)	30. <i>F. pentagona^{isx}</i>	10. Agarciidae				
11. <i>I.</i> sp. 2 (EN-M)	31. F. gr. abdita	27. Pavona				
3. Astreopora	15. Goniastrea	49. <i>P. clavus</i> ^{ISX}				
12. A. sp.	32. G. retiformis	28. Leptoseris				
4. Montipora	33. G. edwardsii	50. <i>L. foliosa?</i> ISX				
13. <i>M</i> . sp. 1 (EN)	16. <i>Platygyra</i>	51. <i>L. gardineri</i> ? ^{ISX}				
14. <i>M</i> . sp. 2 (M)	34. P. pini	29. Pachyseris				
2. Poritidae	35. P. gr. sinensis	52. P. rugosa				
5. Porites	17. Leptoria	53. P. speciosa				
15. P. sp. 1 (BR)	36. L. phrygia	11. Fungiidae				
16. P. sp. 2 (KN)	18. Oulophyllia	30. <i>Fungia</i>				
17. P. sp. 3 (EN)	37. O. sp.	54. <i>F</i> . sp.				
18. <i>P</i> . sp. 4 (M)	19. Montastrea	31. Cycloseris				
6. Goniopora	38. <i>M. curta</i>	55. C. sp.				
19. G . sp. ^{ISX}	20. Leptastrea	12. Pectiniidae				
7. Alveopora	39. L. pruinosa	32. Echinophyllia				
20. A . sp. ^{ISX}	21. Echinopora	56. <i>E. echinata</i> ? ^{ISX}				
3. Astrocoeniidae?	40. <i>E. lamellosa</i>	13. Merulinidae				
8. Stylocoeniella?	41. E. mammiformis	33. Hydnophora				
21. S. sp.	42. <i>E. gemmacea</i> ^{isx}	57. <i>H</i> . sp.				
4. Pocilloporidae	22. Cyphastrea	14. Dendrophylliidae				
9. Seriatopora	43. C. serailia	34. Turbinaria				
22. S. hystrix		58. <i>T</i> . sp.				

ISX The superscript ISX indicates taxa recorded only in the non-in situ fraction.

Table 2

Table 3.	
Results of the similarity percenta	ige analysis (SIMPER).

A. Comparison of	reef cycles				B. Comparison of coral associations ^b						
	Av.Abund ^c	Av.Sim	Sim/SD ^d	Contrib%	Cum.% ^e		Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Cluster 1						Po-Mo-Fa assoc.					
Acropora BR	1.00	11.60	11.63	18.73	18.73	Po/Mo	1.00	12.95	3.19	34.39	34.39
Porites EN	1.00	11.60	11.63	18.73	37.47	Montipora EN	0.83	7.05	1.34	18.71	53.10
Porites M	1.00	11.60	11.63	18.73	56.20	Porites M	0.83	7.05	1.34	18.71	71.82
Stylophora	1.00	11.60	11.63	18.73	74.93	Porites EN	0.67	4.05	0.78	10.75	82.57
Acropora BM	0.67	4.17	0.58	6.73	81.66						
						Poc assoc.					
Cluster 2						Stylophora	0.75	15.71	0.61	67.73	67.73
Acropora BM	1.00	4.67	16.48	7.02	7.02	Acropora BM	0.50	2.22	0.41	9.58	77.31
Isopora M	1.00	4.67	16.48	7.02	14.03	Montipora M	0.50	1.75	0.41	7.56	84.88
Montipora EN	1.00	4.67	16.48	7.02	21.05						
, Montipora M	1.00	4.67	16.48	7.02	28.07	Acro-Iso assoc.					
Porites EN	1.00	4.67	16.48	7.02	35.09	Porites M	1.00	9.41	3.46	18.71	18.71
Porites M	1.00	4.67	16.48	7.02	42.10	Porites EN	0.88	6.98	1.45	13.88	32.59
Po/Mo	1.00	4.67	16.48	7.02	49.12	Montipora EN	0.88	6.63	1.49	13.19	45.78
Stylophora	1.00	4.67	16.48	7.02	56.14	Acropora BM	0.75	4.54	0.97	9.03	54.81
Favia	1.00	4.67	16.48	7.02	63.15	Acropora BR	0.63	3.55	0.68	7.07	61.88
Pectiniidae	1.00	4.67	16.48	7.02	70.17	Acropora BF	0.63	3.38	0.69	6.72	68.60
Acropora BF	0.75	2.39	0.91	3.59	73.76	Stylophora	0.63	2.91	0.69	5.79	74.38
Pocillopora	0.75	2.35	0.91	3.54	77.30	Montipora M	0.63	2.78	0.70	5.53	79.91
Leptoria	0.75	2.21	0.91	3.32	80.61	Isopora M	0.63	2.72	0.70	5.41	85.32

a Results of the similarity percentage analysis for the two clusters identified in figure 6.

b Results of the similarity percentage analysis for the three coral associations (Po-Mo-Fa, Poc and Acro-Iso).

c Frequency of occurrence (1.00 meaning the coral occurs in 100% of the samples).

d Ratio of the contribution of a taxon to the average similarity of a group (SIM) to the standard deviation of their contribution (SD). Taxa with the highest Sim/SD are present most consistently among the samples of a group.

e The listed corals are those which account for 80-85% of the within-group similarity.

Table 3



Supplementary figure S1



Supplementary figure S2



Supplementary figure S3